

# CRISPR-Cas system

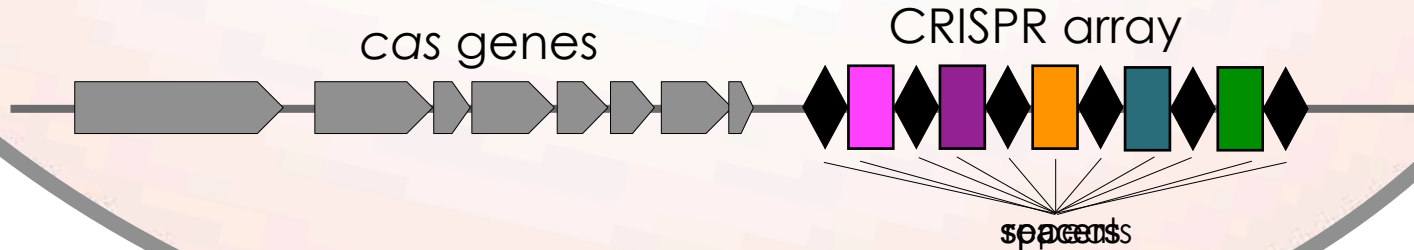
CRISPR (clustered regularly interspaced short palindromic repeats)

Cas (CRISPR-associated) proteins

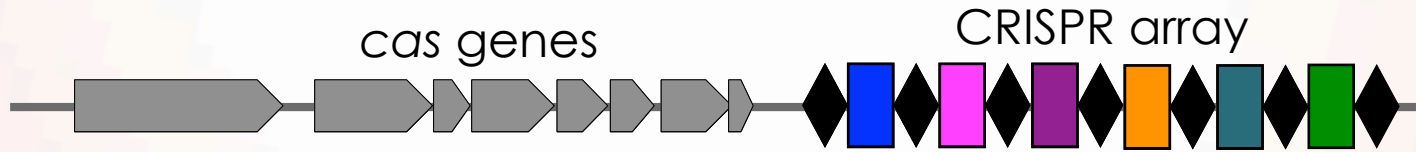
CRISPR-Cas system provide prokaryotes with adaptive immunity

- immunization – cells collect molecular records of encounters with genetic invaders;
- self versus non-self discrimination – cells must avoid autoimmunity but specifically target and destroy foreign genome

