UNIVERSITE PARIS-SACLAY FACULTÉ DE PHARMACIE

FACULTÉ DE PHARMACIE DE CHÂTENAY-MALABRY Identification number :

Last NAME :

First Name :

# MASTER 1: Development of Drugs and Health Products TU09 Analytical sciences

## Exam (no documents allowed) February 4, 2021 from 13:30 to 15:30

## **INSTRUCTIONS ON HOW TO WRITE ANSWERS**

- FILL IN VERY CAREFULLY THE HEADINGS ON THIS FIRST PAGE WHICH ALLOWS YOUR COPY TO REMAIN ANONYMOUS.

- NO MENTION OR SIGN OF RECOGNITION MUST BE INDICATED ON THE FOLLOWING PAGES, UNDER PENALTY OF NULLITY

- CHECK THAT THIS BOOKLET IS NUMBERED FROM 1 TO 8

- NO COMPLAINT CONCERNING THIS BOOKLET WILL BE ACCEPTED AFTER THE FIRST QUARTER OF AN HOUR OF THE EVENT.

- DO NOT WRITE WITH PENCIL.

- ANY COMMUNICATION OR ATTEMPTED FRAUD WILL RESULT IN A ZERO SCORE ON THE TEST.

# A. SEPARATION TECHNIQUES (1 hour: 10 points/20)

<u>Attention</u>: Notation of multiple choices questions: For each question: No difference with the correct response=100% of the note; one difference=50%; two differences= 20%; three or more or no response=0%.

### **Electrophoresis**

1) What are the mode(s) of electrophoresis that allow(s) the separation of proteins by their sizes?

- $\Box$  A. Native PAGE
- □ B. SDS-PAGE
- $\Box$  C. Isoelectric focusing (IEF)
- $\Box$  D. 2D Electrophoresis

2) What are the mode(s) of electrophoresis that allow(s) the separation of proteins by their charges?

- $\Box$  A. Native PAGE
- □ B. SDS-PAGE
- $\Box$  C. Isoelectric focusing (IEF)
- $\Box$  D. 2D Electrophoresis

# 3) We use a zone capillary electrophoresis (CZE) setup with untreated fused silica capillaries. What are the parameters that influence the electroosmotic flow (EOF)?

- $\Box$  A. The charge of the capillary's internal surface
- $\Box$  B. The pH of the buffer (background electrolyte)
- $\Box$  C. The ionic strength of the buffer
- $\Box$  D. The viscosity of the buffer

### Protein analysis and analysis of drugs and macromolecules

#### 4) Critical aspects for protein analysis using size exclusion chromatography are:

- $\Box$  A. Define the permeation range of the column under defined conditions
- B. Avoid interactions protein stationary phase
- $\Box$  C. Form aggregates during the analysis
- $\Box$  D. Insure that interactions protein excipients occurred during the analysis

# 5) Which chromatography mode can be used for detecting charged variants induced by degradations (deamidation, oxidation) or glycosylation?

- $\Box$  A. Size exclusion chromatography
- $\Box$  B. Reverse phase chromatography
- $\Box$  C. Ion exchange chromatography
- D. Hydrophobic interaction chromatography
- E. Hydrophilic interaction chromatography

#### 6) Different approaches for chiral separations:

- □ A. Chromatographic enantiomer separations can be carried out directly using chiral selectors incorporated either in the stationary phase or mobile phase to form diastereomeric derivatives.
- □ B. Chromatographic enantiomer separations can be carried out indirectly by using chiral derivatization reagents to form diastereomeric derivatives
- $\Box$  C. Indirect and direct ways are possible in CE
- $\Box$  D. Only indirect way is possible in CE

#### 7) Enantiomer separation using cyclodextrins (CDs) in CE

- □ A. The chiral recognition mechanism is based on inclusion of a hydrophobic group of the analyte into the cavity of the CD
- B. The chiral recognition is based on the possibility to form hydrogen bonds between the hydroxyl groups at the mouth of the CD and polar substituents close to the chiral center of the analyte
- $\Box$  C. Only neutral substituted CDs can be used
- $\Box$  D.  $\gamma$ CD-derivatives are the most frequently used for molecules bearing aliphatic or substituted aromatic group.

#### **Chromatography exercise**

A mixture of salicylic acid and acetylsalicylic acid (standard solution (b)) was analyzed by Reversed Phase LC. In the same conditions a standard solution containing only salicylic acid (standard solution (a)) was analyzed.



Figure 1 : Compounds structure

The operating conditions are: *Column: Kromasil C18*; L=12,5 cm; *i.d.*=4,0 mm;  $dp = 5 \mu m$ ; Mobile phase: *methanol* / 0,1% *phosphoric acid* (50/50 v/v); *Flow rate:* 1,2 mL/min; *Detection:* 237 nm.

The obtained chromatograms are shown on figure 2 and the corresponding data on table 1.



	Standard (a)		Standard (b)			
	<b>t</b> <sub>R</sub> (min)	<b>ω</b> 0,5 (min)	Area	<b>t</b> <sub>R</sub> (min)	<b>ω</b> 0,5 (min)	Area
Acetylsalicylic acid	-	-	-	1,83	0,09	831,02
Salicylic acid	3,48	0,12	807,73	3,48	0,12	871,18

Table I
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 $t_R$  is the retention time and  $\omega_{0,5}$  is the width of the peak at half of its maximum height and Area the peak area.

