

## Microbial production of therapeutic interest molecules

**Master 2 Microbiology and Biological engineering**  
***Teaching Unit "Medico- Pharmaceutical Applications of microbial biodiversity"***

# Agenda

- Introduction: Health and Biotechnologies
- Production process for therapeutic bioproducts: USP/DSP
- Example for Antibiotics production
- Example for Recombinant Proteins

# Introduction: Biotechnology Concept

Pharmaceutical biotechnology: process using microbial factories, plants or animals for the production of pharmaceutical products

**Bioproduction** is the production of biologics-based therapeutic drugs

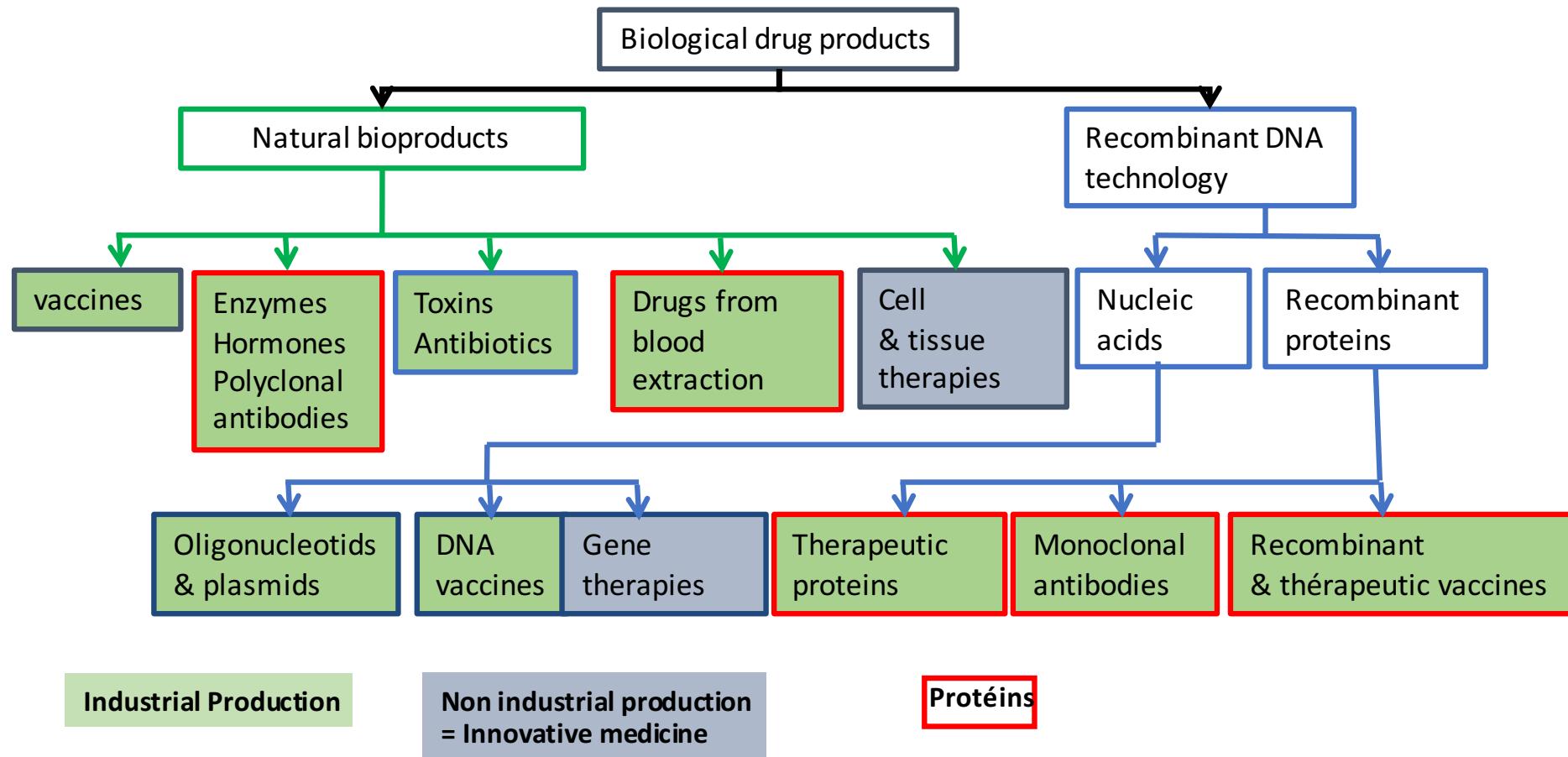
## Biodrug:

- Drug substance produced by or extracted from a **biological source**
- Well characterized & described **manufacturing and purification process**
- **Characterization** and **quality** assessment need a **combination** of physical, chemical and biological assays

*EU definition of Biologic (Directive 2001/83/EC as amended, Annex 1 Active substance 3.2.1.1.b)*

# Biodrug classification

The CONCEPTION and the PRODUCTION  
are a biotechnological process



# Biogrugs and micro-organisms

- **Proteins:**

- Recombinant: hormones, antibodies and derivates, cytokines...
  - Bacteria, yeast, insect cells, mammal cells
- Natural production: enzymes, hormones, antibodies, albumin, collagen...
  - Bacteria, yeats, fungi, mammal cells

- **Nucleic acids**

- DNA vaccines: bacteria
- Plasmids, oligonucleotides: bacteria
- Viral vectors: viruses (gene therapy)

- **Antibiotics:** fungi, bacteria

- **Lipids, polysaccharids, organic complex molecules:** Yeast, bacteria

- **Vaccines**

- Microbial: virus, bacteria
- Recombinants: bacteria, yeast, mammal cells, insect cells

# The production process: an overview

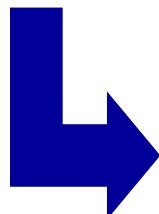
# Bioproducts need complex industrial biological mastered process

- ✓ Biological materials isolation
- ✓ Modification
- ✓ multiplication

- ✓ Characterization & sequencing
- ✓ Cloning
- ✓ Expression

*Genetic engineering*

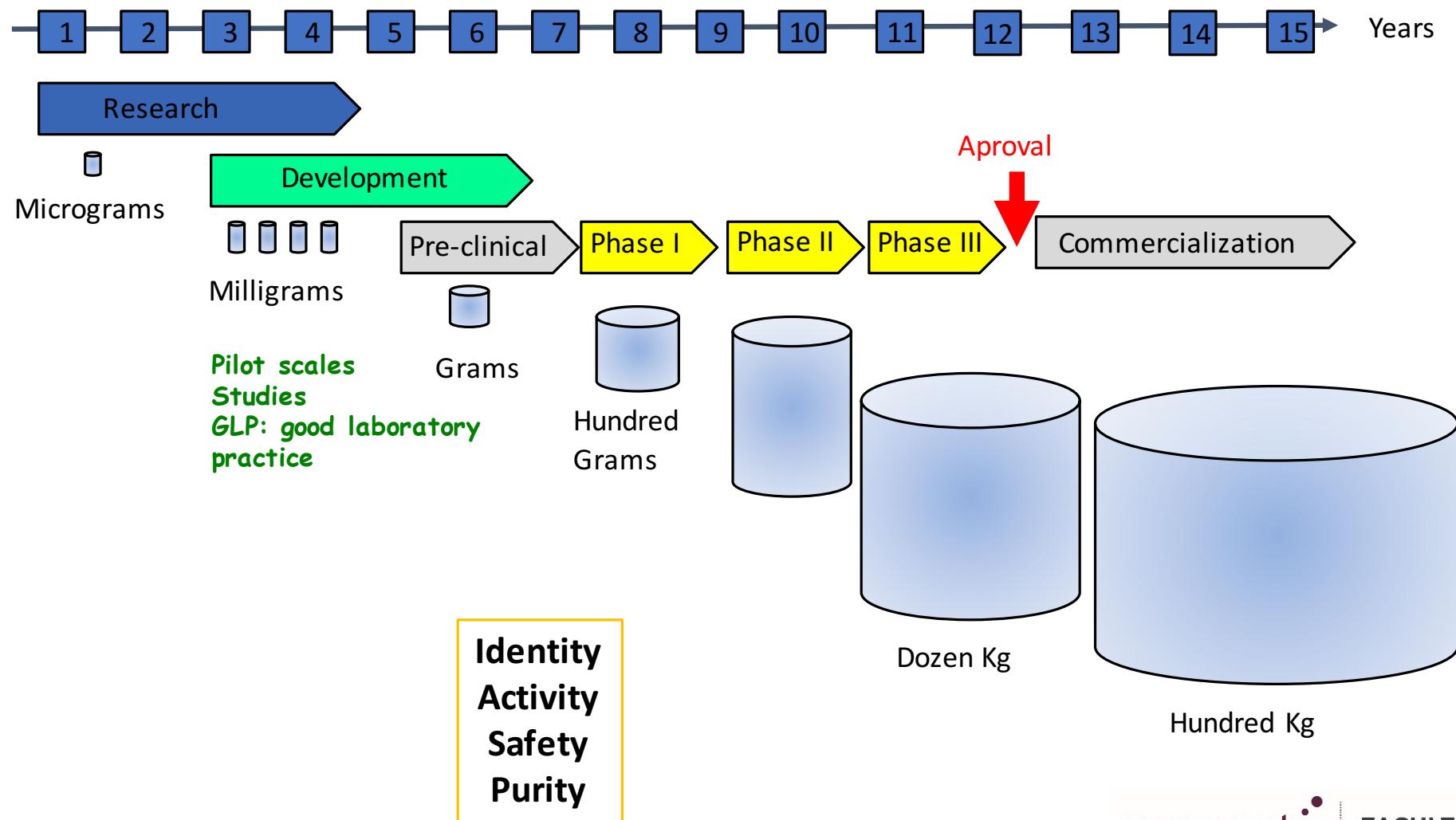
- ✓ Simple and robust process
- ✓ Energy balance, raw materials, waste treatments
- ✓ Regulatory environment



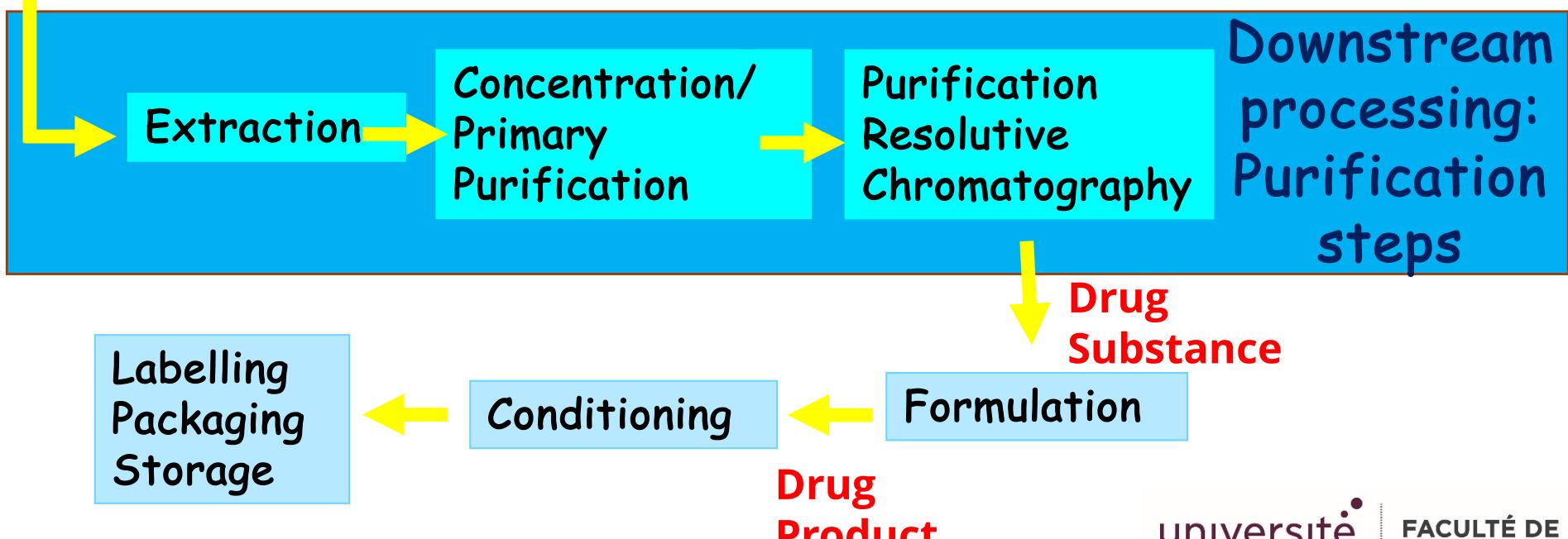
- ✓ Industrial production
- ✓ Purification and characterization
- ✓ Quality control
- ✓ Conditionning



# From research to commercialization



# General scheme of a production process



# Upstream processing (USP)

## Productive strain choice

Rusticity: adaptability to industrial conditions

Performances: growth speed, production speed, production yield...

