

M1 D²HP Development of Drugs and Health Products



OTU 01 Diagnosis of infectious diseases Introduction class

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Content

I- Samples

II- Virological diagnosis

- 1- Direct diagnosis: antigen detection**
- 2- Direct diagnosis: genome detection**
- 3- Direct diagnosis: virus isolation**
- 4- Indirect diagnosis**

III- Context of diagnosis

IV- Presentation of the OTU 01

Conventional microbiological diagnosis

↳ Conventional microbiology techniques

Direct examination after specific staining (Gram)

Detection of specific antigen

Bacterial and fungal culture

Antibiogram on isolated colonies

Cell culture for viruses

Serodiagnosis

↳ Issues

Low sensitivity

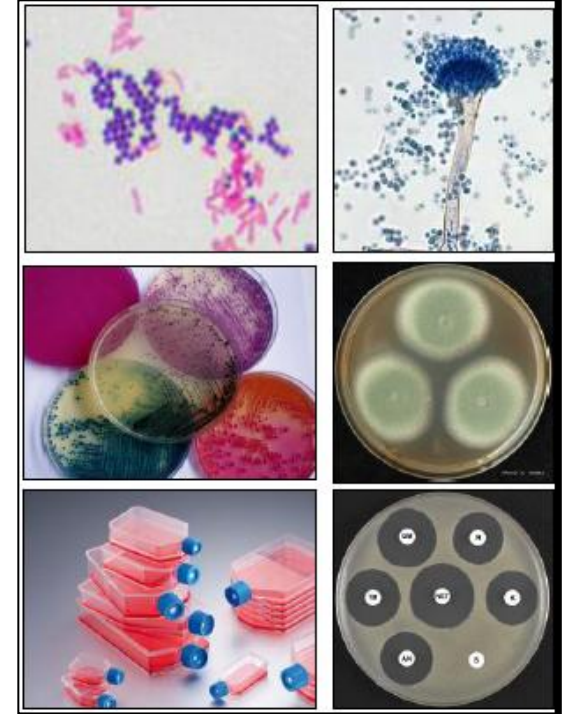
Multiplicity of possible etiologies for the same infectious syndrome

Culture default (patients already under treatment, infectious agents difficult to isolate or not cultivable)

Important delays in obtaining partial and then complete results

↳ incompatible with early adaptation of treatments

⇒ **Emergence of molecular biology techniques +++**



Biological signs and biological diagnosis of infection

☞ **Non-specific diagnosis**

Biochemical analysis

Hematology analysis

Cytopathological analysis, etc.

☞ **Specific diagnosis**

Identification of the infectious agent

Identification of specific antibodies

I- Samples

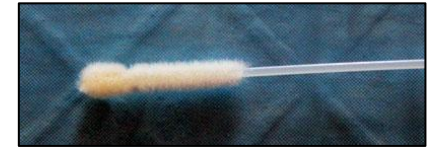
Principle of sampling

- ☞ **Taking samples based on the syndromes and what we are looking for (suspected virus)**
- ☞ **At the site of virus multiplication**
 - ☞ target organ, virus entrance door
 - ☞ if accessible
- ☞ **The earliest, at the beginning of clinical signs**
 - = during the acute phase of the disease
 - ☞ when viral excretion is the highest

Types of samples

- ☞ **Blood, serum**
- ☞ **Stool**
- ☞ **Urine**
- ☞ **Nasal swab, nasopharyngeal aspiration, bronchoalveolar lavage**
- ☞ **Pap smear**
- ☞ **Biopsy**
- ☞ **Cerebrospinal fluid**
- ☞ **Amniotic fluid**

Collection and transportation of the sample



- ☞ For isolation, the viability of the viruses present must be preserved
- ☞ Suitable transport media
- ☞ Protected from drying, heat and pH variations

II- Virological diagnosis

Techniques for virological diagnosis

Direct diagnosis	Indirect diagnosis
Antigen detection	Serology
Immunofluorescence, immunoperoxidase	ELISA
Seroneutralisation	Western-Blot
IHA	Immunochematography
ELISA	IgG avidity assay
Immunochematography	
Genome detection	
PCR (RT-PCR, real time PCR)	
RNA amplification (NASBA, TMA)	
Branched DNA assay	
Hybridization (<i>in situ</i> , liquid phase)	
Virus isolation	
Virus isolation in cell culture (CPE, haemadsorption)	
Electron microscopy	

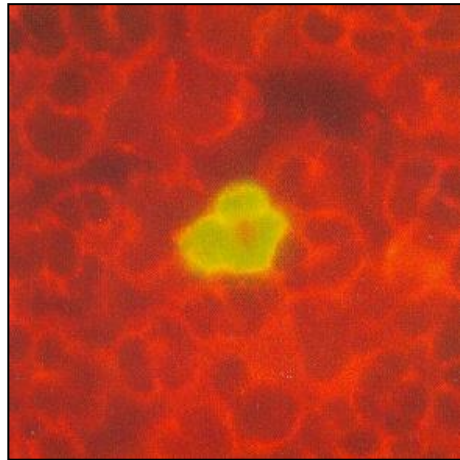
II- Virological diagnosis

1- Direct diagnosis: antigen detection

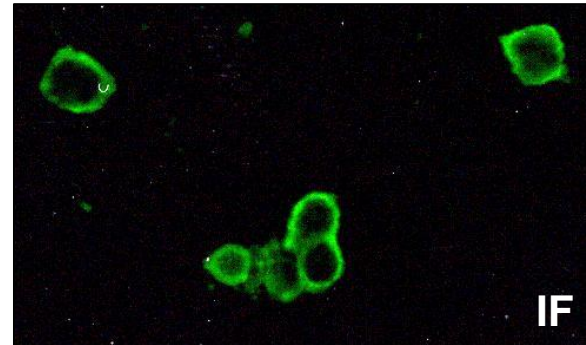
Techniques for antigen detection (1)

☞ Immunofluorescence

Antigenemia pp65 (CMV)



HSV-1 and HSV-2

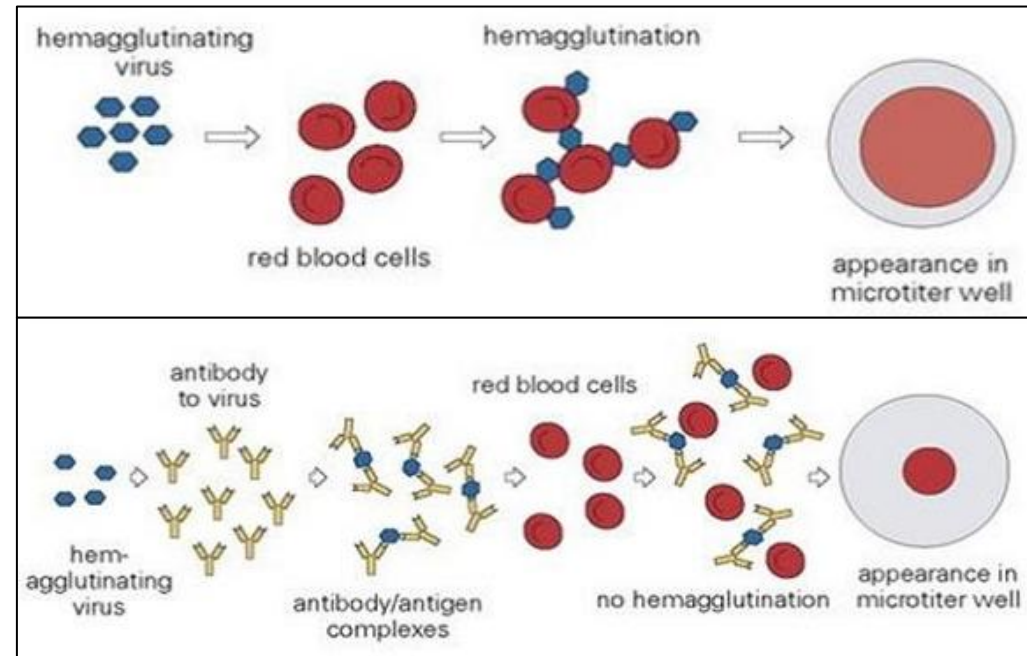


Techniques for antigen detection (2)

↳ Inhibition of hemagglutination (IHA)

