

**université
PARIS-SACLAY**

**FACULTÉ DE
PHARMACIE**

**GENE THERAPY
NON VIRAL VECTORS**

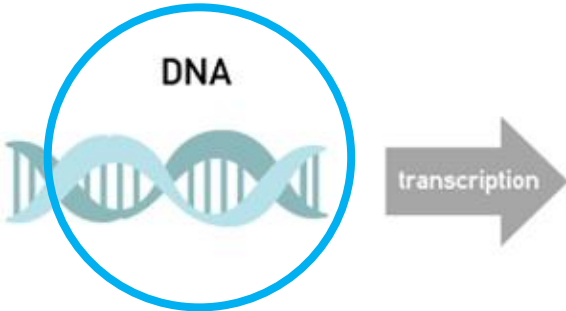
François Fay

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francois.fay@universite-paris-saclay.fr



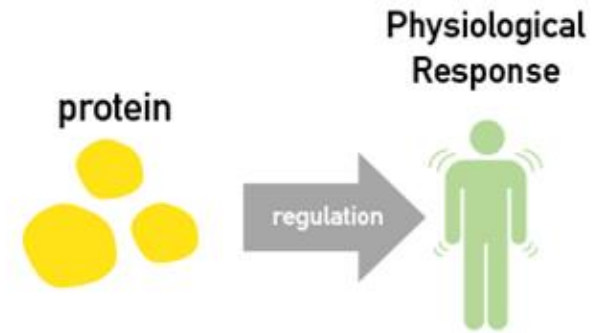
- Add DNA



- Add mRNA



- React with protein



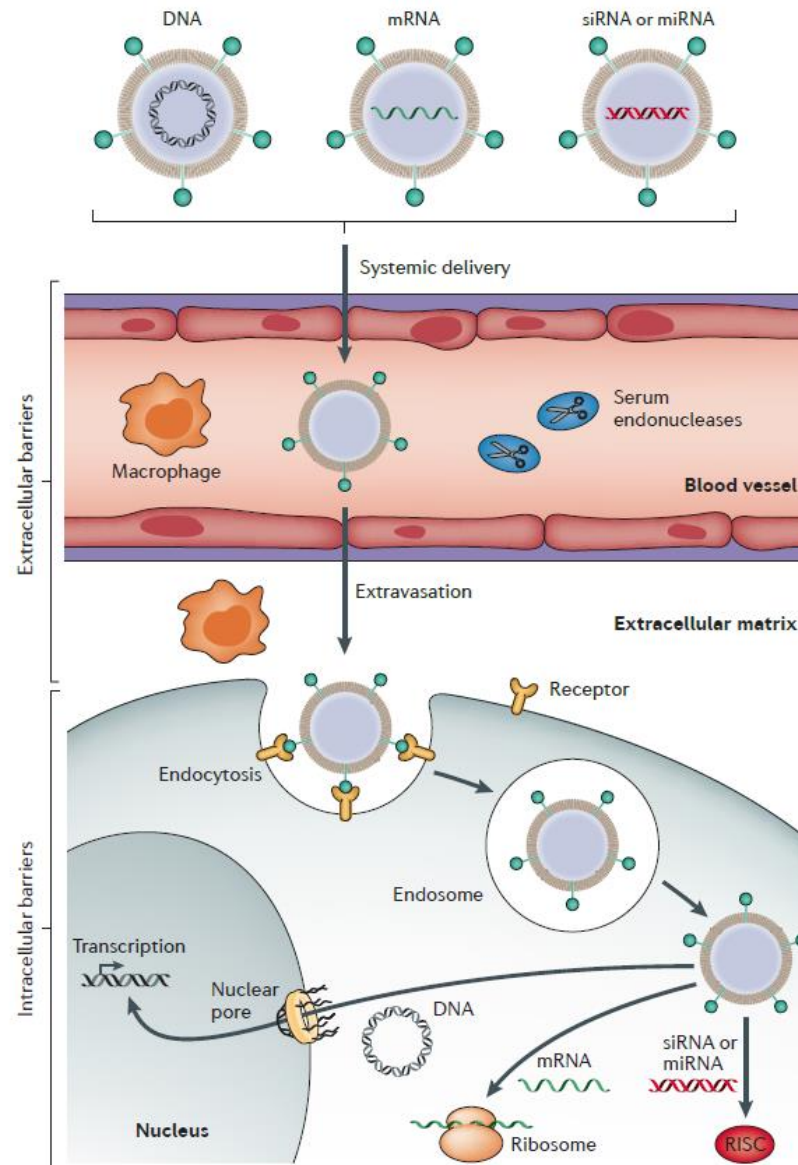
- Remove DNA
- Replace DNA

- Destroy mRNA (prevent translation)
- Modify mRNA (modify translation)

Natural barriers?