Genetic stability

Bioreactors, required functions:

- Confining
- Sterility
- Mixing
- Aeration + air filtration
- Nutrient and fluid supply
- Cleaning procedures

## Microbial Culture parameters

Physical, chemical:  
pH, pO<sub>2</sub>, T°, osmotic pressure

Kinetics:  
Dilution, nutrients, culture length...

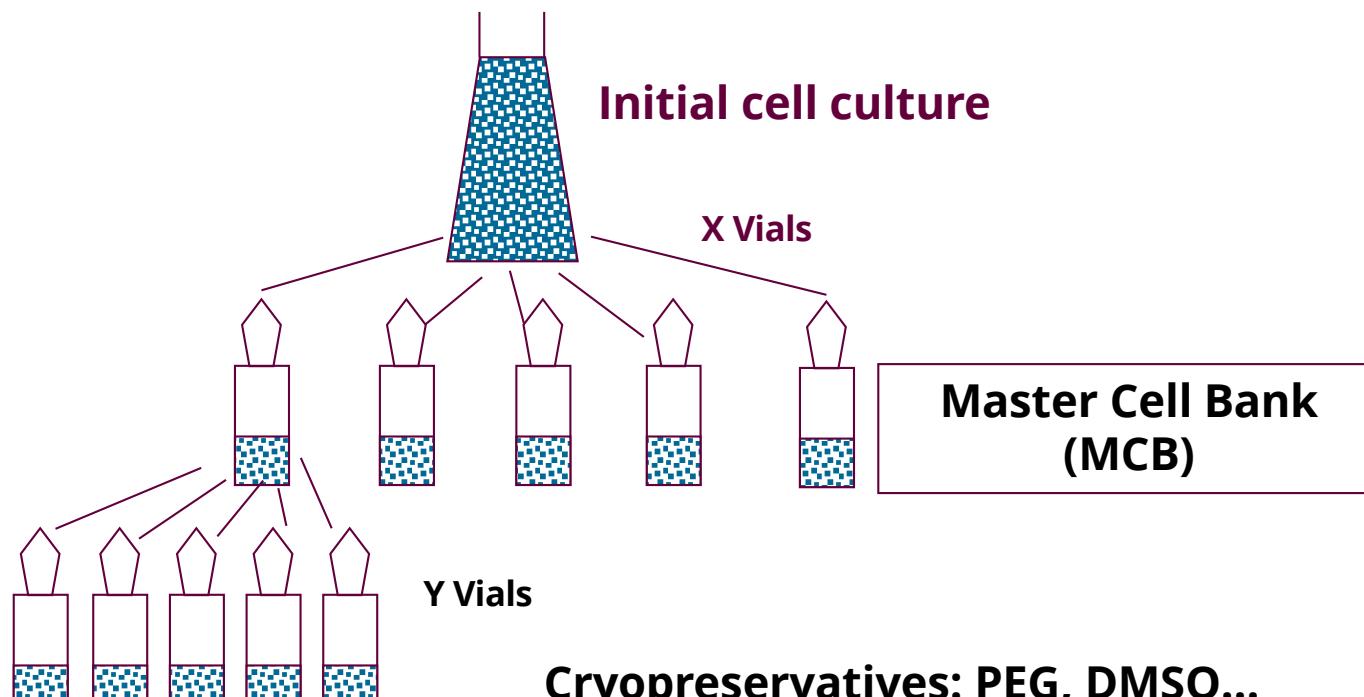
Nutrient composition :  
Nitrogen, carbon, growth factors,  
lipids, oligoéléments (Mn, Fe, Co, Ni, Cu,  
Zn, Mo)...

Metabolit accumulation

Process modelisation and automation

# BioBanking

Goal: to store and securize the well characterized initial cell culture



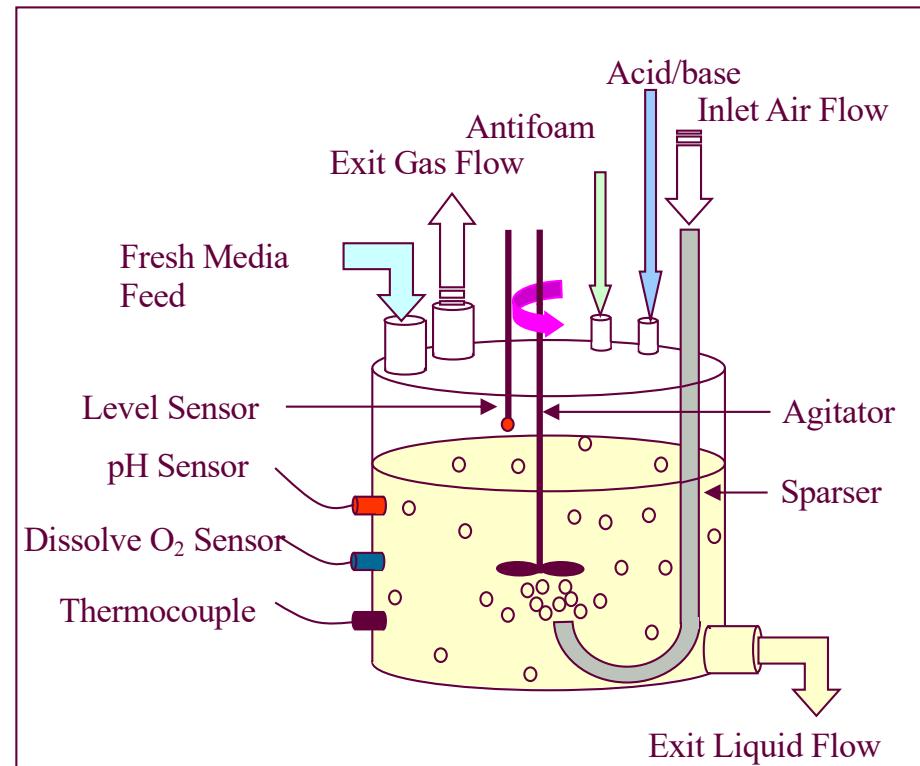
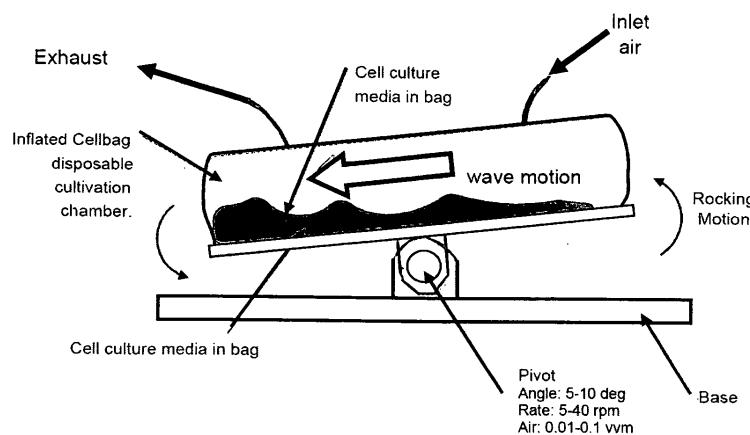
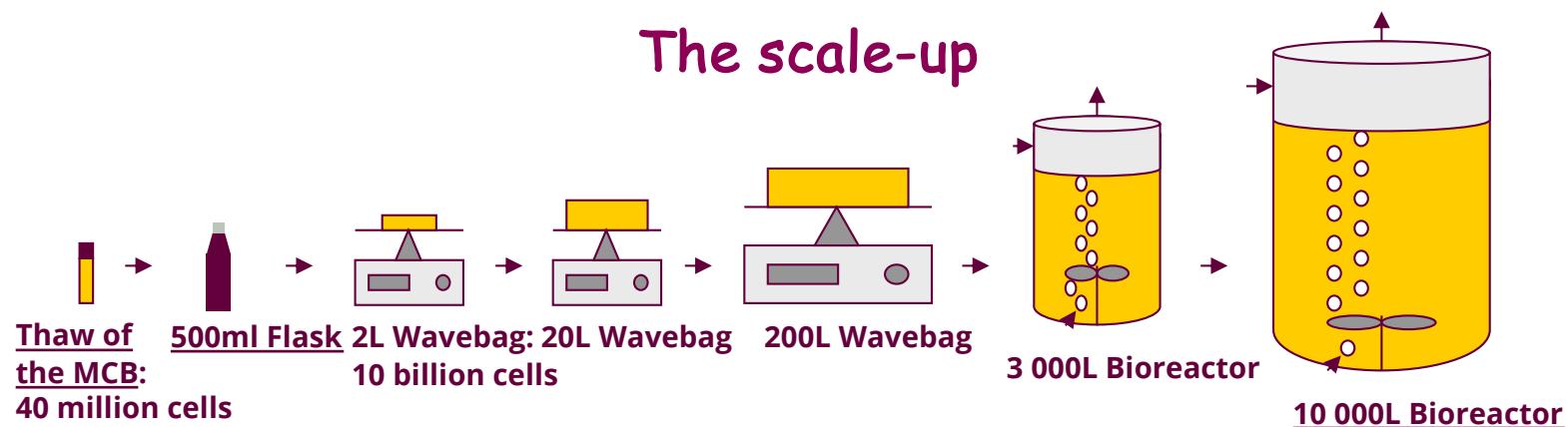
## Quality control

Cell bank characterization tests

Culture purity
Species identity
DNA sequencing
Plasmid identity and integrity
Host strain identity
Viable count
Specific productivity
Plasmid retention
Restriction endonuclease mapping

- ✓ Repetability of cell growth
- ✓ Quality control procedures and cell bank viability

## The scale-up



Stirred-tank bioreactor



## 3 production modes

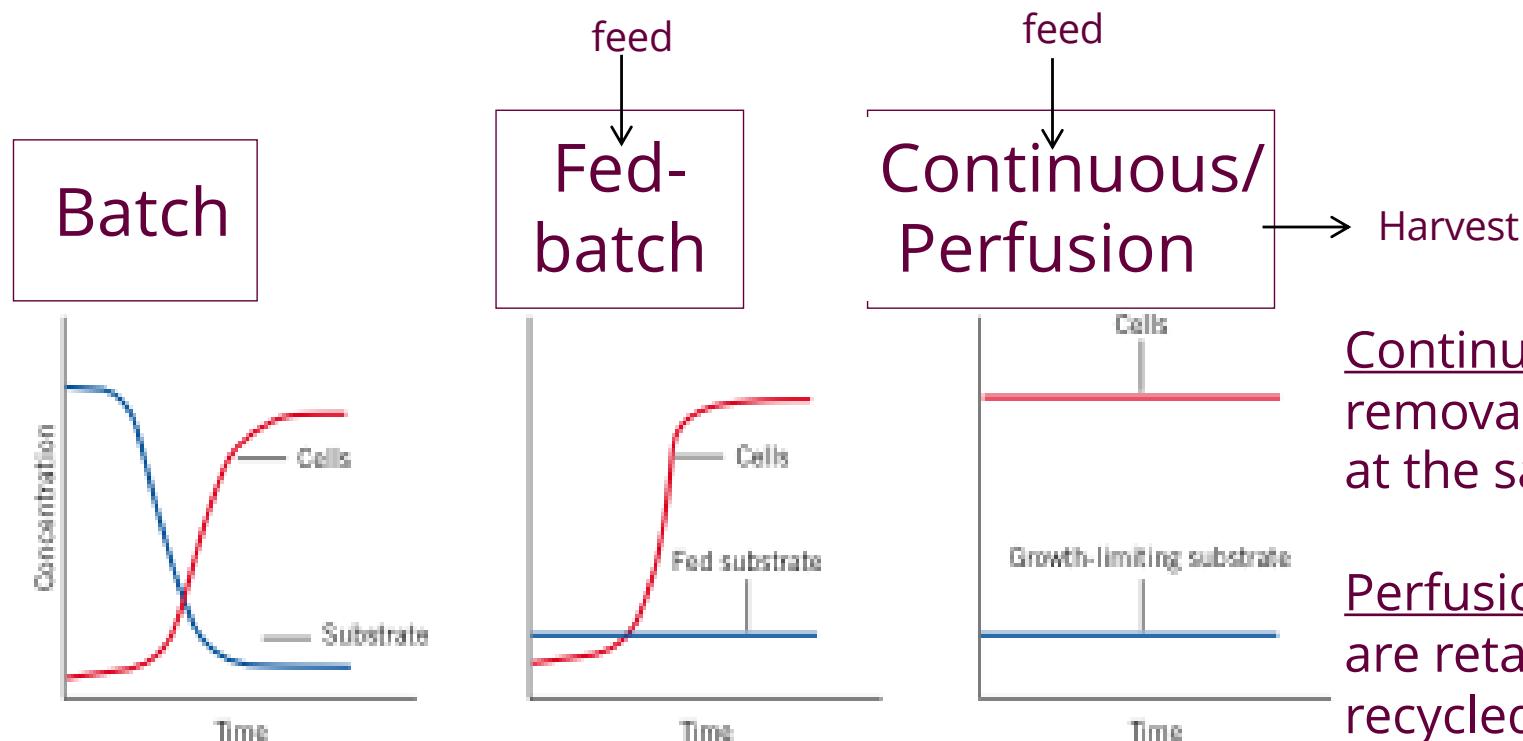


Fig. 95. Comparison of batch and perfusion concerning nutrients and waste products.