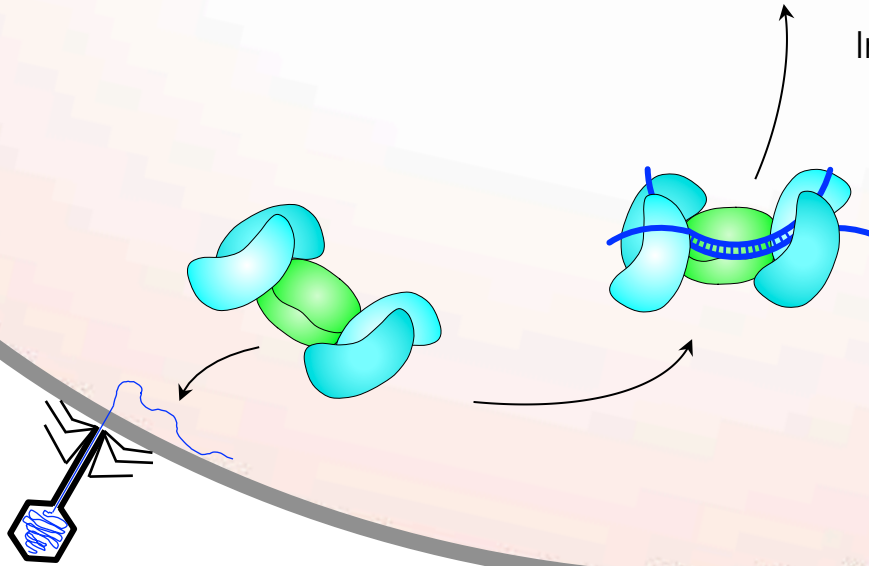
## CRISPR-Cas locus structure



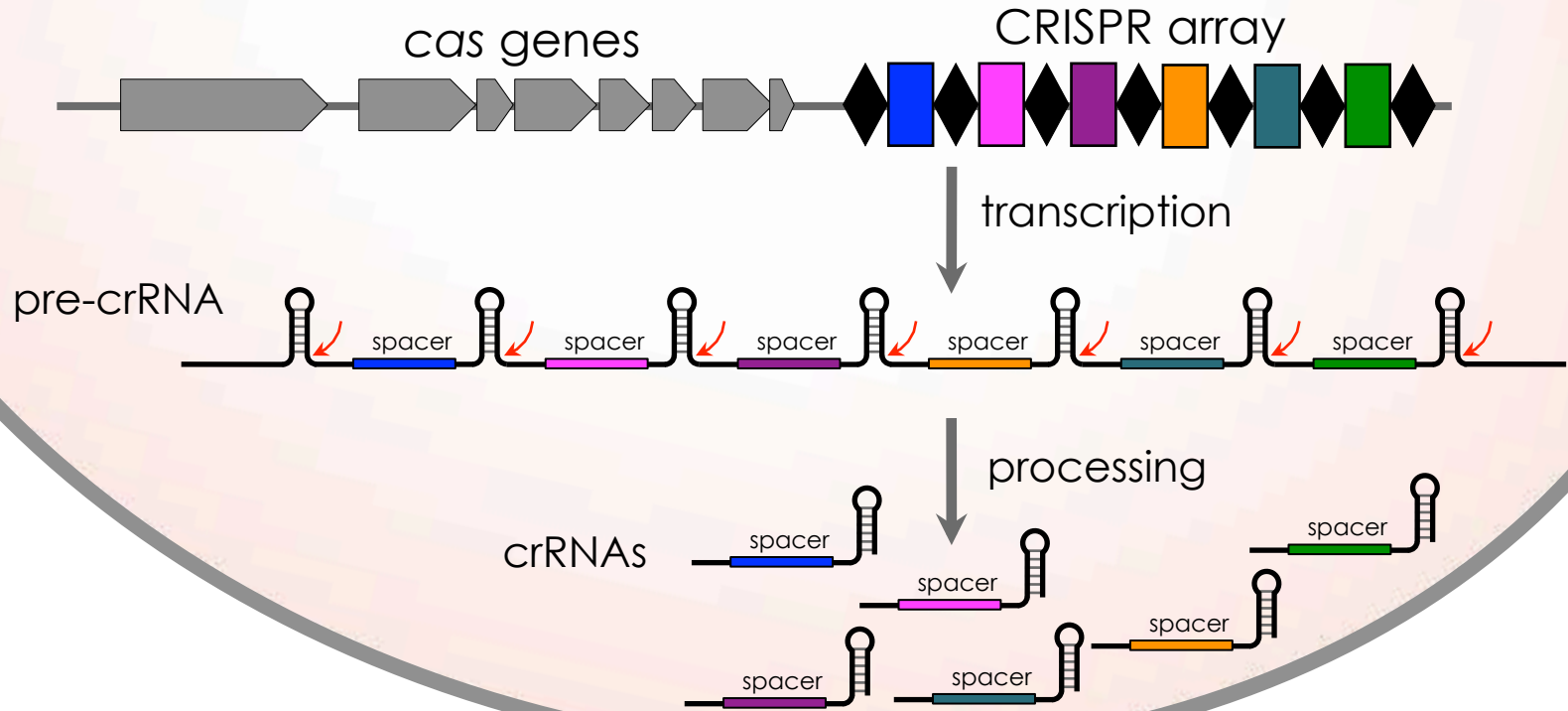
# CRISPR adaptation



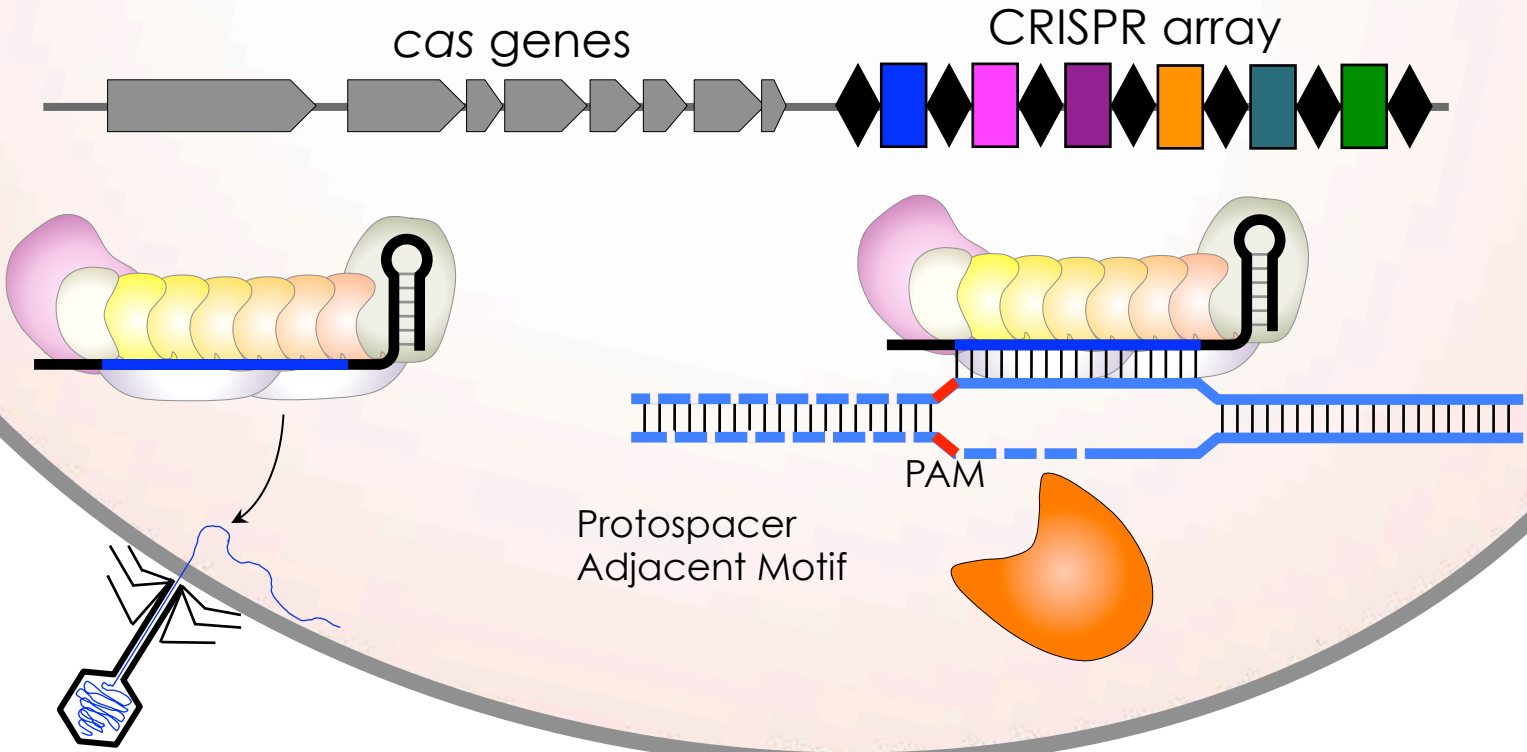
Integration of new  
spacers  
into CRISPR array