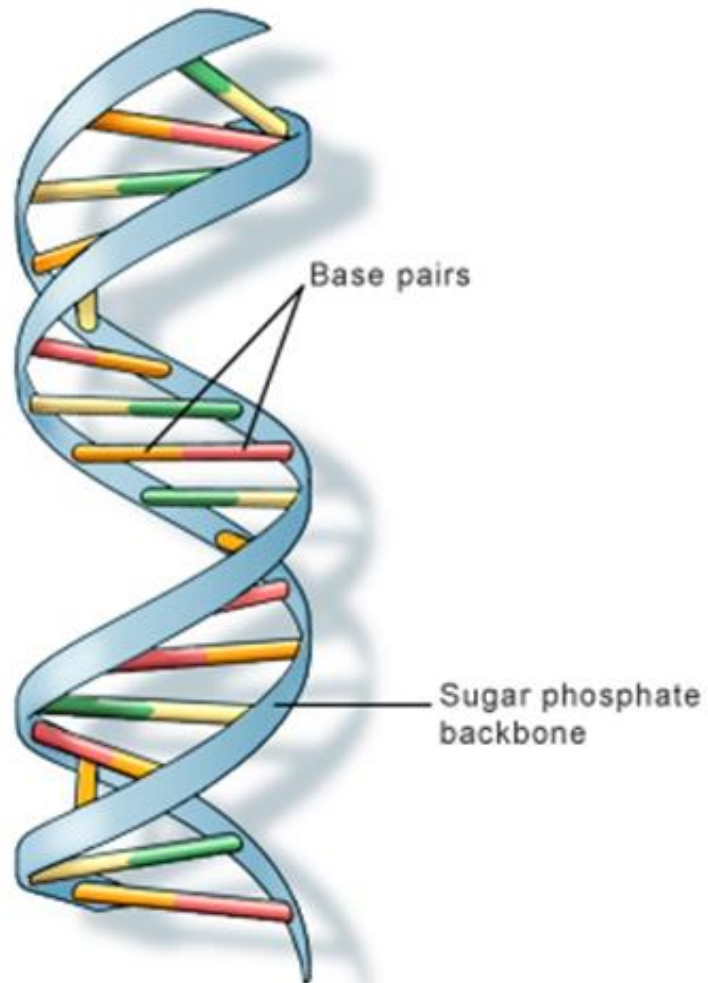
Natural barriers

Physiological barriers



Natural barriers

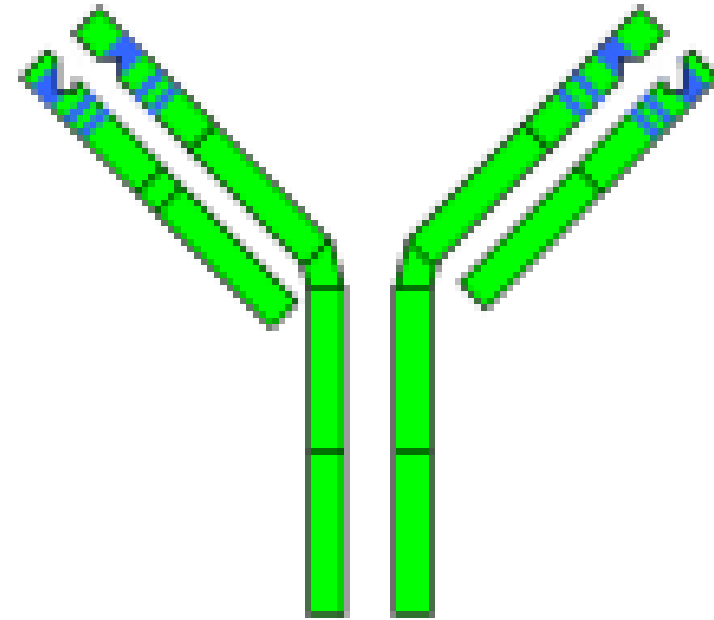
DNA / RNA Biochemistry



•

Aspirin 0,18 kda

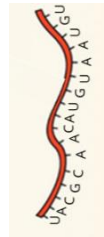
RNA ?