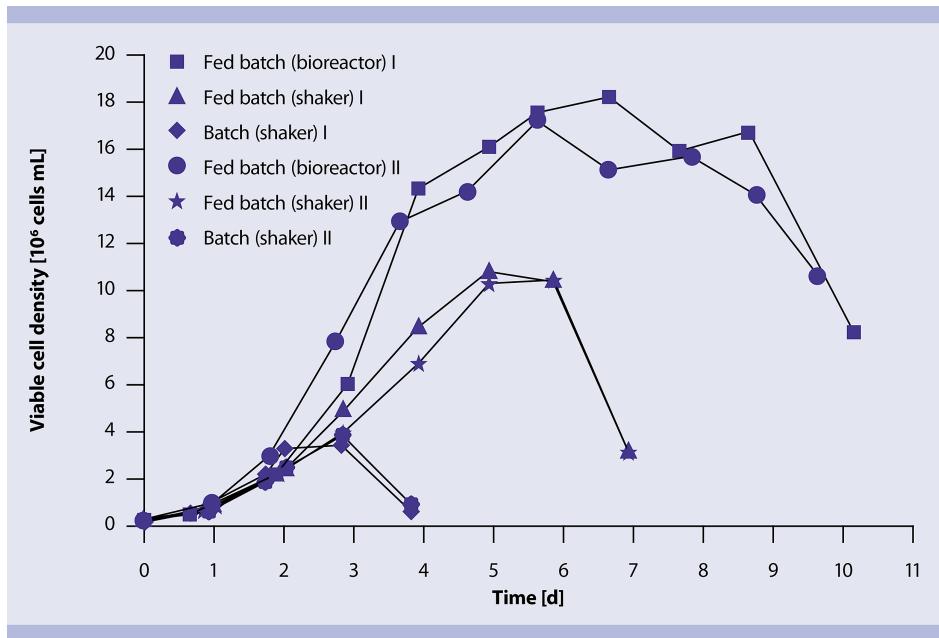
- ✓ Variable medium composition
- ✓ Toxic metabolite accumulation

- ✓ Increase cell density
- ✓ Easy automation
- ✓ Bioproduct accumulation

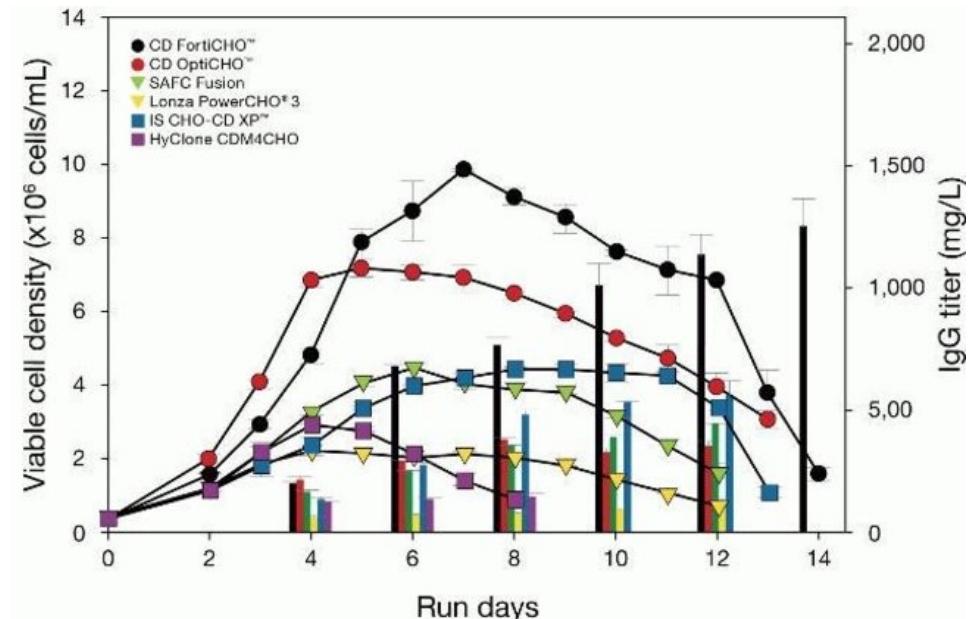
- ✓ Metabolite elimination
- ✓ Stable medium composition
- ✓ Reduced degradation risk

# Process development USP – cell growth optimization

## Bioreactor comparison



## Cell growth media comparison



### Critical issues by regulatory authorities:

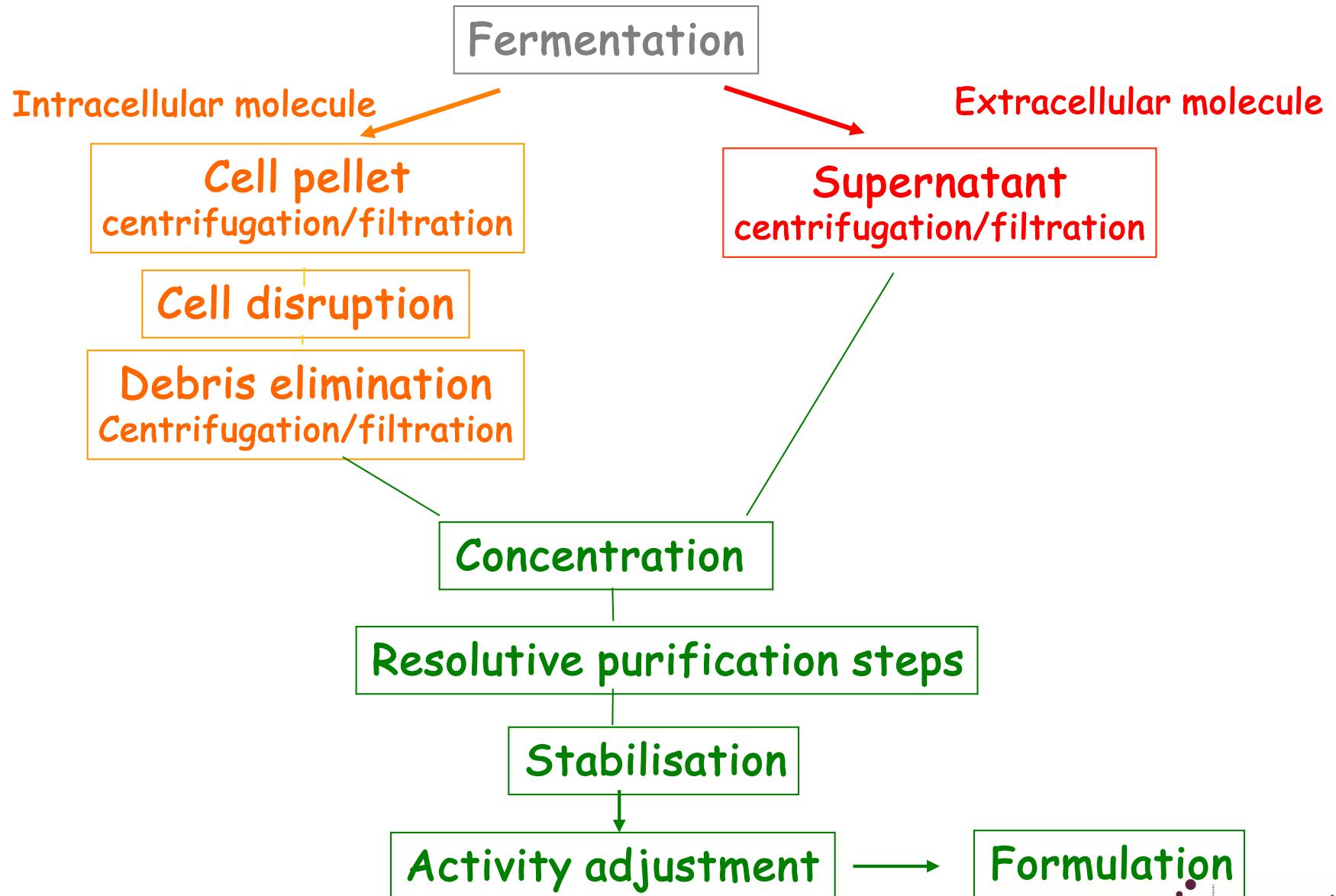
- ✓ safety,
- ✓ genetic and phenotypic stability,
- ✓ monoclonality of the production cell line

→ Rigorous cell line development:

Full documentation

Full characterization of the final production cell line

# Downstream processing (DSP)



## Centrifuge



Filtration installation

Chromatography

# Example 1:

## Antibiotic production

## Main AB producers

**Secondary metabolites:  
stationnary phase  
synthesis**

**80% Bacteria, steptomyces**

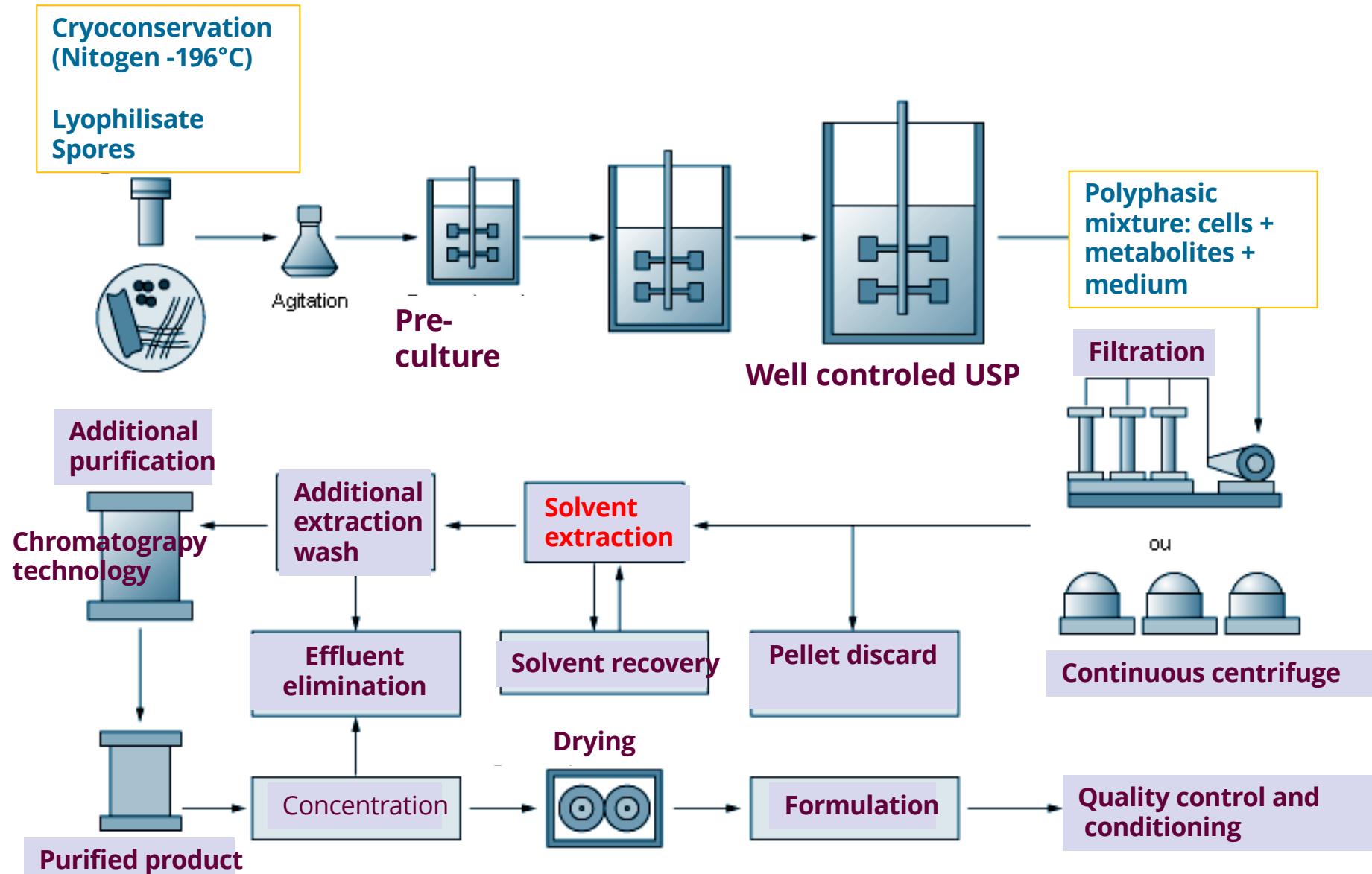
***Bacillus***

**Fungi**

***Aspergillus, Penicillium,  
Cephalosporium,  
Heminthosporium, Fusidium***

Classification	Antibiotique	Organisme producteur
Aminoglycosides	Streptomycine	<i>S. griseus</i>
	Spectinomycine	<i>S. spectabilis</i>
	Néomycine B	<i>S. fradiae</i>
Tétracyclines	Chlortétracycline	<i>S. aureofaciens</i>
	Oxytétracycline	<i>S. rimosus</i>
Polyènes	Nystatine	<i>S. noursei</i>
	Amphotéricine	<i>S. nodosus</i>
Macrolides	Spiramycine	<i>S. ambofaciens</i>
	Érythromycine	<i>S. erythreus</i>
	Rapamycine	<i>S. hygroscopicus</i>
	Natamycine	<i>S. natalensis</i>
	Avermectine	<i>S. avermitilis</i>
	Tylosine	<i>S. fradiae</i>
	Oléandomycine	<i>S. hygroscopicus</i>
	Pristinamycine	<i>S. pristinaespiralis</i>
	Virginiamycine	<i>S. virginiae</i>
Glycopeptides	Vancomycine	<i>S. orientalis</i>
Lincosamides	Lincomycine	<i>S. lincolnensis</i>
Autres	Chloramphénicol	<i>S. venezuela</i>

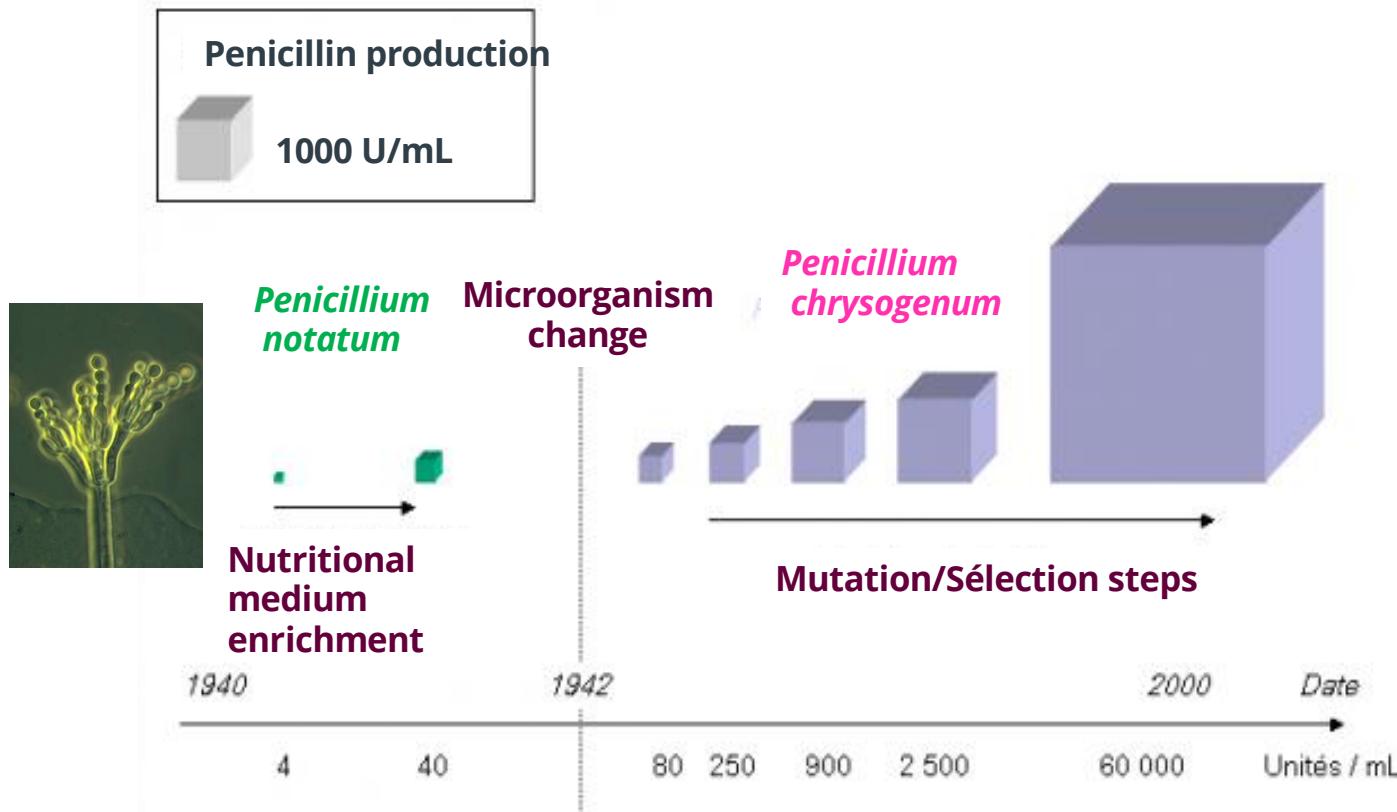
# General Scheme for AB production



Source: Kroschwitz, 1992.

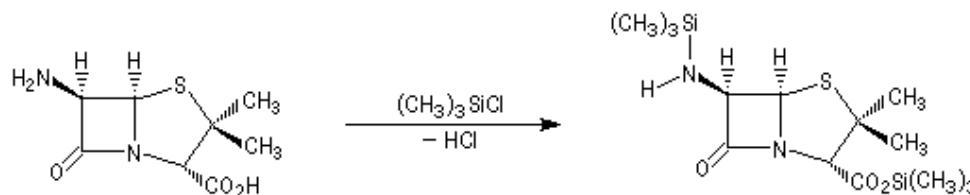
Ideal solvent properties: Solubilisation , Neutrality, Selectivity, Moderate cost, Elimination

- Penicillin example

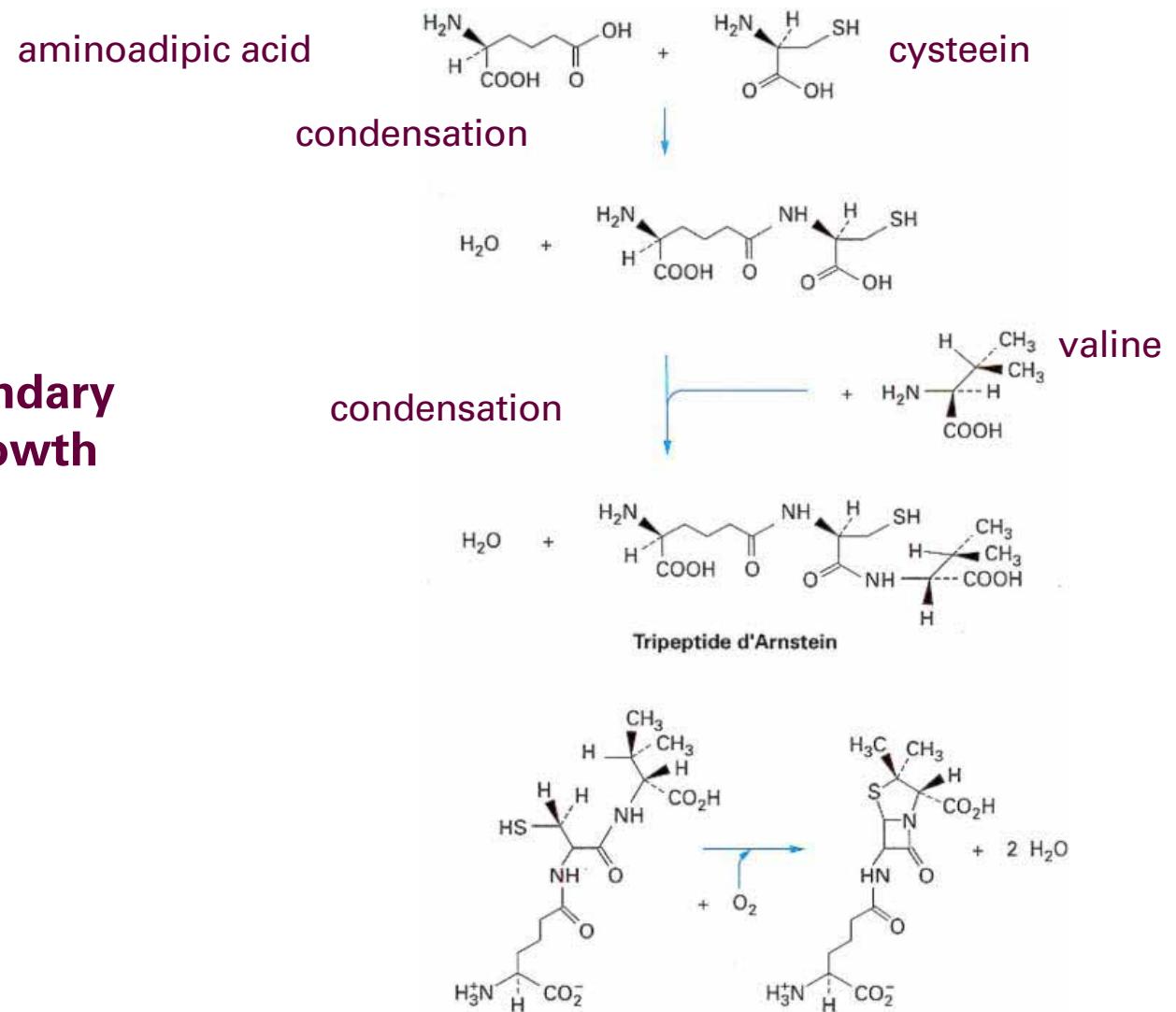


**Production pH≈7  
Penicillin excretion at early stationnary phase;  
Organic solvent extraction**

- ✓ Aqueous phase
- ✓ Penicillin cristallisation
- ✓ Hémisynthesis:  
transformation into derivates AB: ampicillin amoxicillin...



**Penicillin biosynthesis secondary metabolism → end of cell growth**



Oxidative cyclisation → isopenicillin N

# Quality control of antibiotics

**Final step = dessication (=lyophilisation)**

**Essays described in the European Pharmacopeia**

- Identification
- Impurity dosage: derived molecules, pyrogenic substances, histamin...
- Activity

EUROPEAN PHARMACOPOEIA ONLINE



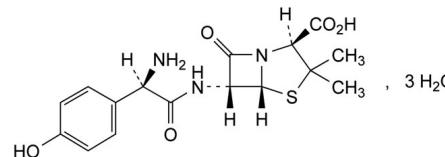
# Example with amoxicillin

## EUROPEAN PHARMACOPOEIA ONLINE

Available With Paris-Saclay Library

### AMOXICILLIN TRIHYDRATE

#### Amoxicillinum trihydricum



C<sub>16</sub>H<sub>19</sub>N<sub>3</sub>O<sub>5</sub>S·3H<sub>2</sub>O  
[61336-70-7]

M<sub>r</sub> 419.4

#### DEFINITION

(2S,5R,6R)-6-[(2R)-2-Amino-2-(4-hydroxyphenyl)acetyl]-amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid trihydrate.