# CRISPR RNA biogenesis



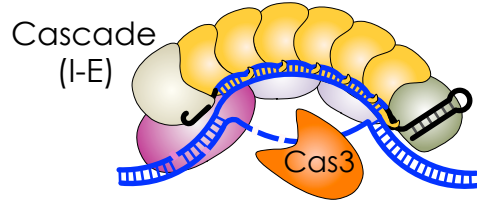
# CRISPR interference



# Classification of CRISPR-Cas systems

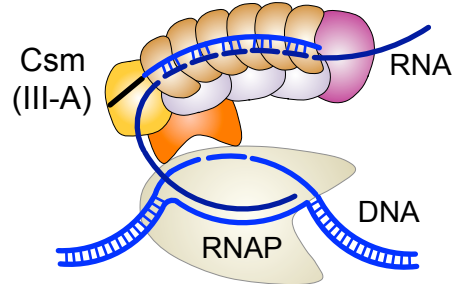
## Class 1 (multi-subunit effector complex)

Type I



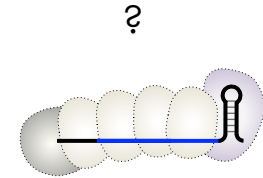
Target: dsDNA

Type III



Target: ssRNA/ssDNA

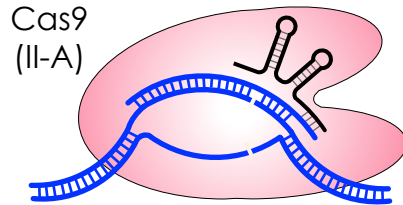
Type IV



Target unknown

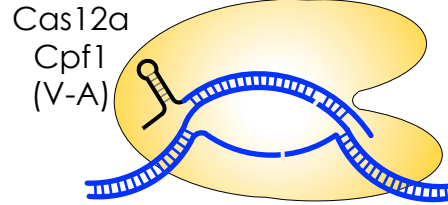
## Class 2 (single-subunit effector complex)

Type II



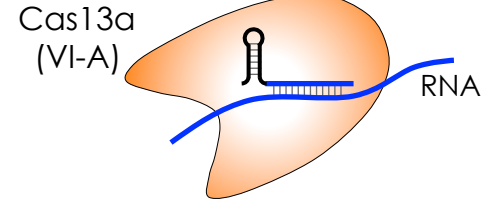
Target: dsDNA

Type V



Target: dsDNA

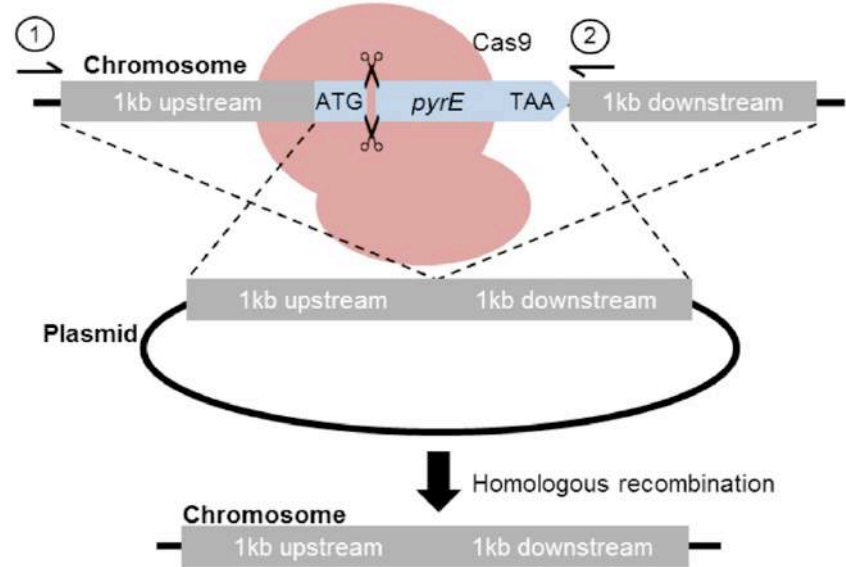
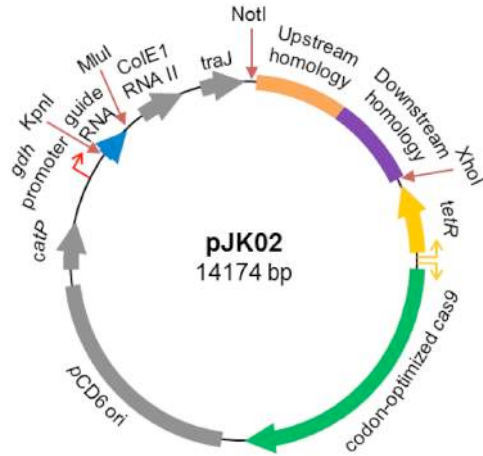
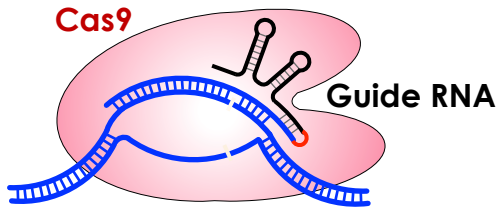
Type VI