↳ antiserum

Rubella virus

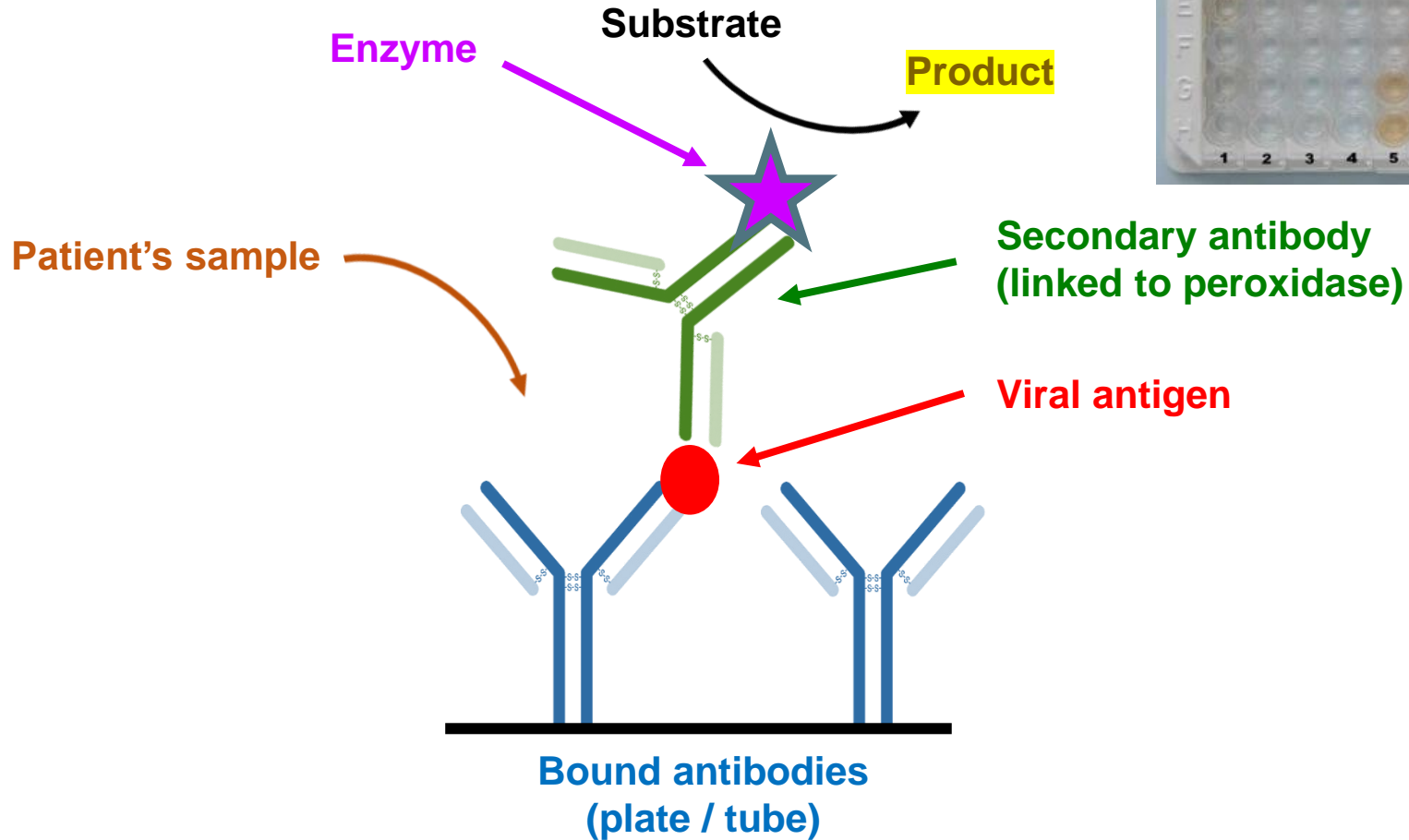
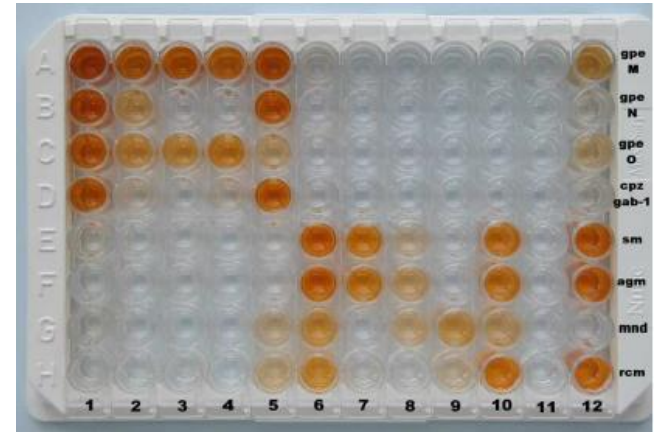


↳ Advantages

- Rapid and easy to use
- Applicable to viruses that are not cultivable or difficult to cultivate

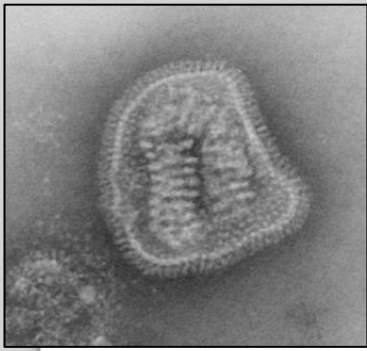
ELISA (Enzyme-linked immunosorbent assay)

Direct ELISA (sandwich)



⇒ Very sensitive method (nM or pM)

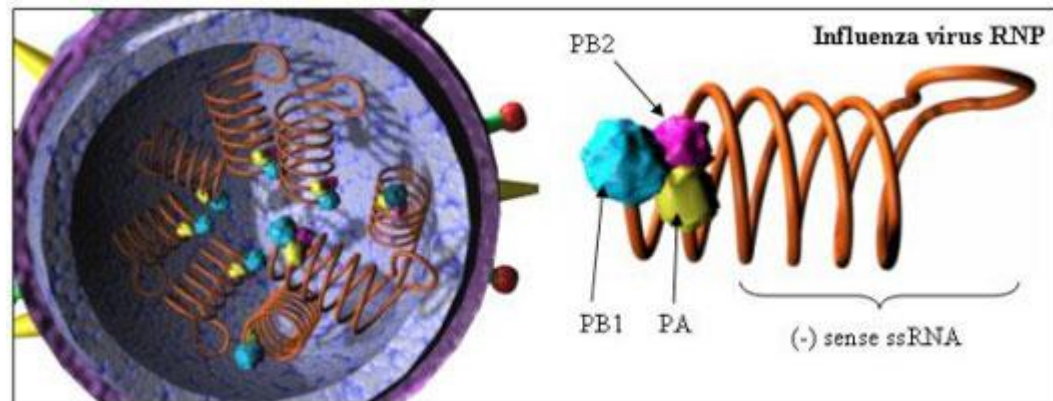
Immunochemistry (1)



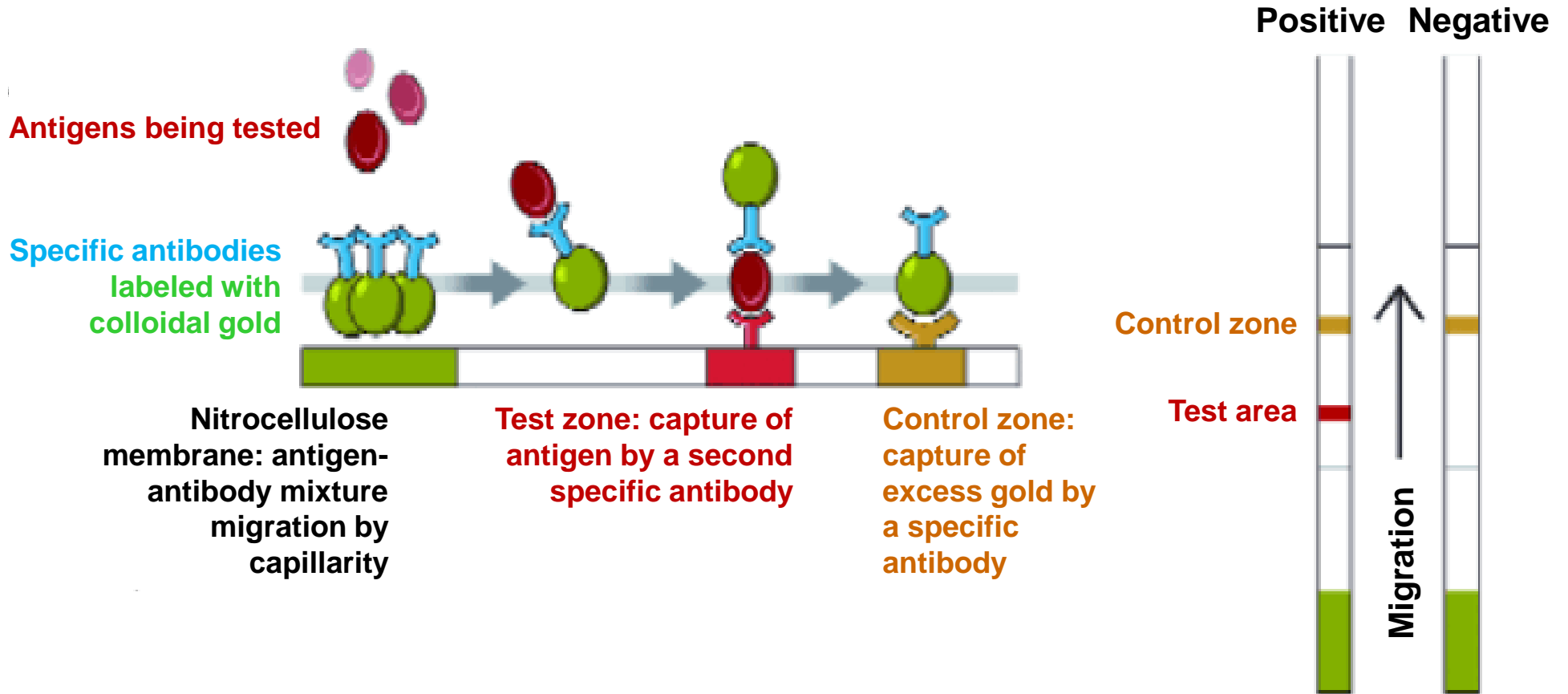
☞ Rapid diagnostic test (RDT)