Dead time, determined by injection of a non-retained compound, is  $t_M=0,75$ min.

1. Calculate, for standard (b) chromatogram, the retention factors (k) for the 2 compounds and as well as the selectivity ( $\alpha$ ) and resolution (R) between them.

- 2. Concentration of salicylic acid in standard solution (a) is 0,01g/L. Calculate salicylic acid concentration in standard solution (b).
- **3.** We increased the methanol percentage to 60%. How would this affect dead time and retention time?
- **4.** Would a gradient mode be useful for this chromatographic separation or an isocratic mode is sufficient? Explain.

List of equations:

$$k = \frac{t_{\rm R} - t_{\rm M}}{t_{\rm M}}$$

$$N = 16 \left(\frac{t_R}{\omega}\right)^2$$
 or  $N = 5,54 \left(\frac{t_R}{\omega_{0,5}}\right)^2$ 

$$R_{S} = 2 \frac{\left(t_{R(2)} - t_{R(1)}\right)}{\left(\omega_{(1)} + \omega_{(2)}\right)} \quad \text{or } R_{S} = 1,18 \frac{\left(t_{R(2)} - t_{R(1)}\right)}{\left(\omega_{0,5(1)} + \omega_{0,5(2)}\right)}$$

$$\Delta P = \frac{\eta \, u \, \Phi_r L}{d_p^2}$$

 $logk = logk_w - S\varphi$  (S=3 for methanol)

## B. SPECTRAL METHODS (45 min: 7.5 points /20)

#### True or false (5 points):

1.	Near infrared spectro on absorption. True	scopy is based on light diffusion while mid infrared spectroscopy is based false
2.	Only the vibrations in True	nducing changes in the dipolar moment can be detected in infrared false $\Box$
3.	The light absorption l aqueous solutions True	by water molecules limits the application of near infrared spectroscopy on false $\Box$
4.	The energy necessar vibrational transition.	y to pass an electronic level is $1/10$ lower than that necessary for a false $\Box$
5.	Beer's law can be spectroscopies True	applied in UV -visible, mid-infrared and near-infrared absorption false $\Box$
6.	In UV-Visible spectron True	oscopy, the transmittance is directly proportional to the concentration. false $\Box$
7.	The X-rays are made True □	of energy quanta called photons false

- 8. The wavelength of X rays is between  $10^{-8}$  to  $10^{-11}$  m True false 🗖
- 9. The energy of X-rays is much lower than infrared false 🗖 True 🛛
- 10. X-rays diffraction arises from constructive interferences between the radiation diffracted by the periodically arranged atoms of molecules composing matter false True 🗖
- 11. X-ray Diffraction is mainly devoted to determination of crystalline structures at micrometric scale

false 🗖 True 🛛

12. STM technique is based on tunneling effect that allows the passage of electrons between the tip and the surface, creating an electric current.

Гrue 🗖	false 🗖
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13. In mass spectrometry, the Maldi ionization is considered as a soft technique

True false	
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14. In mass spectrometry, different types of analyzers can be used, for example: Quadrupole, Time-of-Flight or ion trap
True □ false □

15. Electron ionization and chemical ionization can be used when the mass spectrometer is coupled to Gaz chromatography

True 🗖

false 🗖

#### **NMR Spectroscopy (2.5 points)**

#### All answers have to be properly justified

#### Exercice 1

1) In the **Table 1** are gathered several isotopes of chemical elements identified following the official nomenclature by the symbol  $_Z^A X$ , where A is the mass number of the chemical element X, and Z its number of protons. In the **Table 2**, are indicated different nuclear spin values.

Associate a nuclear spin	value found in	Table 2 to	each isotop	e of <b>T</b>	able 1	1

$14^{-1}$ $14^{-1}$ $3^{-1}$ $15^{-1}$	<sup>28</sup> 14Si	<sup>29</sup> <sub>14</sub> Si	<sup>6</sup> <sub>3</sub> Li	$^{31}_{15}P$
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**Table 1** : Several isotopes of chemical elements identified following the official nomenclature by the symbol  $\binom{A}{Z}X$ , where A is the mass number of the chemical element X, and Z its number of protons

0	1/2	1

Table 2 : Different nuclear spin values

**2**) Boron possess different isotopes, among which one of them has a nuclear spin number equal to 3/2.

Indicate for this isotope in the two following cases (denoted **a** and **b**), the number of nuclear energy levels and their relative position in term of energy:

a) absence of any homogeneous and static magnetic field

**b**) in the presence of an homogeneous and static magnetic field denoted  $B_0$ .

## Identification number:

# C. THERMAL ANALYSIS (15 min, 2.5 points/20)

**DSC** : High density polyethylene (HDPE) is analyzed by Differential Scanning Calorimetry (DSC).

A glass transition at -125°C and a melting peak at 135°C are highlighted.

- 1) Draw the thermogram corresponding to the first temperature rise program.
- 2) Are these transitions of the first or second order?
- 3) Are they reversible or irreversible?

TGA : 1) What is the main information given by TGA? Briefly recall the principle.

2) In order to identify the volatile compounds emitted during a TGA experiment, which couplings with other analytical techniques can be set up ?