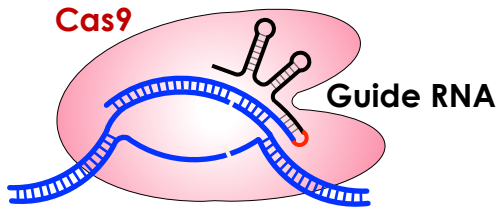
Target: ssRNA

# CRISPR-Cas9-based tool

McAllister et al. *Sci Reports* 2017

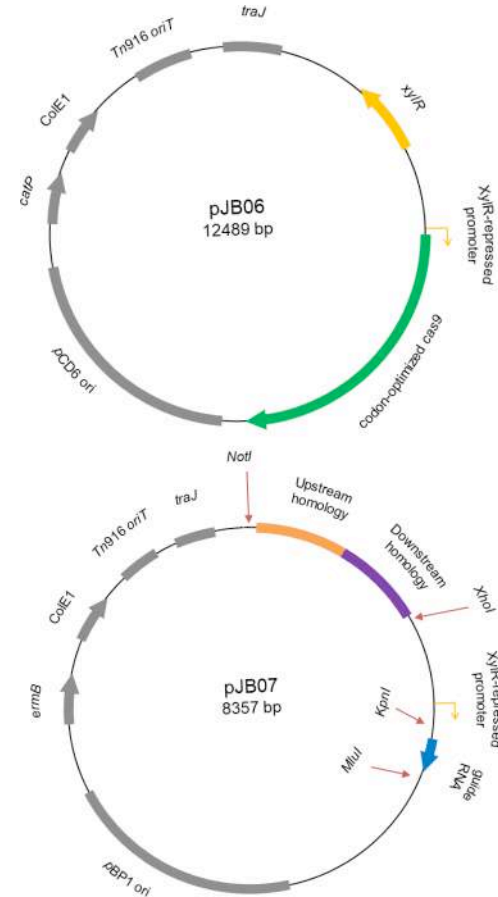


# Optimized CRISPR-Cas9-based tool

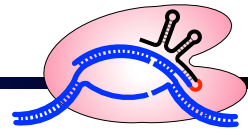


**A two-plasmid mutagenesis system** to avoid toxicity of CRISPR-Cas9 :

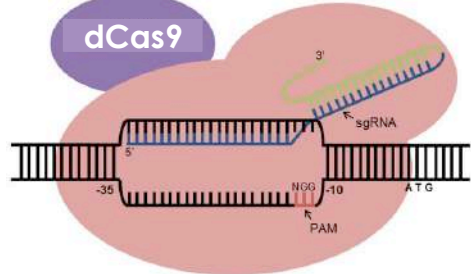
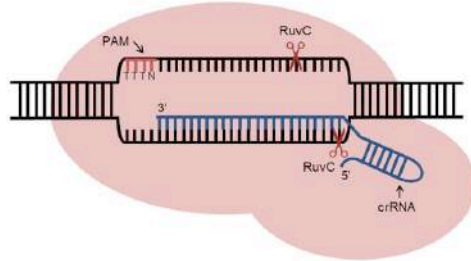
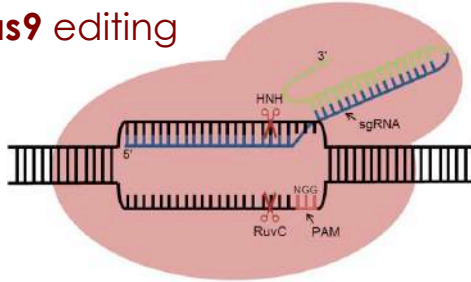
- Xylose-inducible expression of *cas9*
- Xylose-inducible expression of guide RNA



# Variety of CRISPR-Cas-based tools



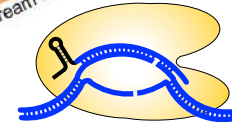
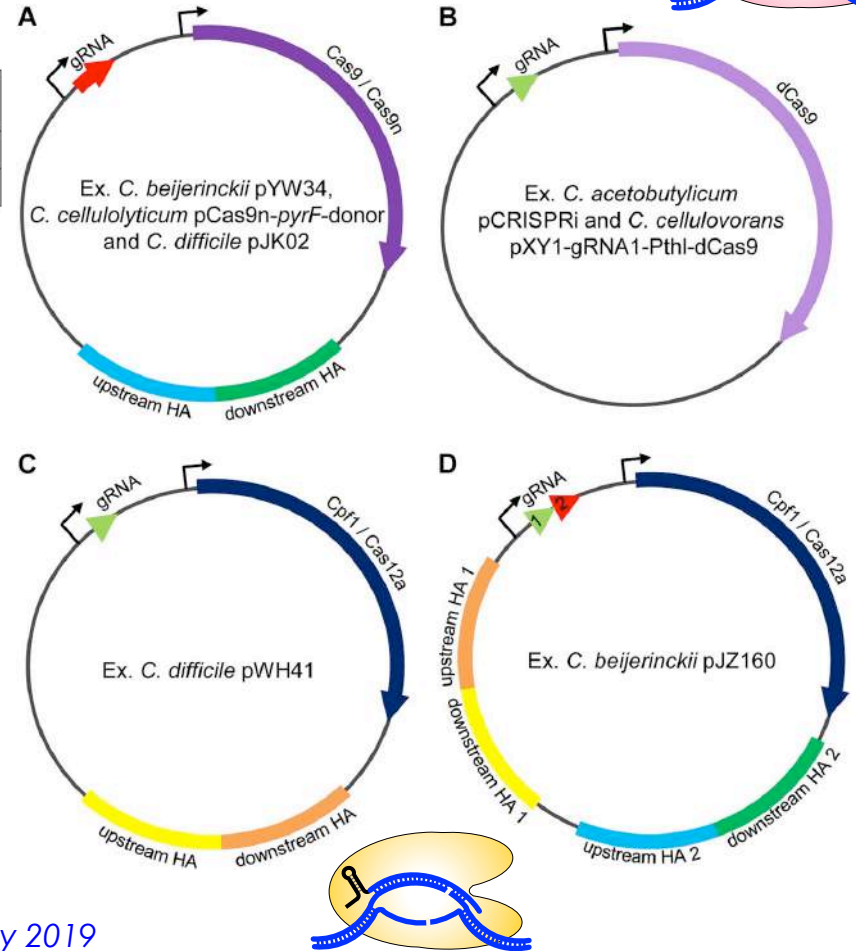
## Cas9 editing



Mutant Alleles		
Mutation	Catalytic activity	Name
D10A	$\Delta$ RuvC	Cas9n, "nickase"
D10A / H840A	Dead	dCas9 / CRISPRi