⇒ Immunochemistry for influenza virus

- Rapid and easy to use
- Results in 15-30 minutes
- Visual reading
- Detection of the nucleoproteins of the influenza viruses A and B
- Detection of all subtypes of influenza A virus
 - ☞ H1N1, H5N1



Immunochromatography (2)



Rapid diagnosis

👉 Indications

⇒ Specific and timely patient management in certain circumstances

- Respiratory infections: influenza virus, SRV (Syncytial Respiratory virus), SARS-CoV-2, etc.
- Infant diarrhea: adenovirus, rotavirus

👉 Advantages

⇒ Speed of rendering result

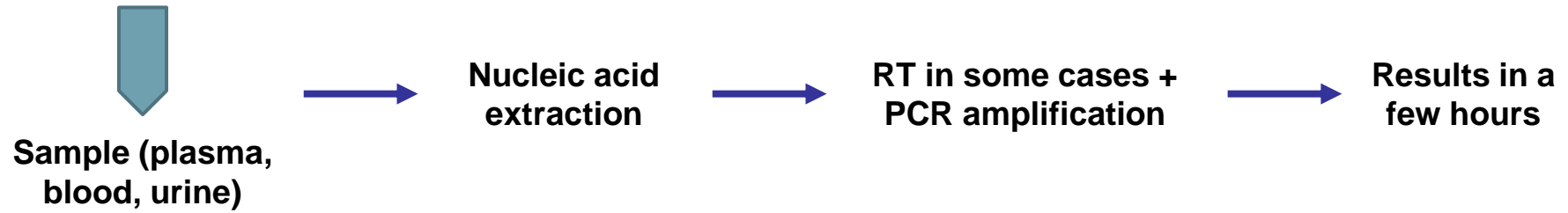
👉 Limitations

⇒ Lack of sensitivity



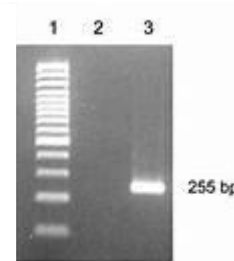
II- Virological diagnosis
2- Direct diagnosis: genome detection

PCR / RT-PCR

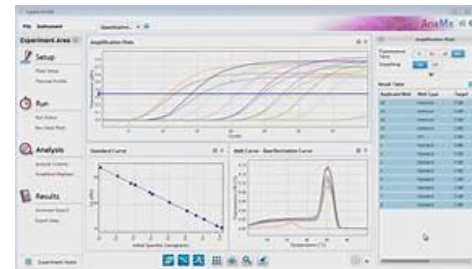


2 methods of detection

Agarose gel



Real-time PCR



⇒ **Very sensitive method**

⇒ **But beware of false negatives**

↳ **highly variable viral genomes**

Reverse transcription (RT)

↳ Reverse transcriptase synthesizes single strand DNA

↳ using an **RNA** as a **matrix**

Retrovirus



1- Reverse transcriptase



RNA



2- The reverse transcriptase synthesizes a single strand DNA molecule = **cDNA**



3- The single strand DNA molecule is used in turn as a matrix



4- Synthesis of double-stranded DNA

⇒ The complementary DNA (cDNA) is then amplifiable by PCR

Gene amplification by PCR technique (1)

☞ **PCR = Polymerase Chain reaction**

☞ production of a large number of copies of a specific DNA sequence

N cycles of PCR

2^N copies of initial DNA

General case: N = 30 cycles

☞ Synthesis in direction 5' to 3'

☞ Reaction mix

Taq polymerase

DNA matrix

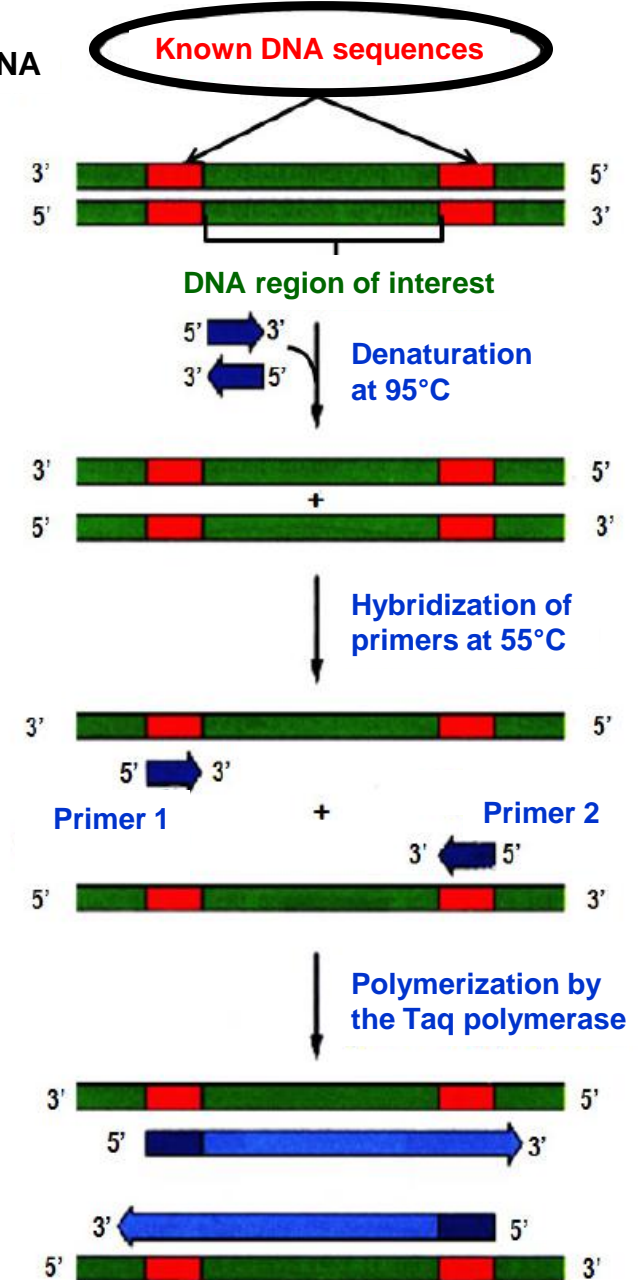
2 primers forward and reverse

Buffer

dNTPs

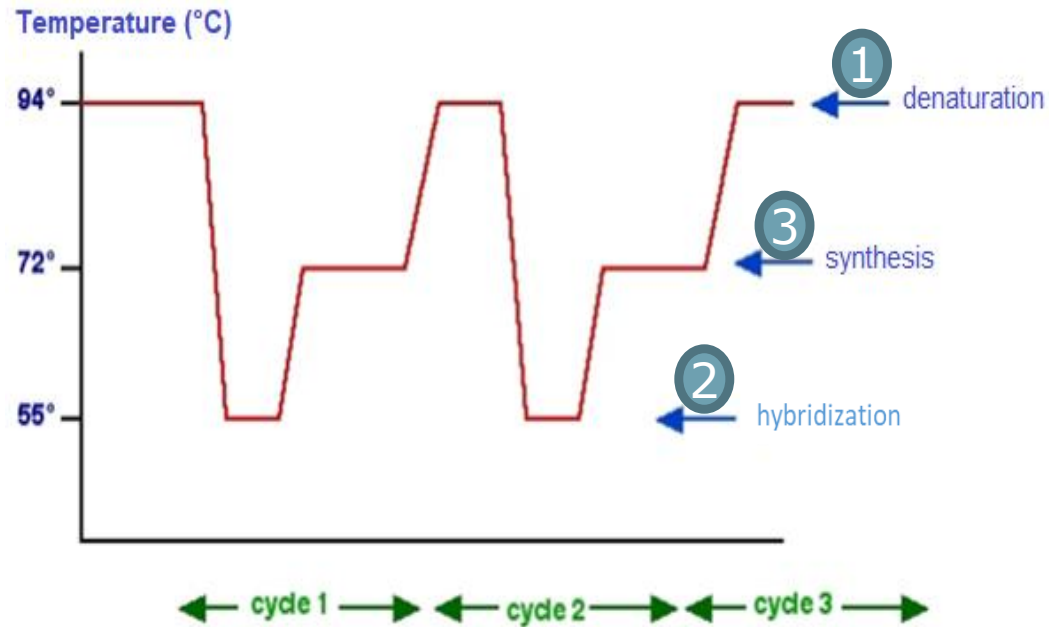
MgCl₂

Chromosomal DNA



Gene amplification by PCR technique (2)