Semi-synthetic product derived from a fermentation product.

Content: 95.0 per cent to 102.0 per cent (anhydrous substance).

#### CHARACTERS

*Appearance:* white or almost white, crystalline powder.

*Solubility:* slightly soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in fatty oils. It dissolves in dilute acids and dilute solutions of alkali hydroxides.

#### IDENTIFICATION

*First identification: A.*

*Second identification: B, C.*

A. Infrared absorption spectrophotometry (2.2.24).

*Comparison:* amoxicillin trihydrate CRS.

B. Thin-layer chromatography (2.2.27).

*Test solution.* Dissolve 25 mg of the substance to be examined in 10 mL of sodium hydrogen carbonate solution R.

*Reference solution (a).* Dissolve 25 mg of amoxicillin trihydrate CRS in 10 mL of sodium hydrogen carbonate solution R.

*Reference solution (b).* Dissolve 25 mg of amoxicillin trihydrate CRS and 25 mg of ampicillin trihydrate CRS in 10 mL of sodium hydrogen carbonate solution R.

*Plate:* TLC silanised silica gel plate R.

*Mobile phase:* mix 10 volumes of acetone R and 90 volumes of a 154 g/L solution of ammonium acetate R previously adjusted to pH 5.0 with glacial acetic acid R.

*Application:* 1 µL.

*Development:* over a path of 15 cm.

*Drying:* in air.

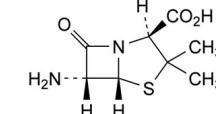
*Detection:* expose to iodine vapour until the spots appear and examine in daylight.

*System suitability:* reference solution (b):

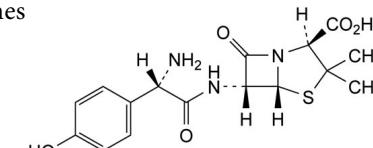
– the chromatogram shows 2 clearly separated spots.

*Results:* the principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

#### IMPURITIES



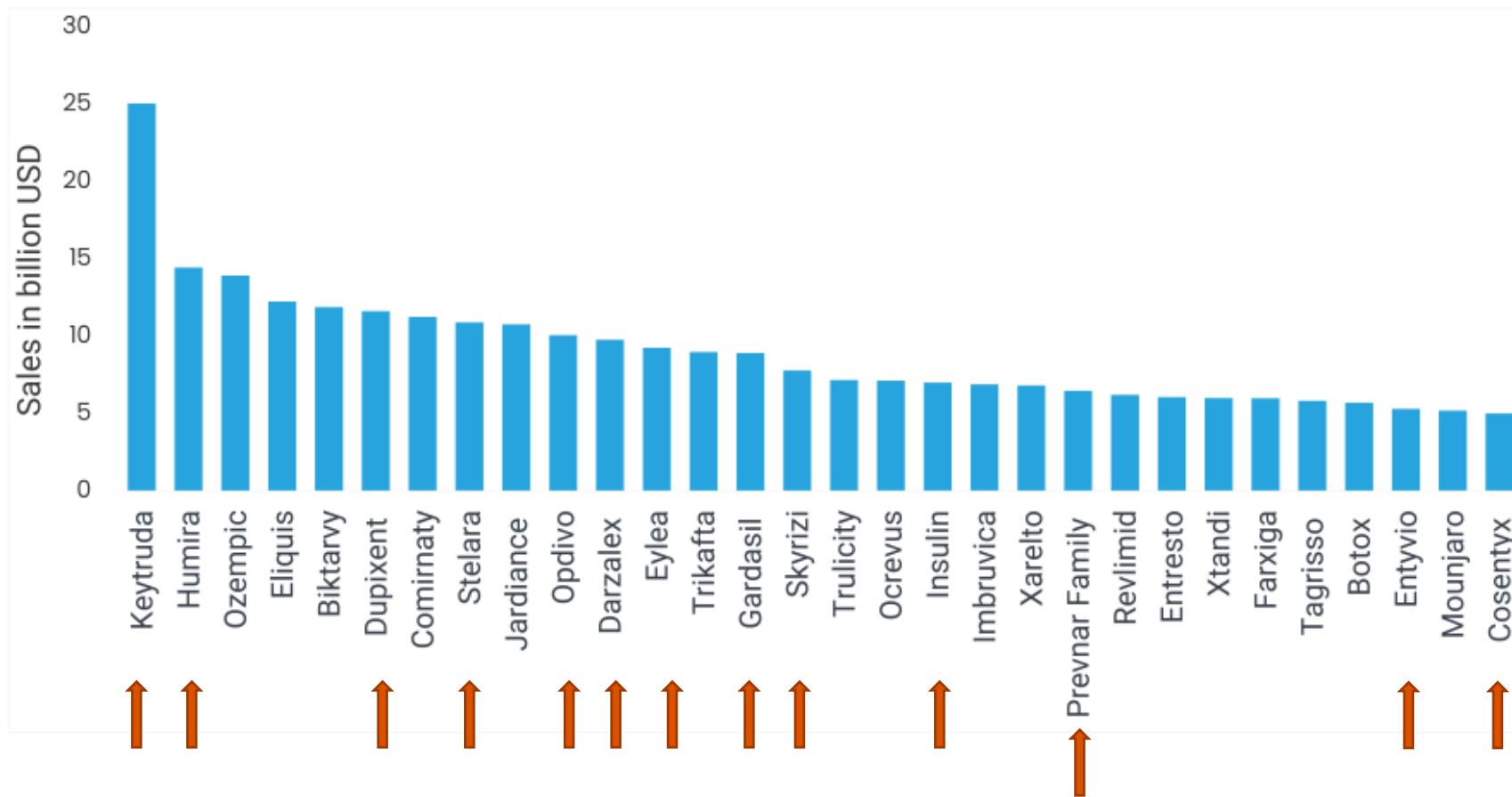
A. (2S,5R,6R)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



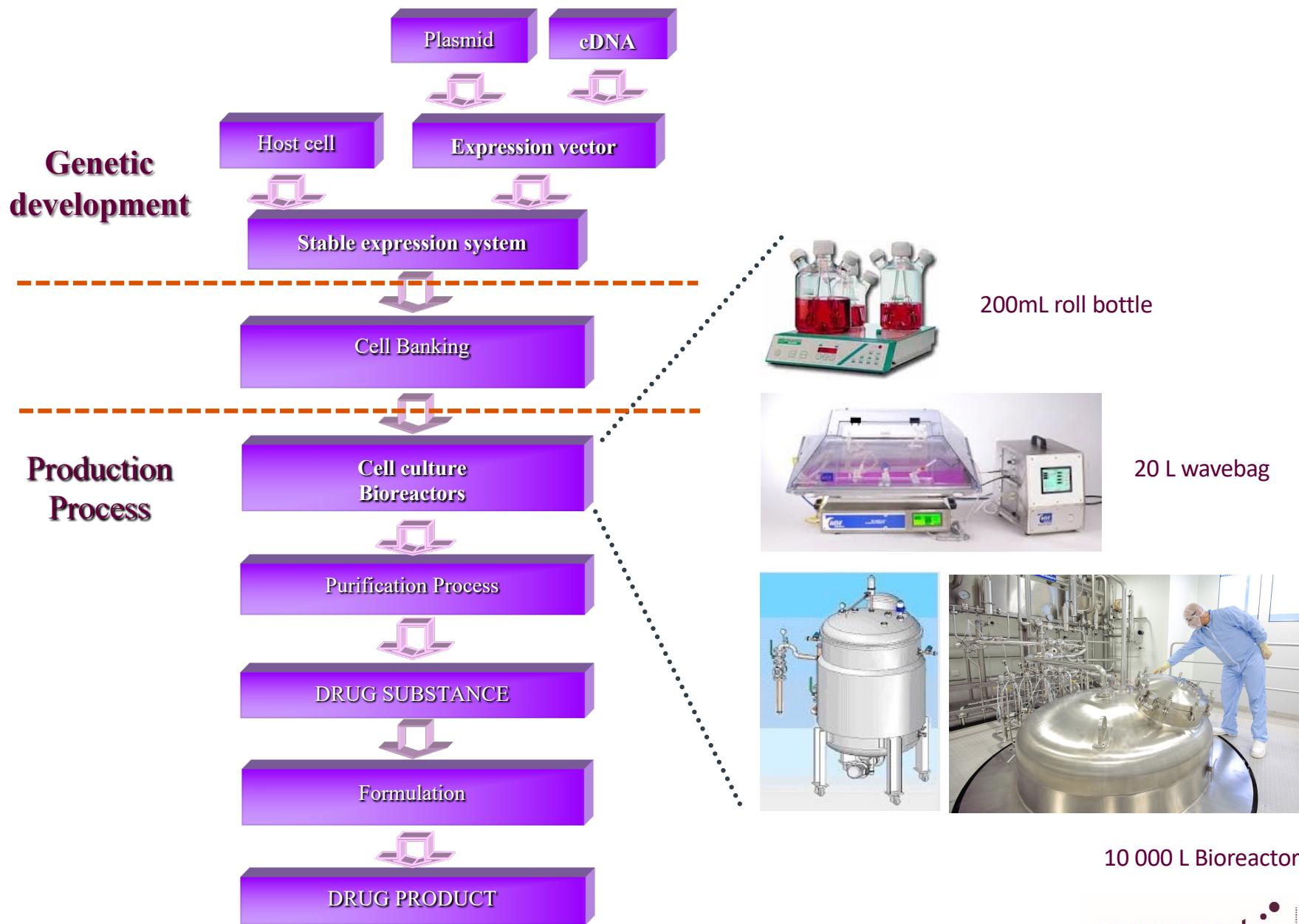
B. (2S,5R,6R)-6-[(2S)-2-amino-2-(4-hydroxyphenyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (L-amoxicillin),

# Example 2: Recombinant protein production

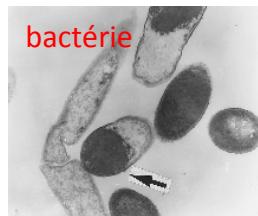
## Top 30 Drugs to Watch in 2024: Insights from 2023 Sales Data



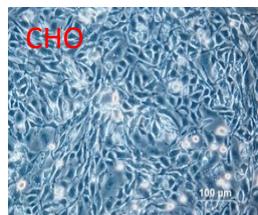
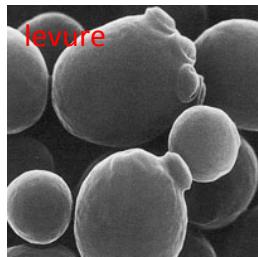
# Recombinant Protein production, an overview



# Recombinant Protein technology, the cell line development



cDNA  
sequence

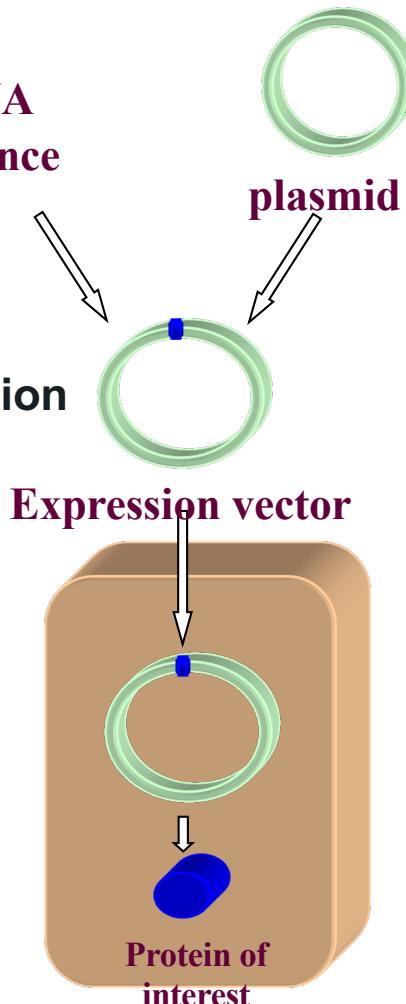


Genetically modified  
organism  
= Host cell

Transfection

Insertion

plasmid



- **GMO choice**
- **Protein expression level**
- **Protein properties**

27

Industrial production  
(cell factory)