## Cpf1/Cas12a editing

## dCas9 CRISPRi gene knock-down





# Endogenous *C. difficile* CRISPR-Cas system



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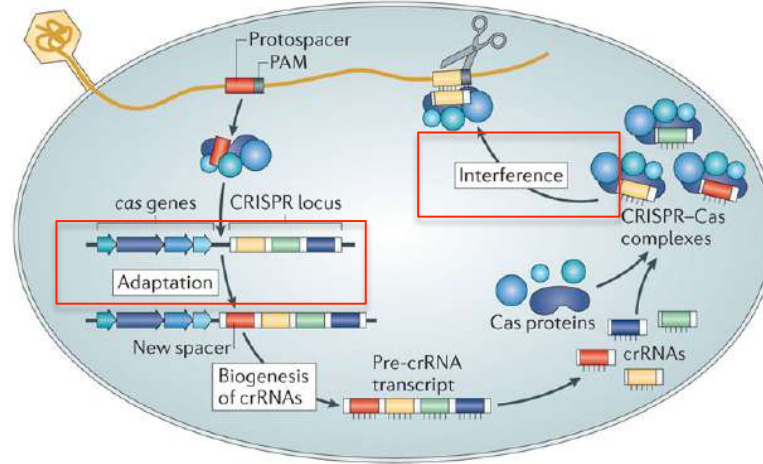
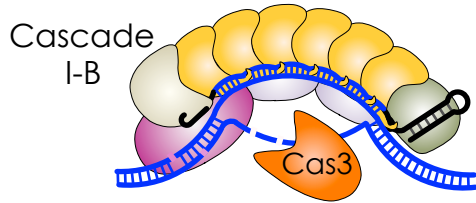


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SAINT-ANTOINE

Boudry et al. *mBio*. 2015  
Maikova et al. *NAR*. 2018  
Maikova et al. *Front Microbiol.*  
2018  
Maikova. PhD thesis  
Maikova et al. *mBio*. 2021  
Muzyukina et al. *mSphere*. 2023

Hargreaves et al. 2014  
Andersen et al. 2016



In CD strain 630 : 12 CRISPR regions exist and are expressed

In epidemic CD strain 027: 9 active CRISPR arrays

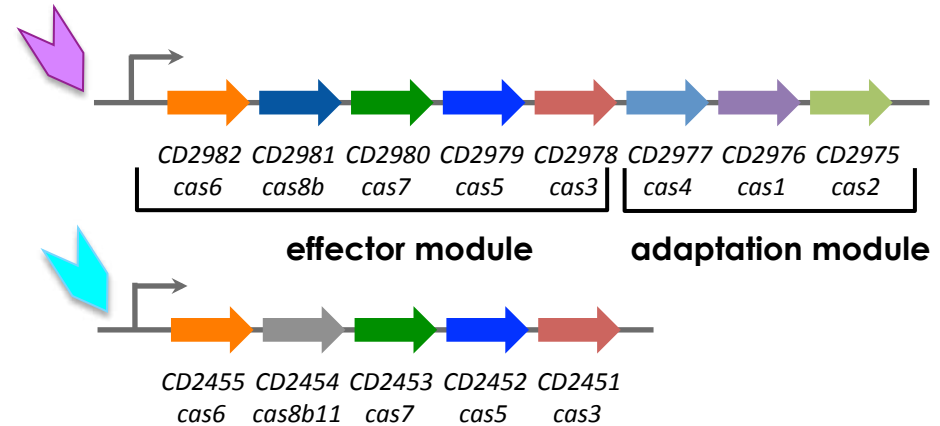
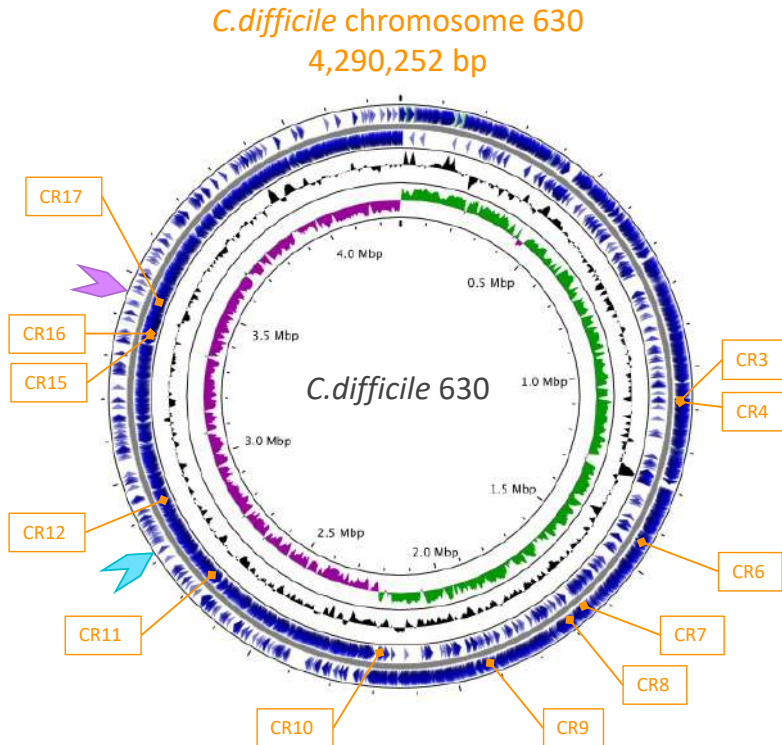


.... A total of 819 spacers from nine CD strains

**Large defence capacity within phage-rich gut communities**

# CRISPR analysis in *C. difficile*

- **Type I-B CRISPR-Cas system** (active for interference and adaptation)
- **Large number of arrays** (the average of 8 array/genome, 12 in 630 strain)
- **2-3 cas gene sets (type I-B)** present in the majority of strains (about 2000)
- Location of arrays in **phage** regions (**5** in 630 strain, 3 highly expressed)



Boudry et al. *mBio*. 2015  
Maikova et al. *NAR*. 2018  
Maikova et al. *Front Microbiol*. 2018  
Maikova et al. *AEM* 2019  
Maikova. PhD thesis  
Maikova et al. *mBio*. 2021  
Muzyukina et al. *mSphere*. 2023