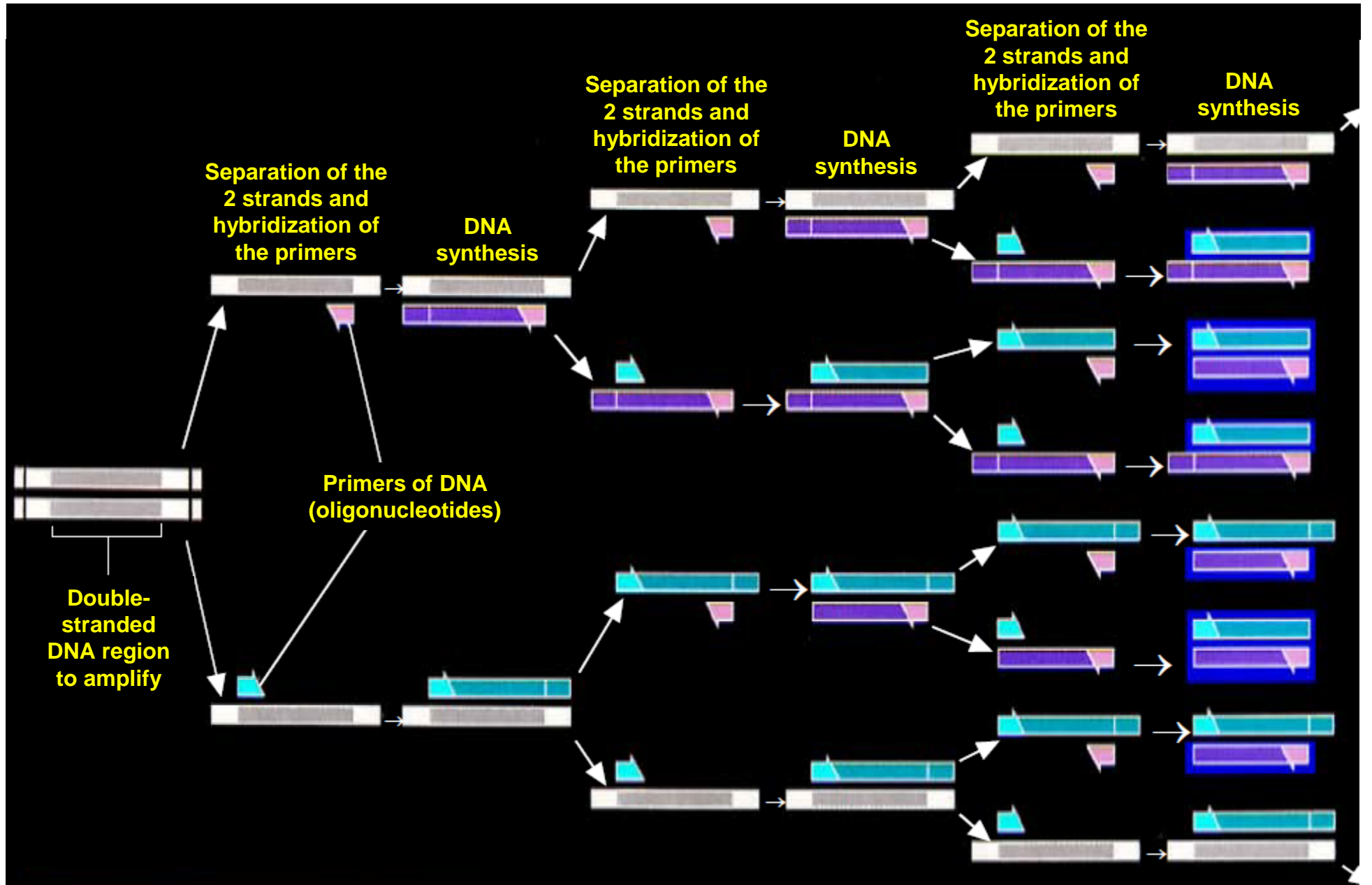
☞ N cycles of PCR



Thermocycler



Gene amplification by PCR technique (3)



Real-time PCR

👉 Principle

↳ real-time measurement of quantities of PCR products synthesized during amplification

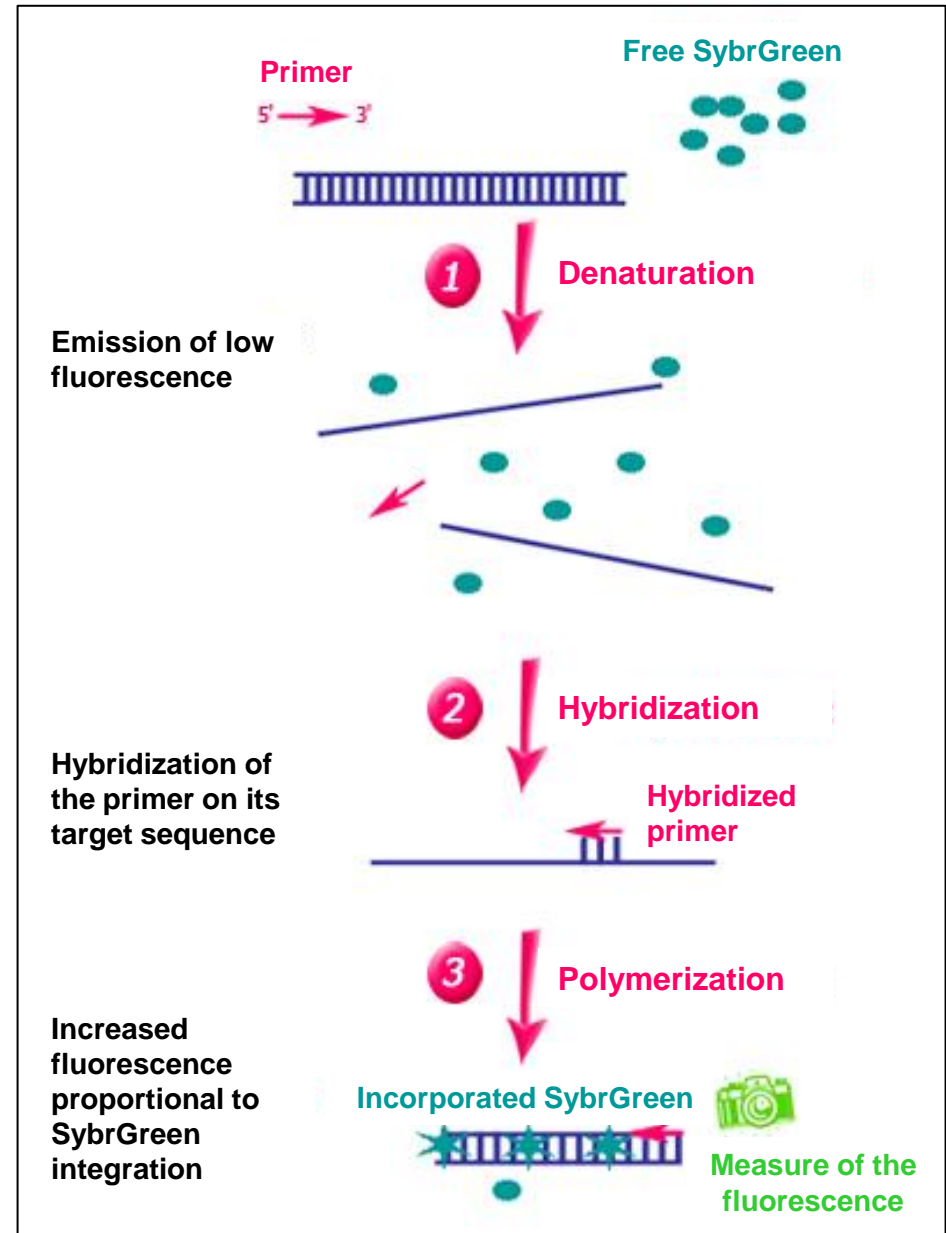
👉 Requirement

- emission of a signal proportional to the amount of amplicon produced
- signal measurement tool
- computer tools for analysis