**Objective:**  
to generate THE producing  
cell

- ✓ Expression vector optimization
- ✓ Targetting the insertion site
- ✓ Transfected cells selection
- ✓ High producers selection
- ✓ Cell line genetic characterization
- ✓ Culture condition optimization

	E. Coli	Yeast	Mammalian	Insect
<b>Proteolytic clavage</b>	?	?	yes	yes
<b>Glycosylation</b>	non	?	yes	?
<b>Secretion</b>	?	yes	yes	yes
<b>Folding</b>	?	?	yes	yes
<b>Phosphorylation</b>	No	?	yes	?
<b>Acetylation</b>	No	yes	yes	?
<b>Amidation</b>	No	yes	yes	yes
<b>% P / total</b>	>50%	1%	<1%	>30%
<b>MM, quantity</b>	60-70 kDa 100g/L	30kDa 10g/L	<300kDa 1-5g/L	60kDa 200g/L

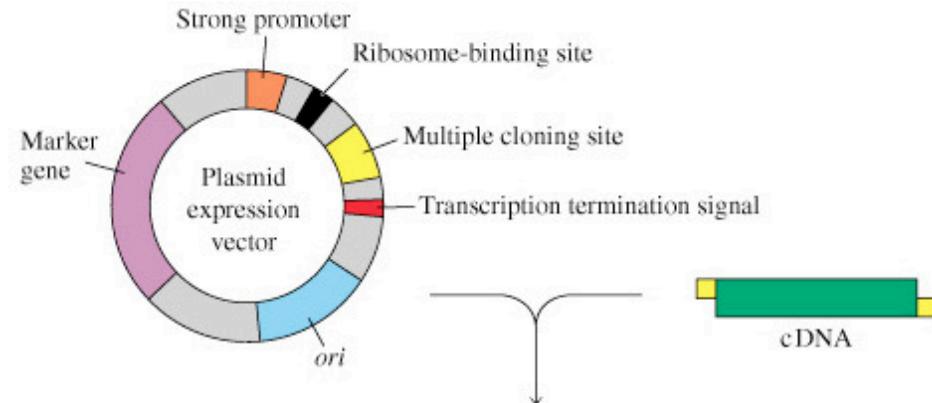
Protein	System	Production level
Hirudin	<i>S. cerevisiae</i> (Y) <i>H. polymorpha</i> (Y)	60 mg/L –
Interferon $\alpha$ -2b	<i>H. polymorpha</i> (Y)	120 mg/L
Hepatitis B vaccine	<i>H. polymorpha</i> (Y)	–
Angiostatin	<i>P. pastoris</i> (Y)	108 mg/L
Anti-HBs Fab	<i>P. pastoris</i> (Y)	50 mg/L
Human serum albumin	<i>K. lactis</i> (Y) <i>S. cerevisiae</i> (Y) <i>P. pastoris</i> (Y)	3 g/L 3 g/L 10 g/L
Human interleukin 6	<i>A. niger</i> (F)	150 mg/L
Human apolipoprotein AI	CHO cells (M)	80 mg/mL
Insulin precursor	<i>P. pastoris</i> (Y) <i>S. cerevisiae</i> (Y)	3 g/L 98 mg/L
Human tPA	CHO cells (M)	34 mg/L
Human gonadotropin	CHO cells (M)	3 g/L
Erythropoietin (epoetin $\alpha$ )	CHO cells (M)	–
Monoclonal Ab	NSO cells (M)	3 g/L
HPV vaccine (Cervarix <sup>TM</sup> )	Insect cells	–
Human proapolipoprotein AI	Insect cells	80 mg/L
Clotting factor VIIa	BHK cells (M)	–

Current Opinion in Biotechnology 2012, 23:965–971

# Protein production in Bacteria (*E.coli*)

## Host bacteria characteristics:

- **recA (-)**
- **dam ou dcm (-)**
- **No endogenous plasmid**

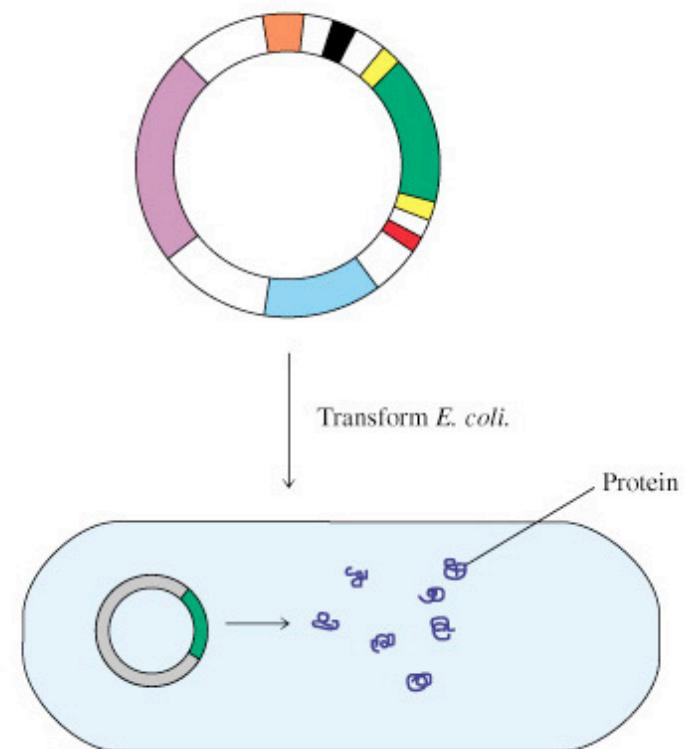


## Intra cellular production

- ✓ Few folding proteins
- ✓ No post-translational modification
- ✓ Reducing environment



Inclusion bodies  
=protein  
precipitation



## PROS

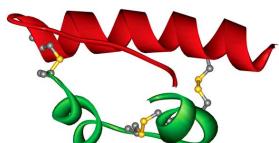
- Fast and inexpensive growth
- Well-known genetic system, many improved strains
- Controllable gene expression
- Many expression vectors available, ease of transfection into host bacteria
- Good protein production yields (>10 g/L culture)
- Cytoplasmic inclusion bodies: purification ease

## CONS

- No post-translational modification
- Inclusion body: insoluble, poorly folded protein
- Presence of bacterial endotoxins: the purification process must include elimination and control steps

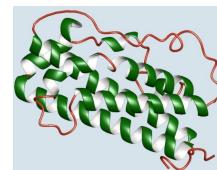


A few therapeutic proteins produced in *E Coli*:



30

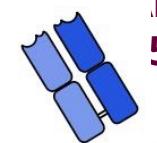
Insulin  
16kDa



Growth hormone  
22kDa

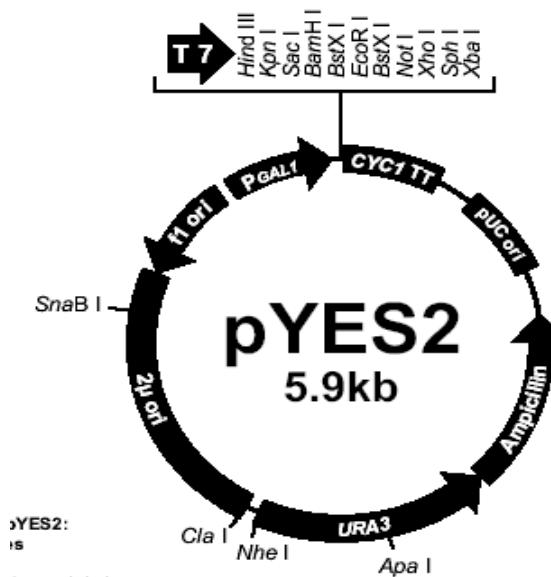
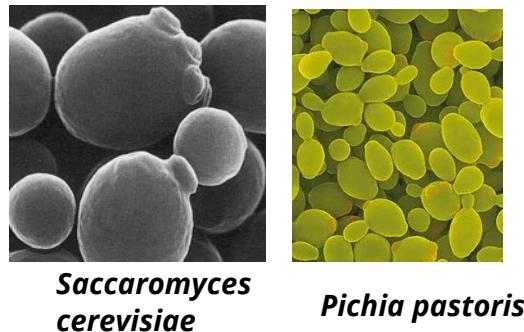


Interferons  
20 kDa



ntibody fragment  
5 kDa

# Protein production in Yeast



## PROS

- Small, eukaryotic genome and well characterized
- Absence of endotoxin
- Inexpensive fermentation
- Good yields ( grams per liter of culture)
- Simple post-translational modifications
- Secretion of the protein of interest

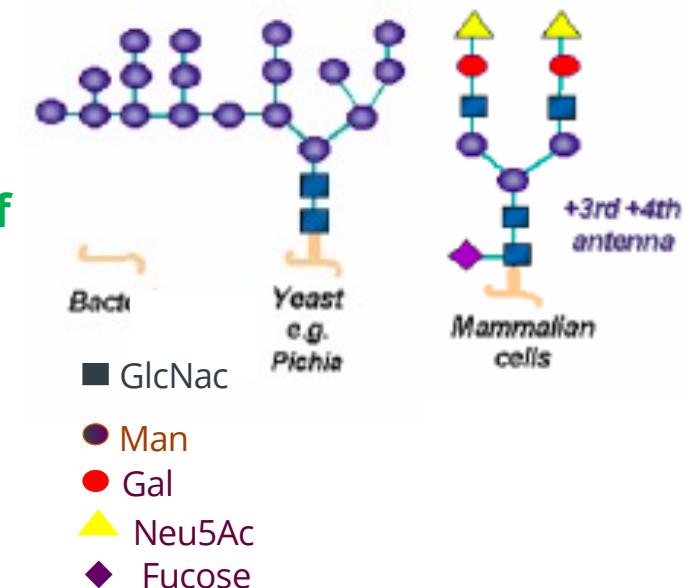
Proteins produced in yeast

Insulin  
GM-CSF  
Vaccine HBV  
Vaccine HPV  
Glucagon

## INCONVENIENTS

- Hypermannosylations
- Folding

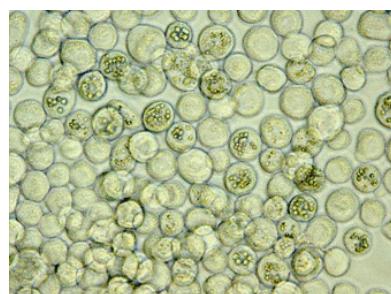
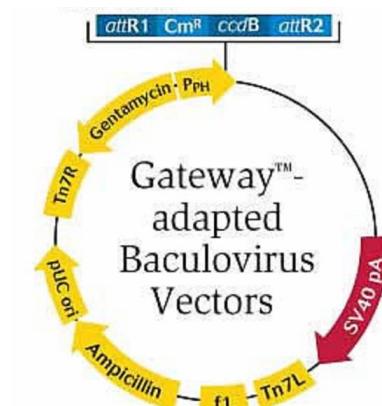
Glyco-engineering for humanization



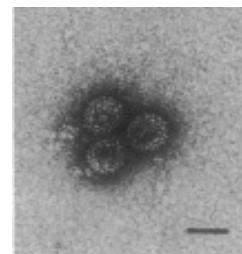
# Protein production in insect cells

## Baculovirus: insect cell virus

- Use of recombinant baculovirus (bearing the cDNA sequence of interest) to infect insect cell suspension
- Protein synthesis



➤ **CERVARIX® (EMA, 2007 and FDA, 2009) bivalent vaccine bivalent against papillomavirus: prévention of cervical cancer**  
HPV-16 L1 protein + HPV-18 L1 protein