# C. difficile CRISPR-Cas system applications



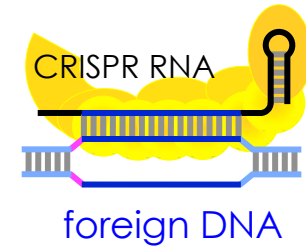
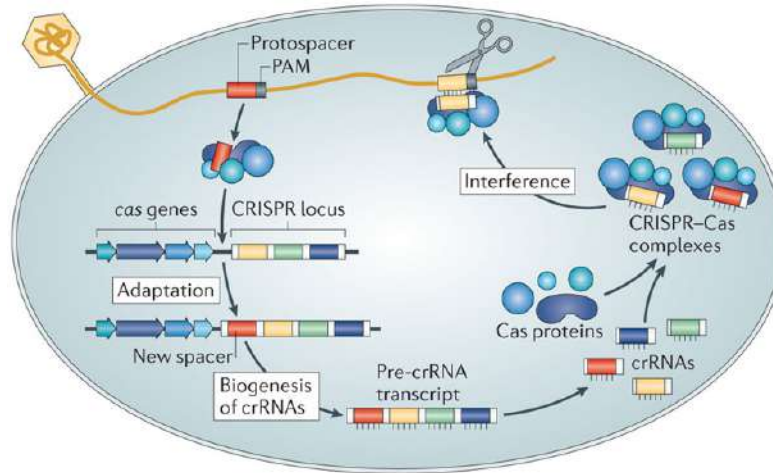
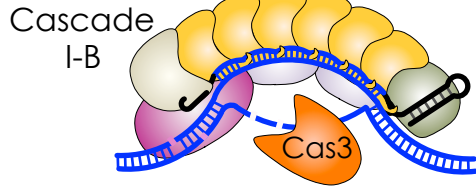
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Boudry et al. *mBio*. 2015  
Maikova et al. *NAR*. 2018  
Maikova et al. *Front Microbiol*. 2018  
Maikova. PhD thesis  
Maikova et al. *AEM*. 2019  
Muzyukina et al. *In preparation*  
Andersen et al. 2016



Sensitive high-resolution CRISPR-based typing for epidemiology & CD microevolution survey

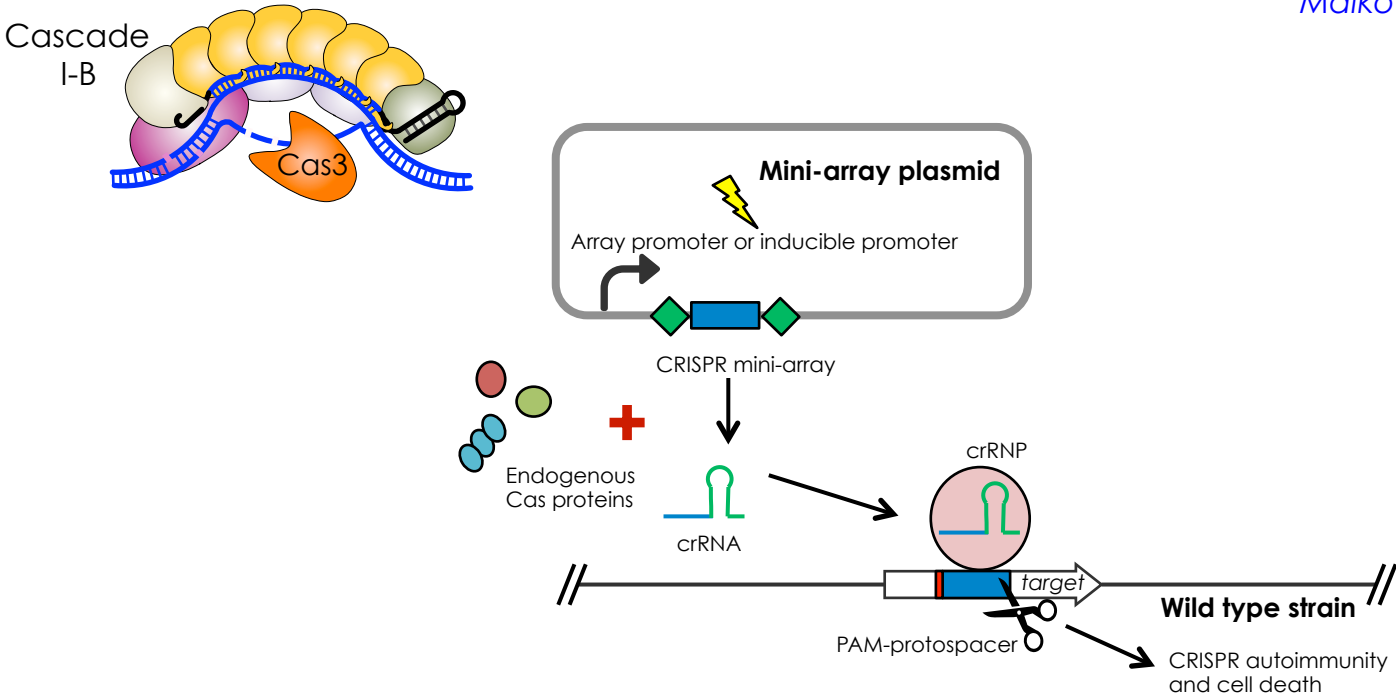
**Endogenous system for CD genome editing**