Real-time PCR and SybrGreen

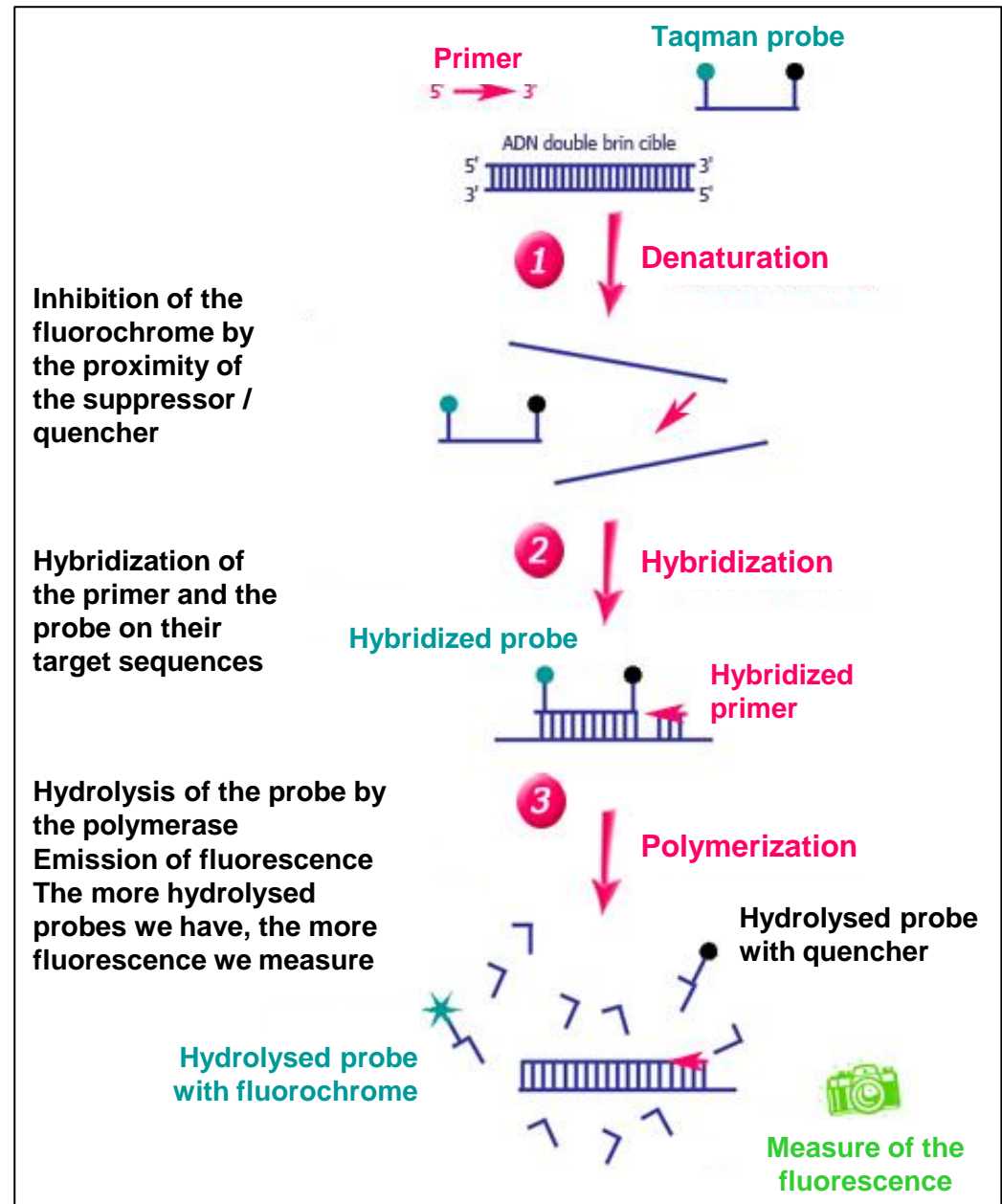
👉 SybrGreen

- ⇒ double-stranded DNA binding agent
- = intercalant of DNA



Real-time PCR and Taqman probe

👉 Taqman hydrolysis probe



Inhibition of the fluorochrome by the proximity of the suppressor / quencher

Hybridization of the primer and the probe on their target sequences

Hydrolysis of the probe by the polymerase
Emission of fluorescence
The more hydrolysed probes we have, the more fluorescence we measure

Hydrolysed probe with fluorochrome

Hydrolysed probe with quencher

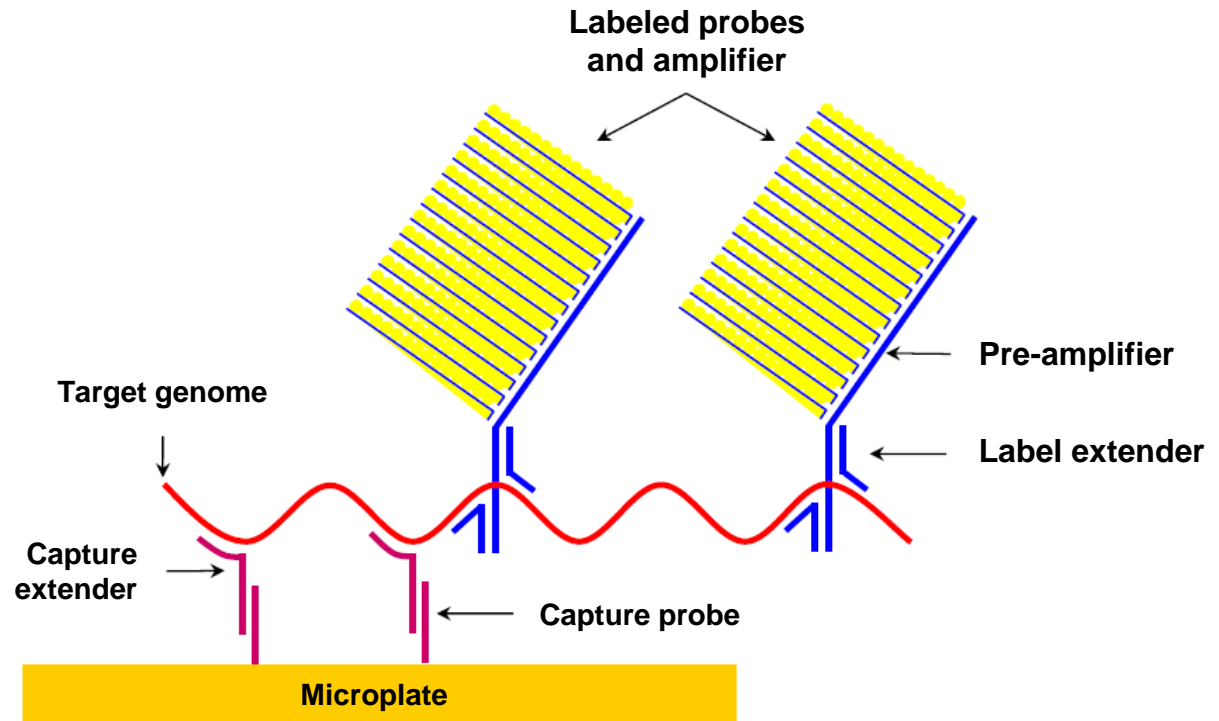
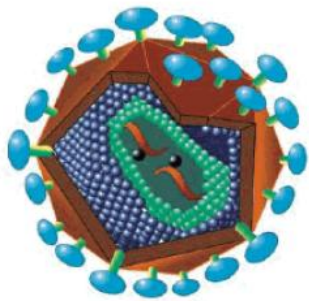
Measure of the fluorescence