**Papillomavirus L1 major capsid protein self-assembles into virus-like particles that are highly immunogenic**

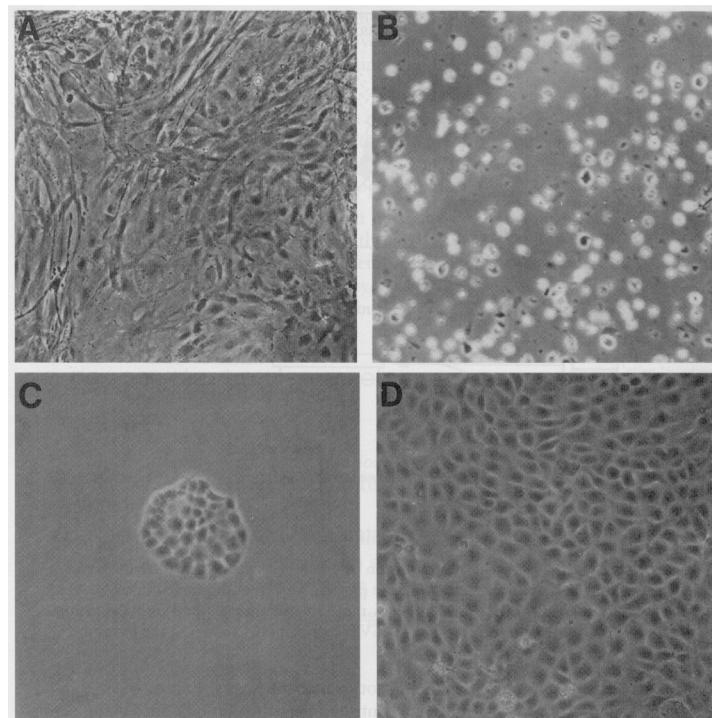
R. KIRNBAUER\*, F. BOYD†, N. CHENG†, D. R. LOWY\*, AND J. T. SCHILLER\*

*Proc. Natl. Acad. Sci. USA, 1992*

➤ **PROVENGE® (FDA, 2010)**  
Autologous product for prostate cancer cell therapy: PSA produced in *S. frugiperda*

➤ **FluBlok® (FDA, 2011)**  
Recombinant influenza vaccine → Time and cost-reduced compared to egg production protocol

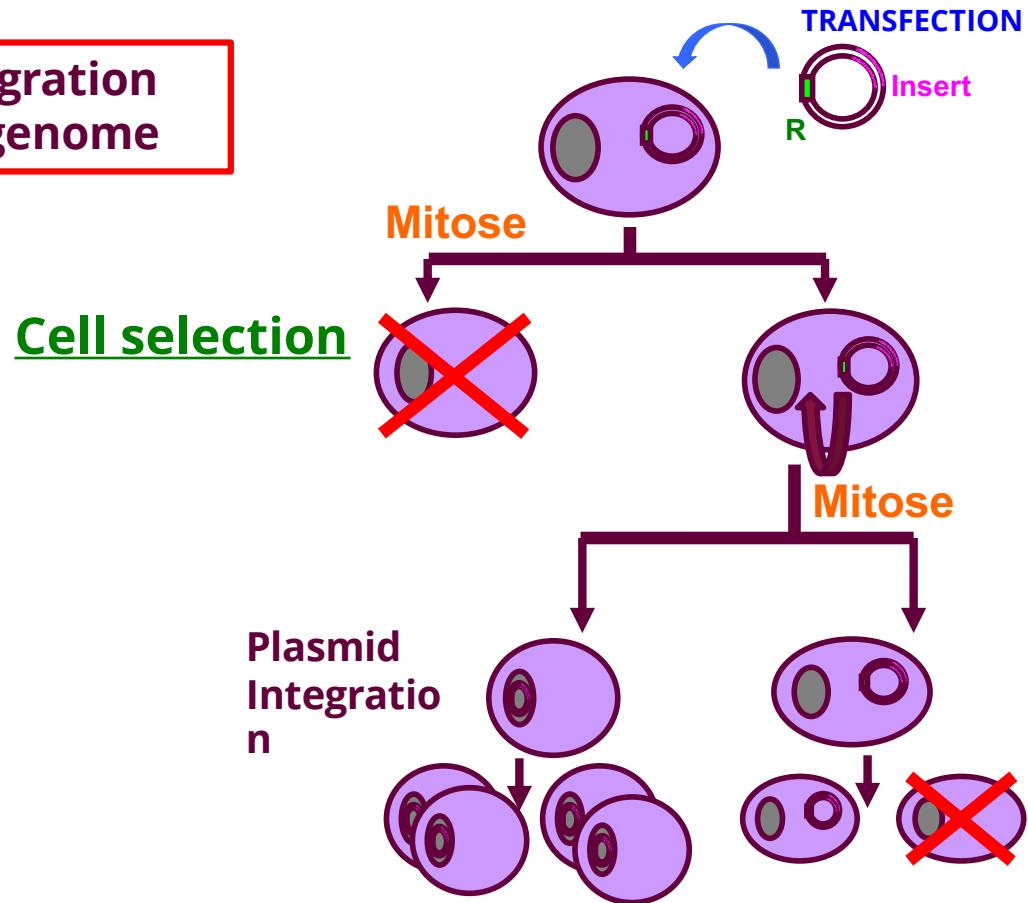
# Protein production in mammalian cells



A Non transfected cells

B-D Transfected cells with selection pressure

Vector integration in the cell genome



→ Stable cell line for protein production

Secretion of the protein

Tight quality control of the cell-line

No integration: plasmid loss

# Protein production in mammalian cells

ECACC : European Collection of Cell Culture [www.ecacc.org.uk](http://www.ecacc.org.uk)

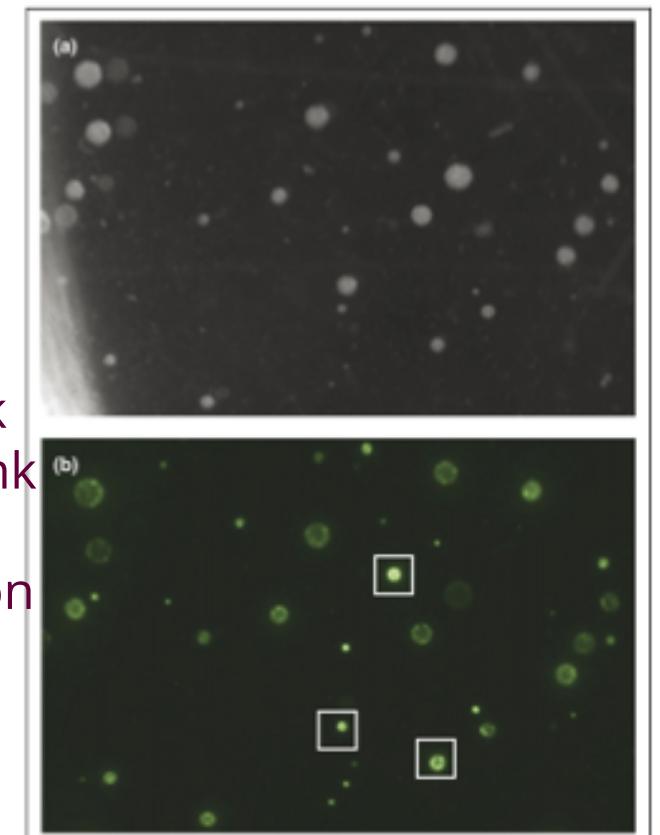
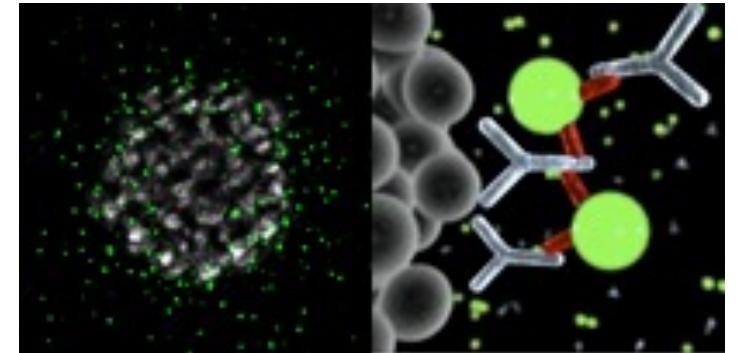
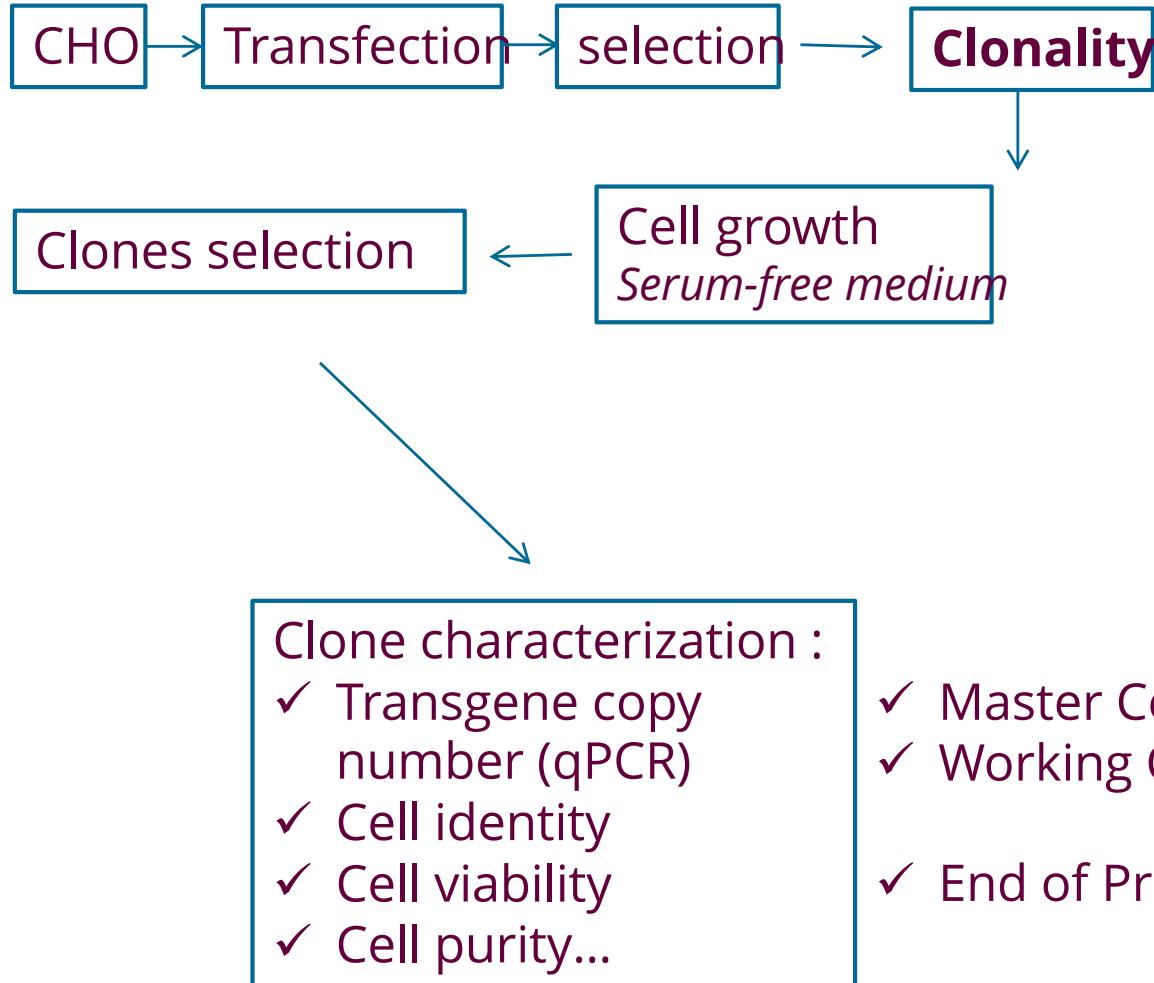
Cell	Biological source	Some common uses
CHO	Chinese Hamster Ovary cells	Recombinant proteins
VERO	Kidney epithelial cells extracted from an African green monkey	Human and veterinary viral vaccines (rotavirus, rabies, poliovirus)
BHK	Baby Hamster Kidney fibroblasts	Factor VIII, veterinary viral vaccines
HEK 293	Human embryonic kidney	Adenoviral vectors (SARS-CoV2)
Hybridomas	Murine hybrid cell line	Monoclonal antibodies
PER.C6	Derived from human embryonic retinal cells	Recombinant proteins, adenoviral vectors (SARS-CoV2)
NS0	Derived from the non-secreting murine myeloma	Recombinant proteins
MDCK	Madin-Darby canine kidney cells	Viral vaccines (flu)
MRC5	Human fetal lung fibroblast cells	Human viral vaccines (flu)