Self-targeting, autoimmunity

**Great potential for genome editing and therapeutic applications**

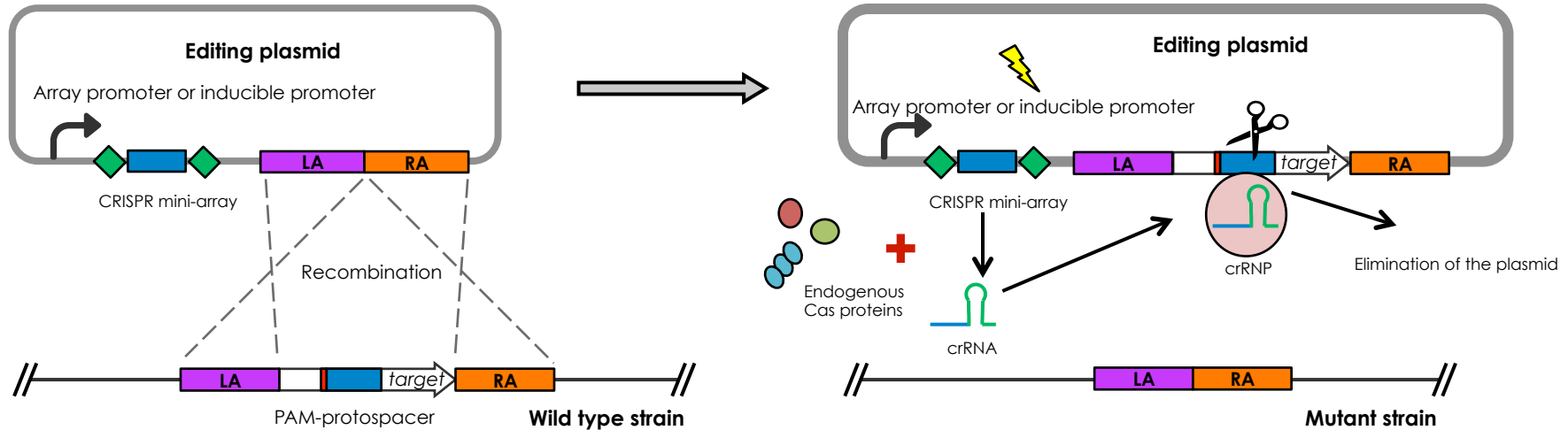
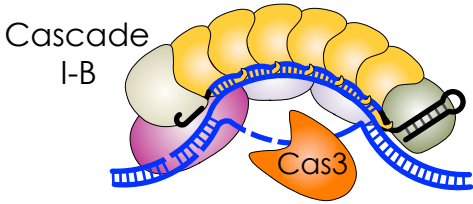
# Genome editing in *C. difficile* using the endogenous CRISPR-Cas system

Maikova et al. *Frontiers Microbiol.* 2018  
Maikova. PhD thesis  
Maikova et al. *AEM.* 2019



# Genome editing in *C. difficile* using the endogenous CRISPR-Cas system

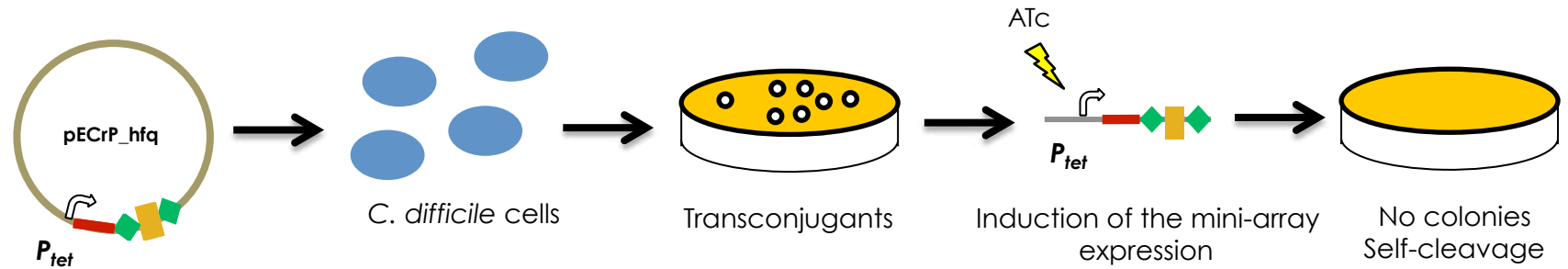
Maikova et al. *Frontiers Microbiol.* 2018  
Maikova. PhD thesis  
Maikova et al. *AEM.* 2019



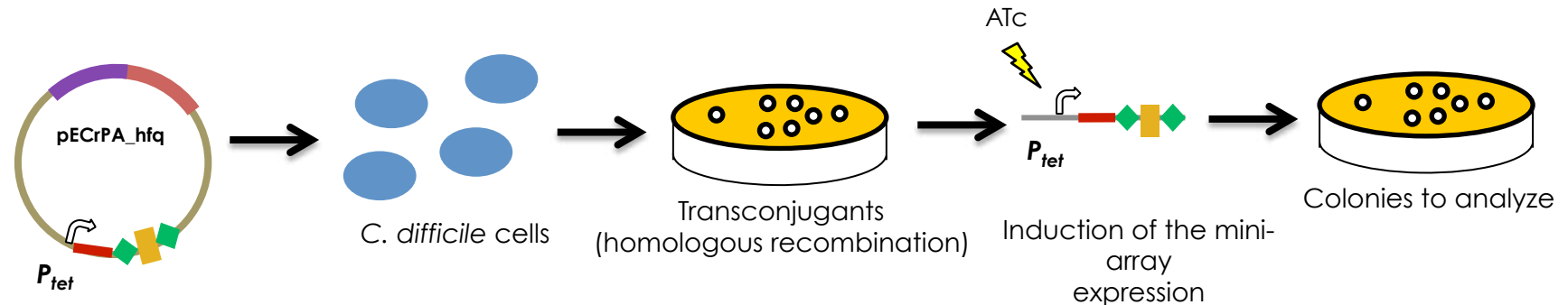
No need of unstable plasmid:  
CRISPR-Cas cleavage