Branched DNA Assay

☞ Numerous complementary probes to different regions

↳ detection of highly variable viruses

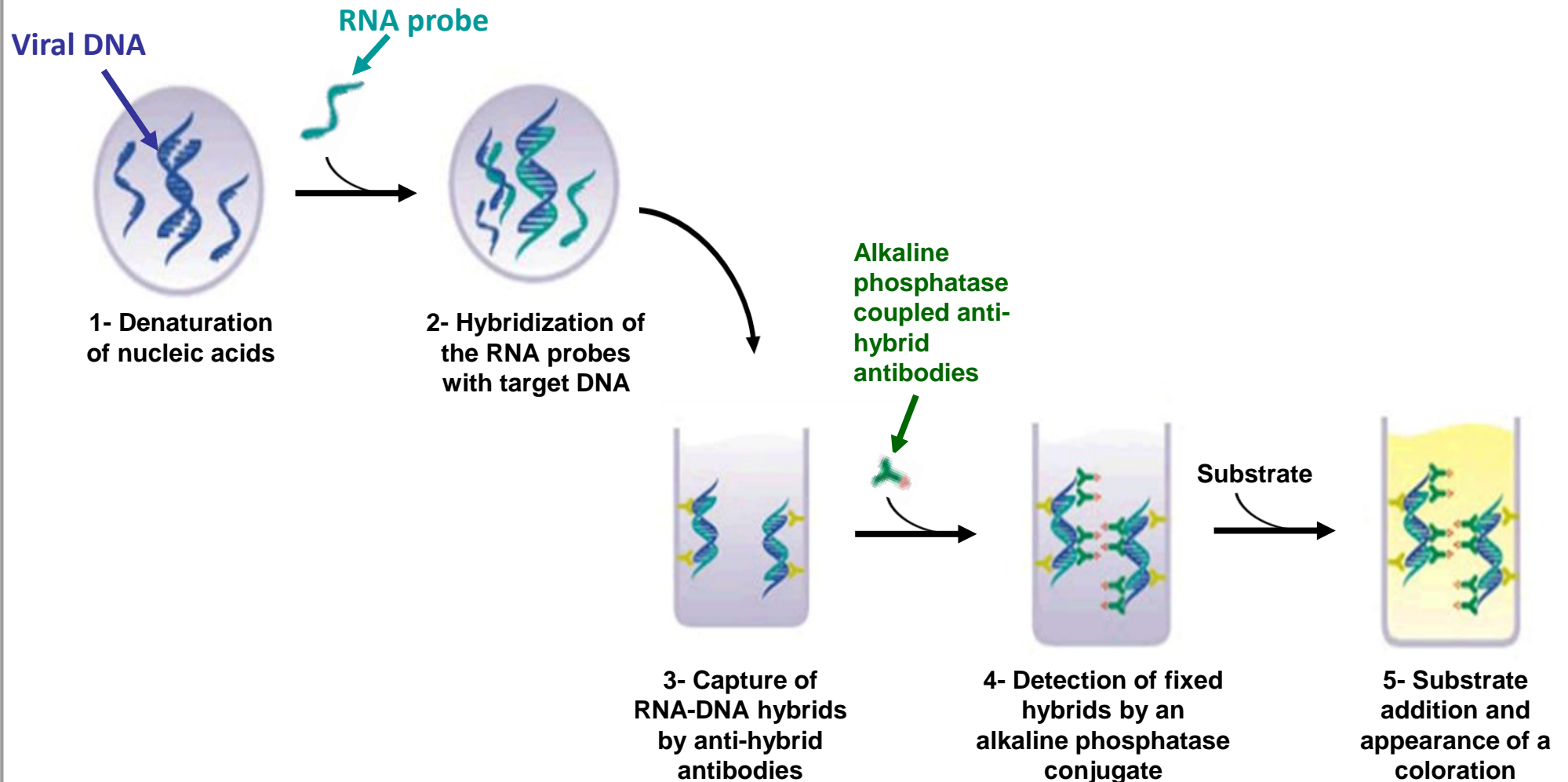
HIV RNA, HCV RNA, HBV DNA



Liquid phase hybridization

☞ Capture of hybrids on solid phase with anti-hybrid antibodies

Digene HPV test ⇒ detection of 13 high-risk HPV types



Molecular diagnosis (1)

☞ Qualitative techniques

⇒ Show the presence of a virus

☞ Quantitative techniques

⇒ Follow-up of patients treated with antivirals (HIV, HBV)

☞ Sequencing

⇒ Characterization of viral genotypes (HCV)

⇒ Detection of nucleotide mutations associated with antiviral resistance (HCV, HIV)

⇒ Characterization of viral tropism (HIV)

⇒ Phylogenetic analysis to trace the origin of an infection

Molecular diagnosis (2)

☞ Advantages

- ⇒ High sensitivity of available tests
- ⇒ Automation +++
- ⇒ Result rendering time shortened (emergencies)

☞ Limitations

- ⇒ No evidence of the infectivity of the virus
- ⇒ Equipment and experienced personnel
- ⇒ Relatively high cost

II- Virological diagnosis
3- Direct diagnosis: virus isolation

Virus culture

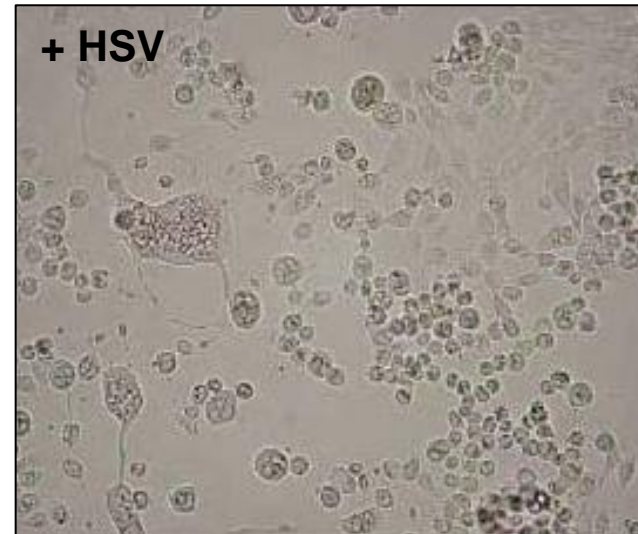
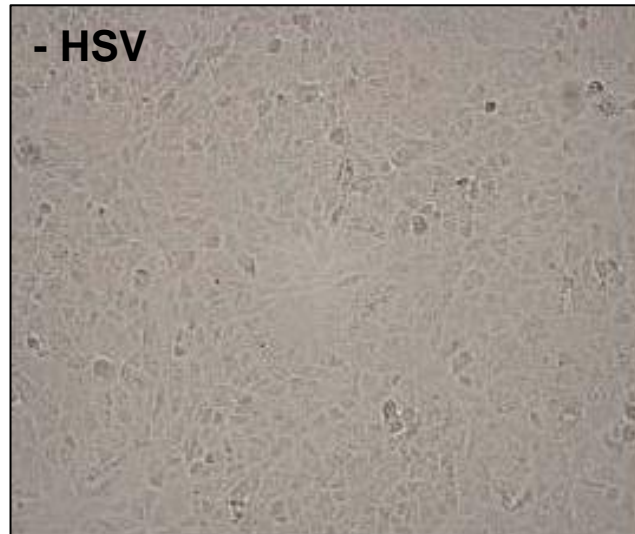
- ☞ **Infection of cells in culture**
- ☞ **Long-standing reference technique**
- ☞ **Use of permissive cells**
- ☞ **Cytopathic effect (CPE)**
- ☞ **Haemadsorption**

Cytopathic effect (1)

☞ Changes in cell morphology

- rounding
- size increase
- support detachment
- lysis of infected cells

HSV



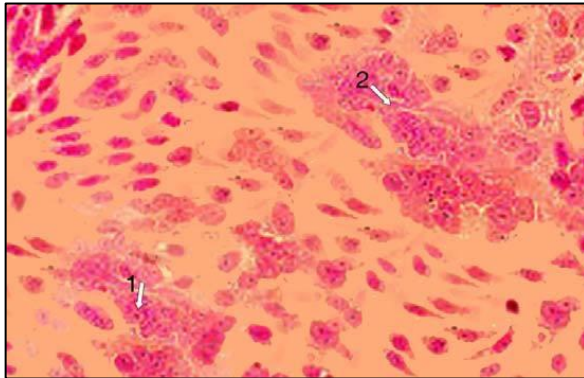
Cytopathic effect (2)

↳ Changes in cell architecture

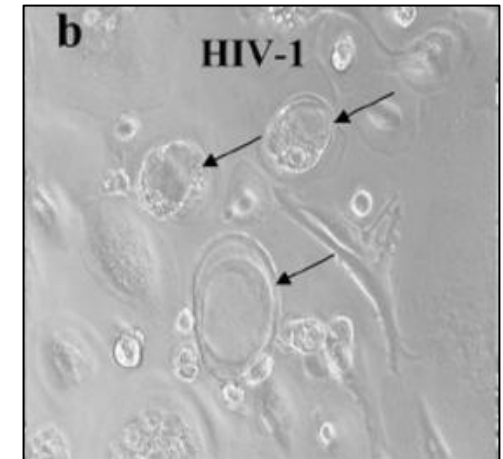
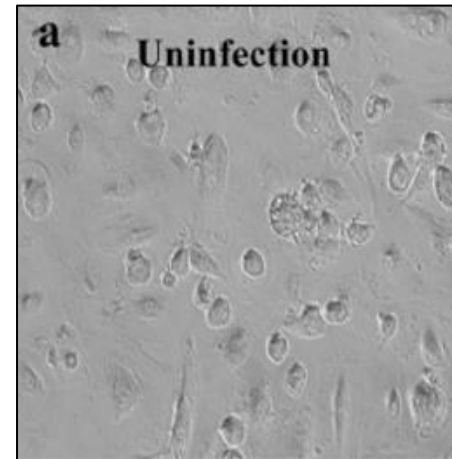
↳ syncytium formation

↳ giant cell formed by membrane fusion of several cells

RSV infection on
Hep-2 cell culture



HIV-1



Cytopathic effect (3)

☞ Formation of inclusions

↳ visible after accumulation of viral material

☞ Cytoplasm

Poliovirus

Rabies virus: Negri bodies

☞ Nucleus

Herpesvirus

Adenovirus



Negri body

Virus isolation

👉 Indications

- ⇒ Proof of infectivity
- ⇒ Characterization of the phenotype of a viral strain for antiviral resistance (HIV) or tropism (HIV)
- ⇒ Epidemiological studies
- ⇒ Vaccine production (flu)

👉 Advantages

- ⇒ Very good sensitivity = **reference method**
- ⇒ Essential for the availability of strains used in vaccine development (flu vaccine)
- ⇒ Reasonable cost

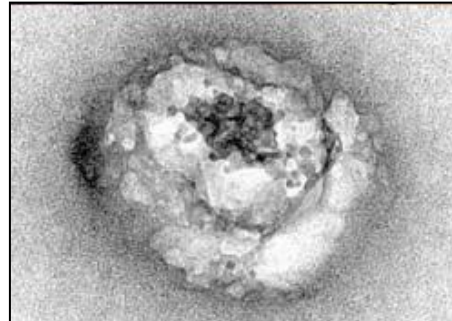
👉 Limitations

- ⇒ Time to result: 48 h to 10 days
- ⇒ Possible subjectivity of the reading
- ⇒ Non-cultivable virus problems: HPV