<https://www.mabdesign.fr/wp-content/uploads/2021/07/Immunowatch-Edition-3-Bioproduction-2.pdf>

# Best producing clones selection

<http://www.genetix.com>



# Quality control of the producing cell cultures, an example

The first recombinant human coagulation factor VIII of human origin: human cell line and manufacturing characteristics

Casademunt et al., European Journal of Haematology. 2012; 89 (165-176)

Table 2 Overview of tests performed and results obtained

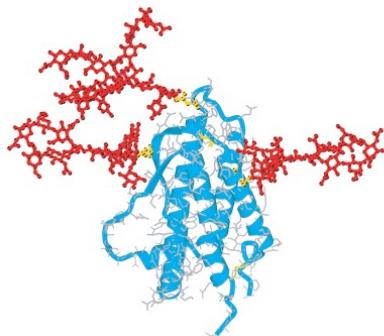
	MCB	WCB	End of production cells	Bulk harvest
Identity				
Isoenzyme analysis	HEK 293	–	–	–
RAPD	HEK 293	HEK 293	HEK 293	–
Microbiological tests				
Sterility	Complies	Complies	Complies	Complies
Mycoplasma	Negative	Negative	Negative	Negative
Cell identity				
Microbiological tests				
Viral tests				
<i>In vitro</i> assay for the detection of adventitious viruses (28 d, three detector cell lines: Vero, MRC5 and 293)	Negative	–	Negative	Negative
<i>In vivo</i> assay for the detection of adventitious viruses (adult and suckling mice, embryonated eggs)	Negative	–	Negative	Negative
QF-PERT	Negative	–	Negative	Negative
PCR screen for human viruses (HIV 1/2, HTLV 1/2, CMV, EBV, HHV 6/7/8, HBV, HCV, B19, HPV, HPyV)	Negative	–	–	–
PCR screen for AAV-2	Negative	Negative	Negative	Negative
TEM	No virus detected	–	No virus detected	No virus detected
MMV infectivity assay	Negative	–	–	–
<i>In vitro</i> bovine virus screen (BVDV, BAV, BRSV, BPV, REO3, BTV, RV)	Negative	–	–	–
PCR screen for bovine polyoma virus	Negative	–	–	–
<i>In vitro</i> porcine virus screen (PPV, PAV, TGE, HEV)	Negative	–	–	–

MCB, master cell bank; WCB, working cell bank; EPC, end of production cells; BH, bulk harvest; RAPD, random amplified polymorphic DNA; QF-PERT, quantitative fluorescent product-enhanced reverse transcriptase; PCR, polymerase chain reaction; HEK, human embryonic kidney; HIV, human immunodeficiency virus; HTLV, human T-cell lymphotropic virus; CMV, cytomegalovirus; EBV, Epstein-Barr virus; HHV, human herpes virus; HBV, hepatitis B virus; HCV, hepatitis C virus; B19, human parvovirus B19; HPV, human papilloma virus; HPyV, human polyoma viruses JC and BK; AAV, adeno-associated virus; TEM, transmission electron microscopy; MMV, mouse minute virus; BVDV, bovine viral diarrhoea virus; BAV, bovine adenovirus; BRSV, bovine respiratory syncytial virus; BPV, bovine parvovirus; REO, reovirus; BTV, bluetongue virus; RV, rabies virus; PPV, porcine parvovirus; PAV, porcine adenovirus; TGE, transmissible gastroenteritis virus; HEV, haemagglutinating encephalitis virus.

– indicates tests have not been performed.

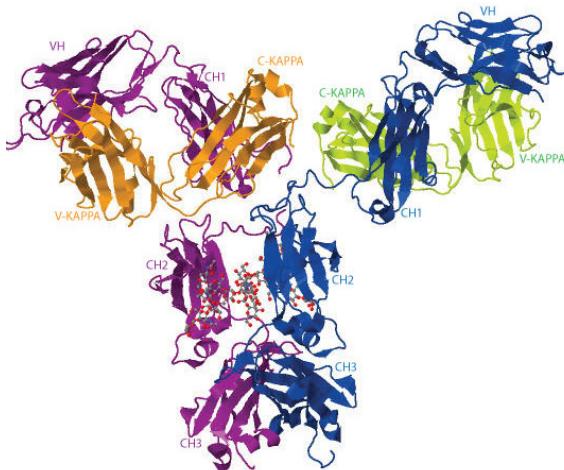
Viral security

# A few proteins produced in mammalian cell lines



## Erythropoïétin (EPO)

165 aa, MM : 30,6 kDa ; 40% glycosylation



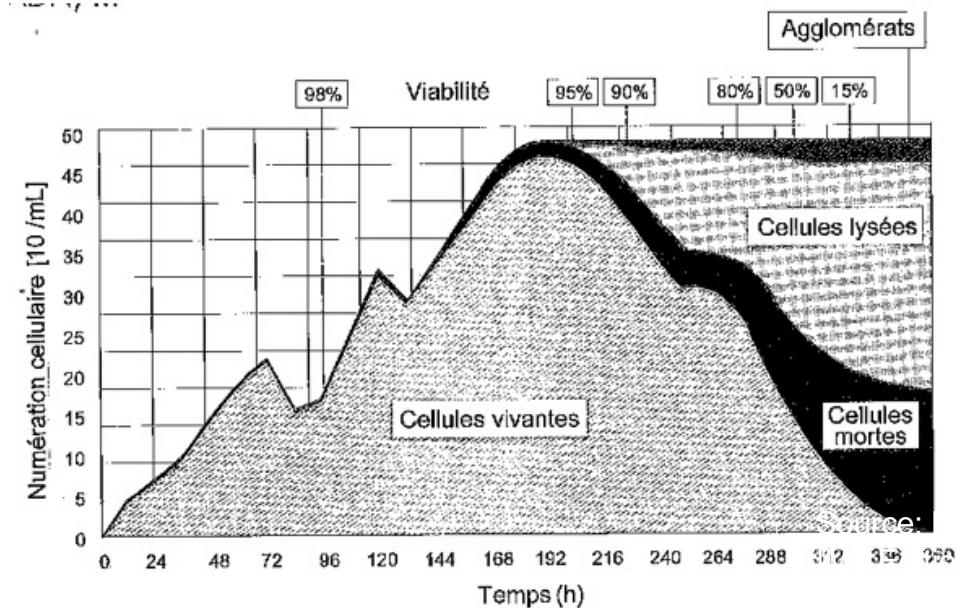
**Full antibodies (mAb): 1200 aa  
MM:150kDa, 1 glycosylation site**

- Full antibodies and complex proteins
- Optimal glycosylation
- Full functional activity
- Optimized expression vectors

- Cost of the culture media
- Process lenght: cell line establishment, cell banks...
- Weak yields
- Viral security

# Downstream process: What substances to eliminate during the purification steps?

- ✓ Host impurities:  
Cell debris, nucleic acids, lipids  
Host-cell proteins (HCP)
  
- ✓ Microorganisms:  
Viral particles, Bactéria,  
Pyrogen substances
  
- ✓ Process-related impurities:  
Remaining buffer, chromatography résin, Métals, polymers  
«extractibles and leachables »...
  
- ✓ Product-related impurities:  
Aggregated, truncated, unfolded...protein of interest forms



# Industrial production of monoclonal antibodies mAbs

28 days



USP

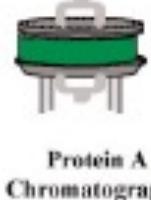
1 day

Cells removal



DSP

mAb Capture  
Most impurities  
removal



Intermediate:  
DNA, Host cell  
proteins,  
endotoxins  
removal

Cation Exchange  
Chromatography

Polishing: remove  
degraded mAbs

Anion Exchange  
Chromatography

10 days

Buffer  
change

Viral  
Filtration

UFDF

Product  
characterization  
10 WEEKS

Yield 70-80%

UNIVERSITÉ PARIS-SACLAY  
FACULTÉ DE PHARMACIE

Harvest  
(centrifugation and  
depth filtration)



Centrifuge



Filtration



# Therapeutic protein characterization, A battery of validation tests

## ▪ Identity / structure

- ✓ Primary structure (AA composition)
- ✓ Secondary structure
- ✓ Glycosylation analysis
- ✓ Physical parameters: Mol weight, isoelectric point...

## ▪ Purity:

- ✓ Host cell impurities (DNA, proteins, lipids...)
- ✓ Fabrication process impurities: leachates and extractables
- ✓ Product-related impurities: unfolded, truncated, aggregated, chemically degraded mAbs

## ▪ Activity:

- ✓ Target binding (affinity measurement)
- ✓ *In vitro* assays
- ✓ *In vivo* assays

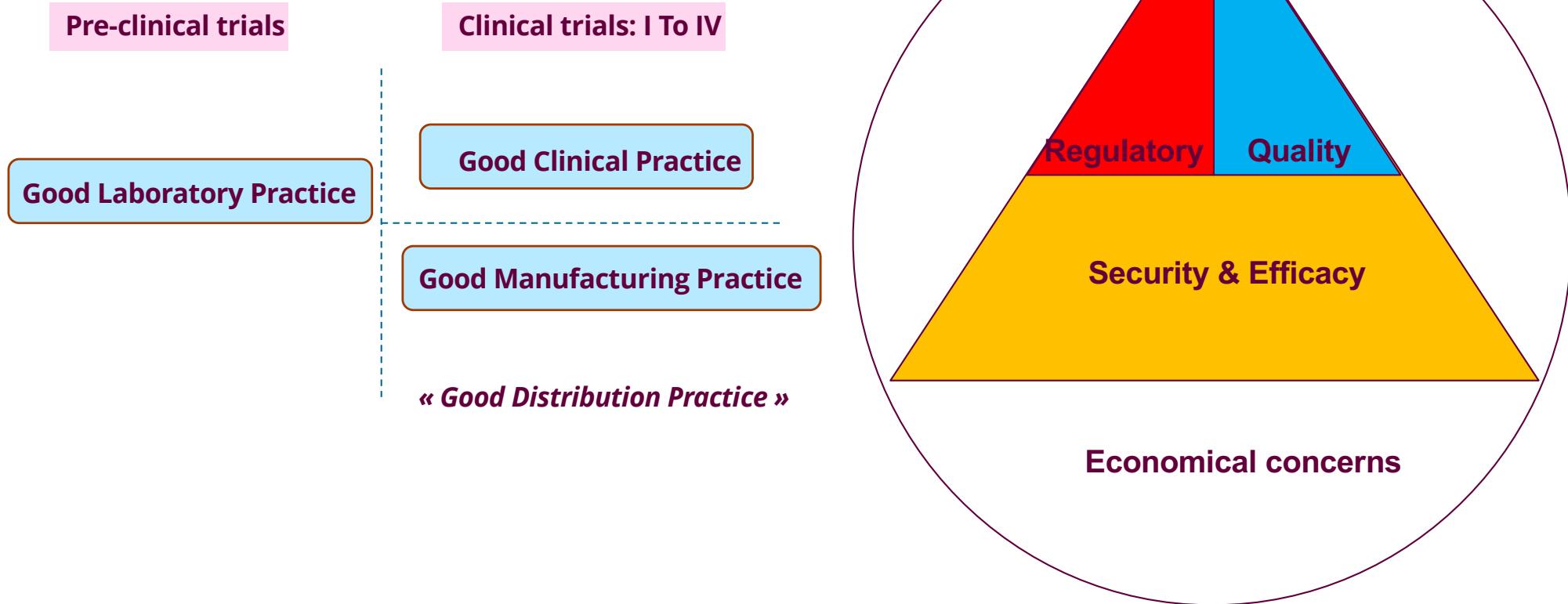
## ▪ Security: Virus particles or genome

Endotoxin detection

Which level?



# Conclusion



Bioproduct production is under pharmaceutical regulatory concepts that are adapted to living material use

Define critical points at early development stages (MCB conception):

Process complexity

Quality controls

Equipment costs, Staff costs, time consuming



« DEVELOPING A PROCESS  
WITH THE END IN MIND »