# Genome editing in *C. difficile* using the endogenous CRISPR-Cas system

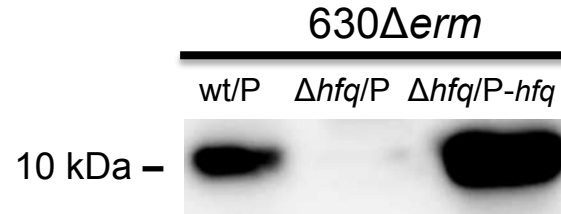
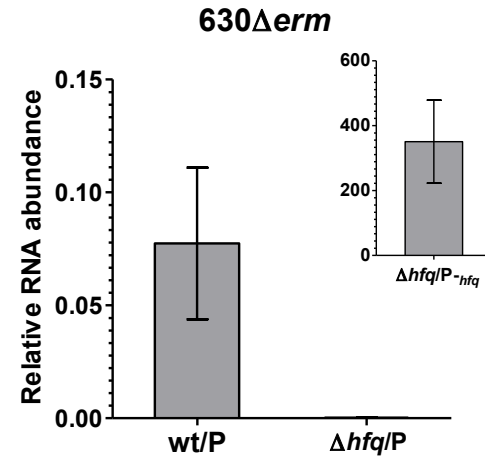
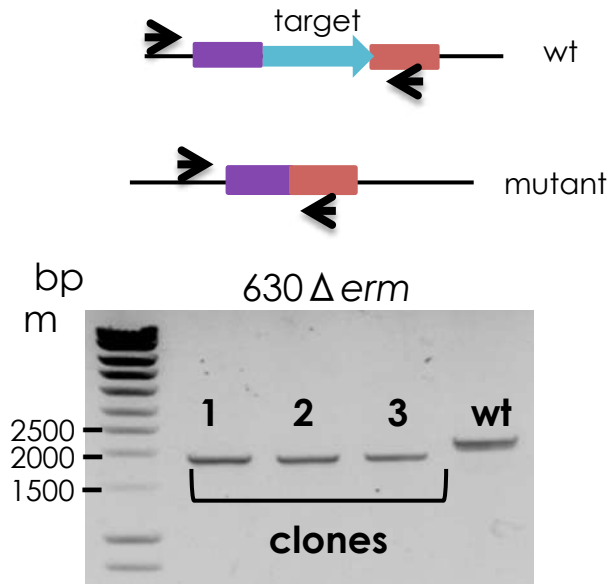
Induced endogenous **CRISPR-Cas autoimmunity** to check **CRISPR spacer efficiency**



**Mutant construction** using plasmid with homologous arms



# Genome editing in *C. difficile* using the endogenous CRISPR-Cas system



- **Δhfq mutants** in 630 and R20291 strains
- **Could not be obtained** using **other** genome editing methods (codA allele exchange and Clostron)
- **Efficient plasmid** loss



# Various mutagenesis types using CRISPR-Cas

## Arms variants

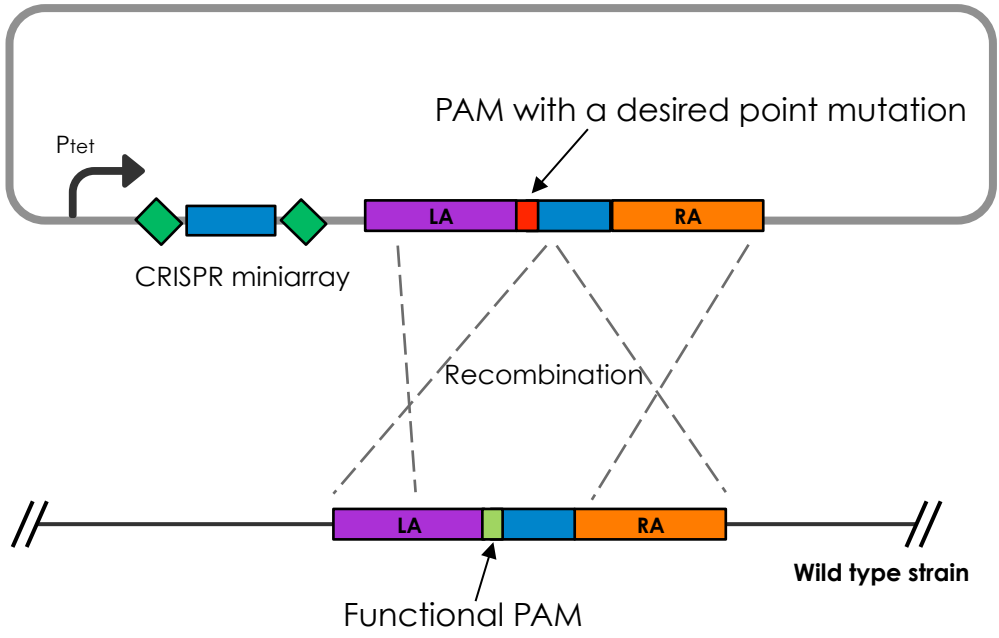
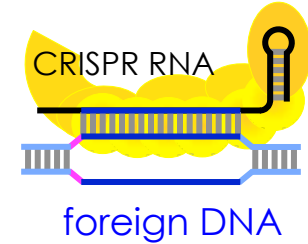
Deletion



Insertion



Point mutation



*C. difficile* functional PAMs:  
**CCN and TCN**

Mutations in PAM for example:  
CTN, ACN

Mutations in first protospacer position  
(seed region from +1 to +6)

Combination of mutations

# CRISPR-Cas-based tools

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## Types of mutations:

- Deletions (marker-less gene inactivation)
- Substitution (promoter replacement ...)
- Insertion (tag for chromosomal gene copy, complementation ...)
- Point mutations
- Gene knockdown by CRISPRi
- Multiple targeting (several crRNAs or gRNAs)

## Remaining challenges:

- Conjugation efficiency in epidemic strains
- Homologous recombination efficiency

# Clostridioides *difficile*

## Challenges

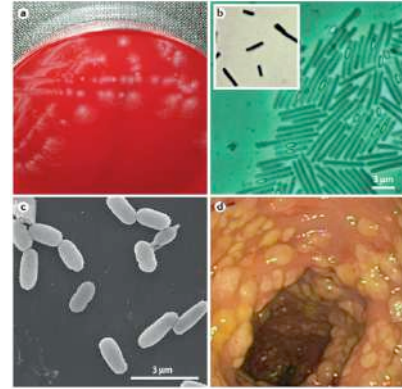


✓ **Difficult** to isolate

✓ **Difficult** to culture requiring strictly anaerobic conditions

✓ **Difficult** to manipulate genetically

Lack of efficient genetic tools



*Smits et al. Nat Rev Dis Primers 2016*