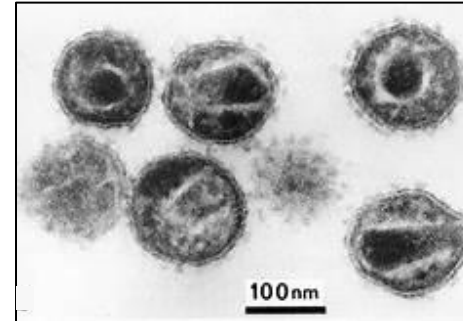
Electron microscopy

☞ Morphological study of virus

HCV



Retrovirus



☞ Study of the virus inside cells

☞ Very high resolution: 1 nm

☞ Interest of electron microscopy mainly in research

II- Virological diagnosis
4- Indirect diagnosis

Indirect diagnosis

- ☞ **Antibody screening**

 - ↳ serological tests

- ☞ **Seropositivity**

- ☞ **Seroconversion**

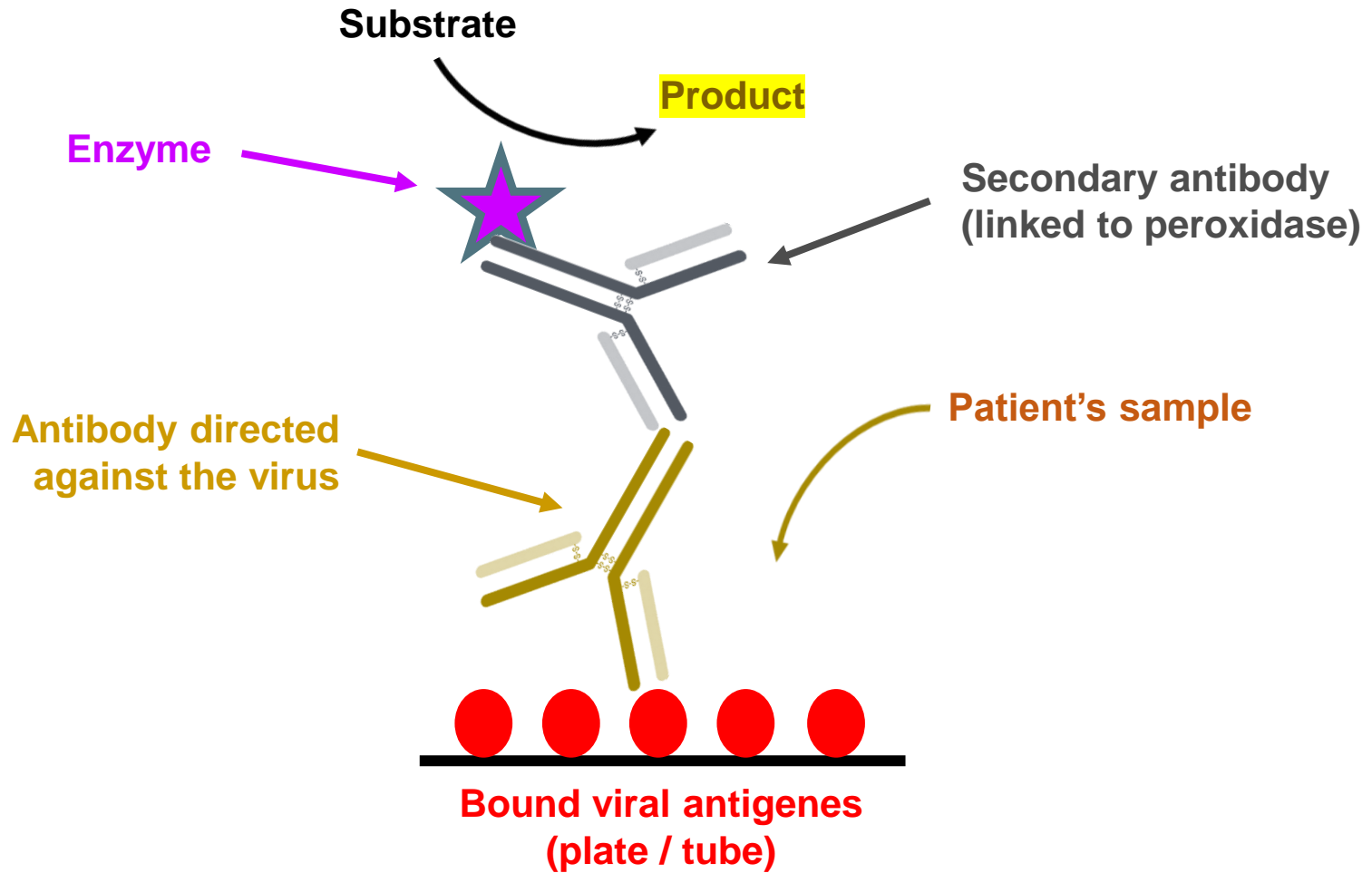
- ☞ **IgM and IgG detection**

- ☞ **IgG avidity assay**

 - CMV, Rubella virus

ELISA (Enzyme-linked immunosorbent assay)

Indirect ELISA



Rapid Diagnostic Test (RDT) (1)



☞ **Unit test**

☞ **Visual reading**

☞ **Rapid**

☞ **Detection of anti-HIV-1 and anti-HIV-2 antibodies**

2 tests detect p24 antigen and anti-HIV-1 and anti-HIV-2 antibodies

Rapid Diagnostic Test (RDT) (2)

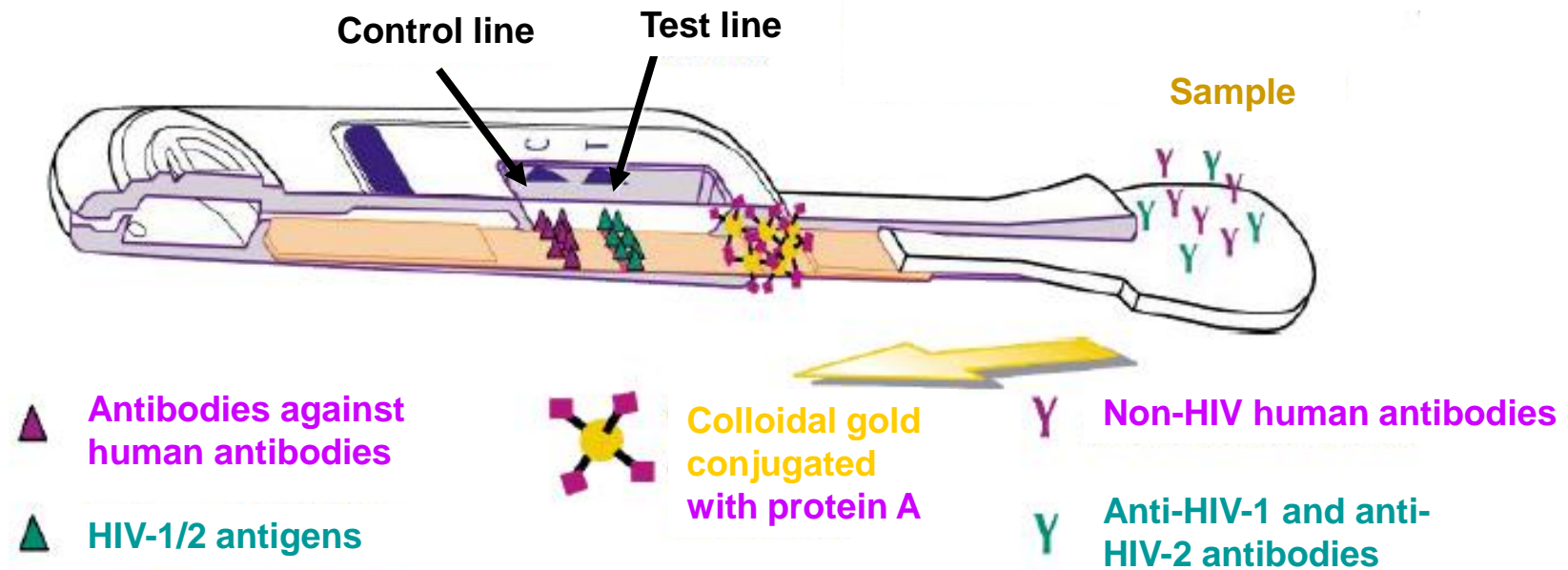
Negative Positive

Positive control band →
HIV band →
Deposit area →

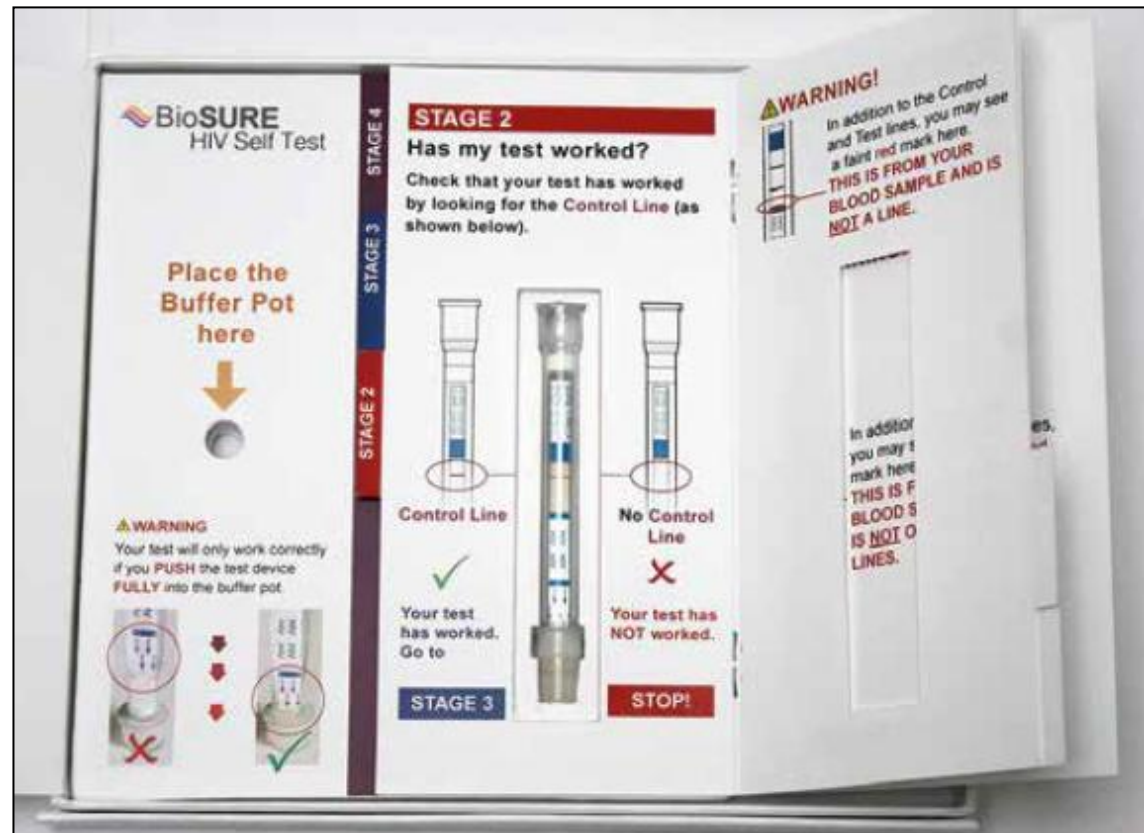


Direction of migration

Principle of immunochromatography for HIV

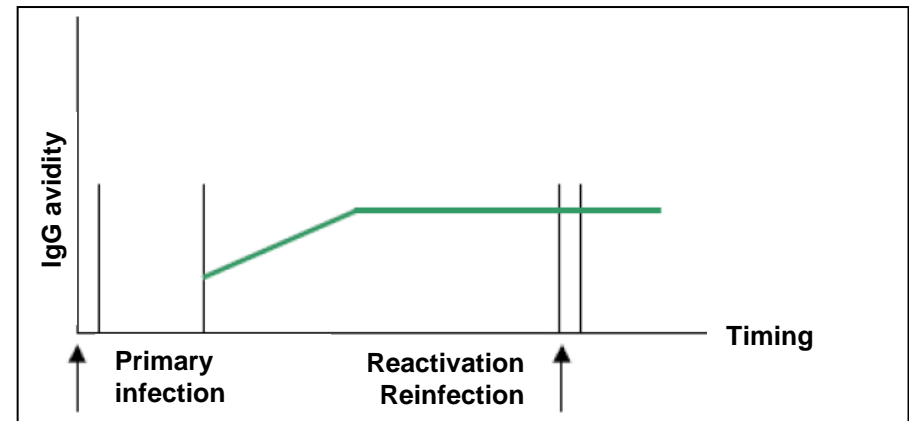
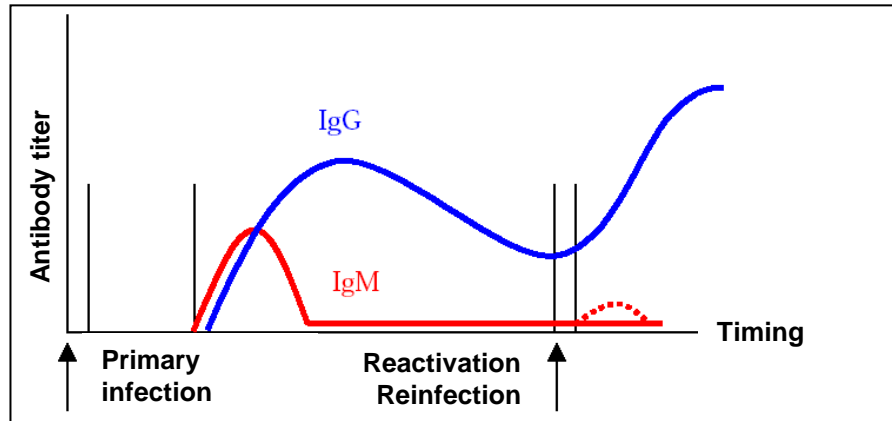


HIV self-tests



IgG avidity assay

👉 For CMV



Serology (1)

☞ Indications

⇒ Evidence of more or less recent contact with a virus

- Seroconversion of specific antibodies

 - ☞ requires 2 samples at 15 days interval

- Presence of specific IgM

- Significant increase in specific IgG

- Measurement of antibody avidity

Serology (2)

☞ Advantages

- ⇒ **Good sensitivity and specificity**
- ⇒ **Automation**
- ⇒ **Time to result**

☞ Limitations

- ⇒ **Reduced sensitivity in certain categories of patients**
 - ☞ **infants and immunocompromised**
- ⇒ **Sometimes difficult interpretation**
 - ☞ **cross-reactivity within the same family of viruses, presence of IgM during reactivation (CMV)**

III- Context of the diagnosis

How and when to use these diagnostic methods?

- ☞ **Characterization of a virus**
- ☞ **Confirmation of an acute infection**
- ☞ **Evidence of the viral origin of an infection**
- ☞ **Search for immunity through ancient infection or vaccination**
- ☞ **Detection of a persistent infection with or without symptoms**
- ☞ **Monitoring of the progress of a viral infection with or without treatment**
- ☞ **Monitoring of the effectiveness of treatment (viral load)**
- ☞ **Blood, organ and tissue donations**
- ☞ **Epidemiological studies**

Viral load

- ☞ **(RT)-PCR measurement / molecular hybridization**
 - ↳ **by quantitative methods**
- ☞ **Viral load**
 - = virus concentration in the blood**
- ☞ **Follow-up of infection**
- ☞ **Monitoring of treatment effectiveness**
- ☞ **Introduction of treatment**
 - = preemptive treatment**

IV- Presentation of the OTU 01

Content of practicals

- ☞ **Theory about diagnostic methods**
- ☞ **Virus isolation: HSV**
 - Preparation of the cells
 - Inoculation of the different samples
 - Observation of CPE 1 week later
- ☞ **ELISA: HBV diagnosis**
 - Ag HBs, anti-HBs and anti-HBc
- ☞ **Real-time PCR: CMV**
 - Taqman technology
- ☞ **Diagnosis of toxoplasmosis**
- ☞ **Tutorials about bacteria diagnosis and clinical cases**

Infectious risk management

- ☞ Potentially pathogenic samples
- ☞ Handling with gloves and a lab coat



+



= bad idea



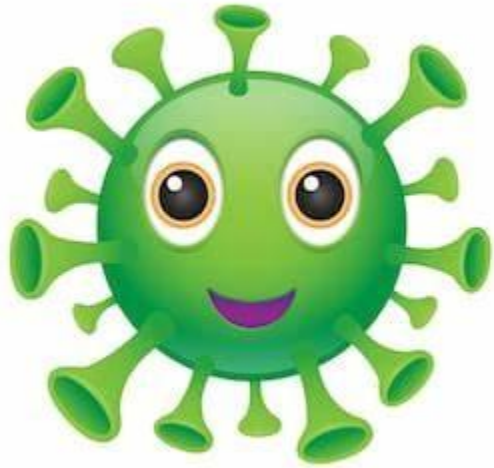
- ☞ Waste management

☞ yellow bin for biological waste

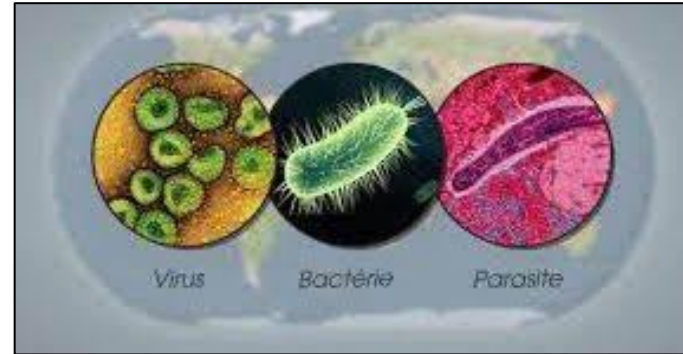


Biological safety hoods





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