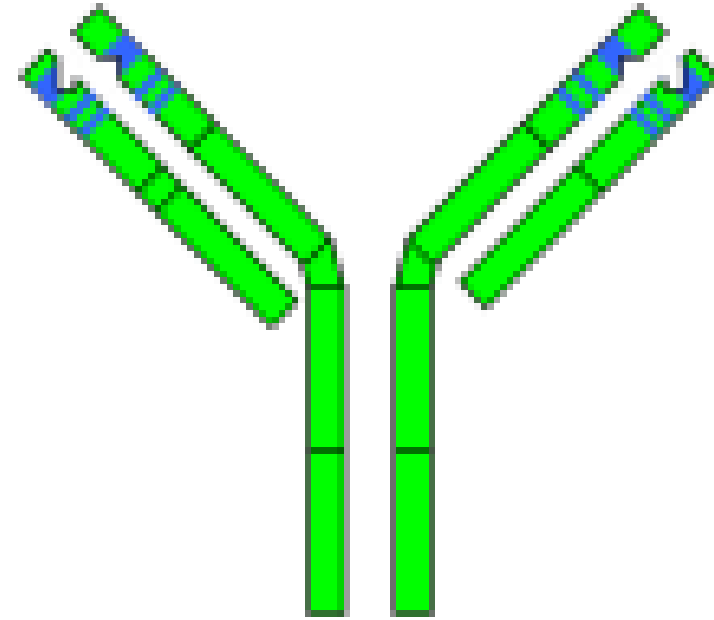
Antibodies 150 kda



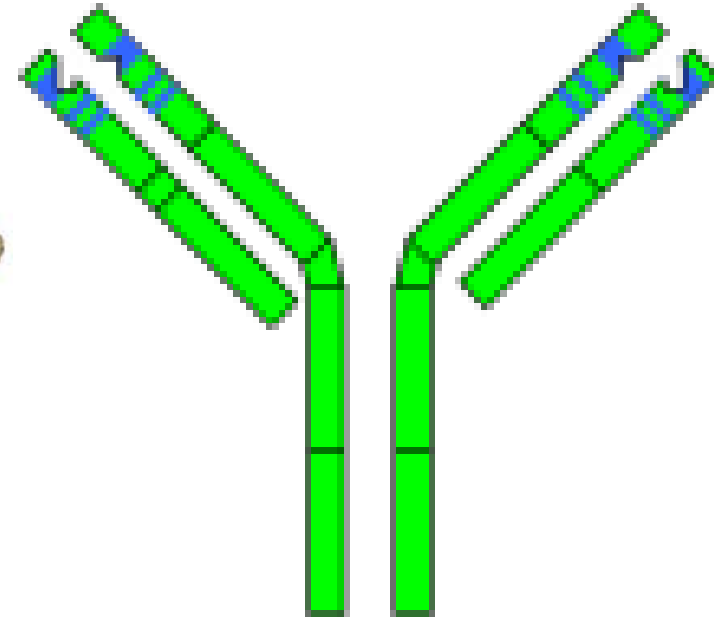
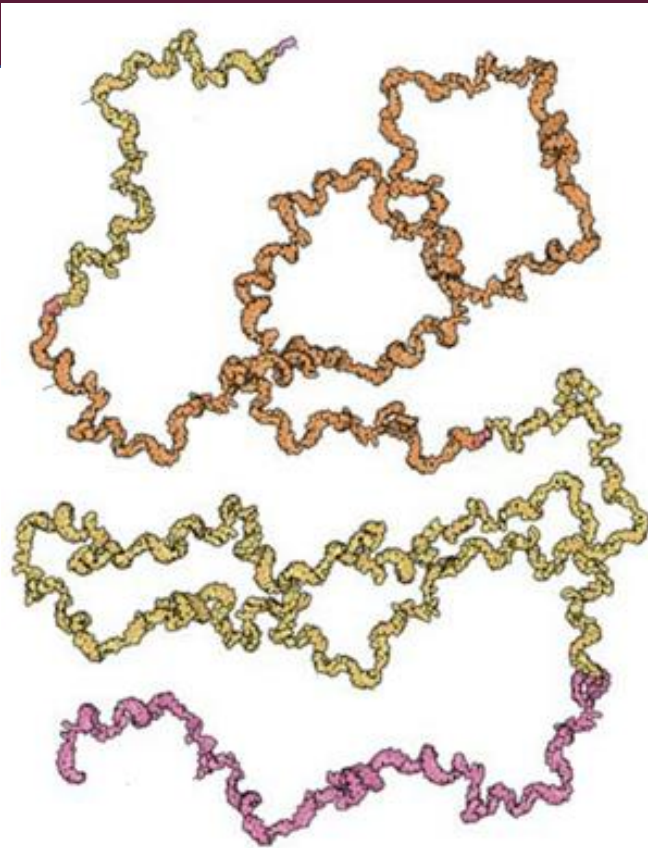
Aspirin 0,18 kDa



iRNA ~ 10 kDa



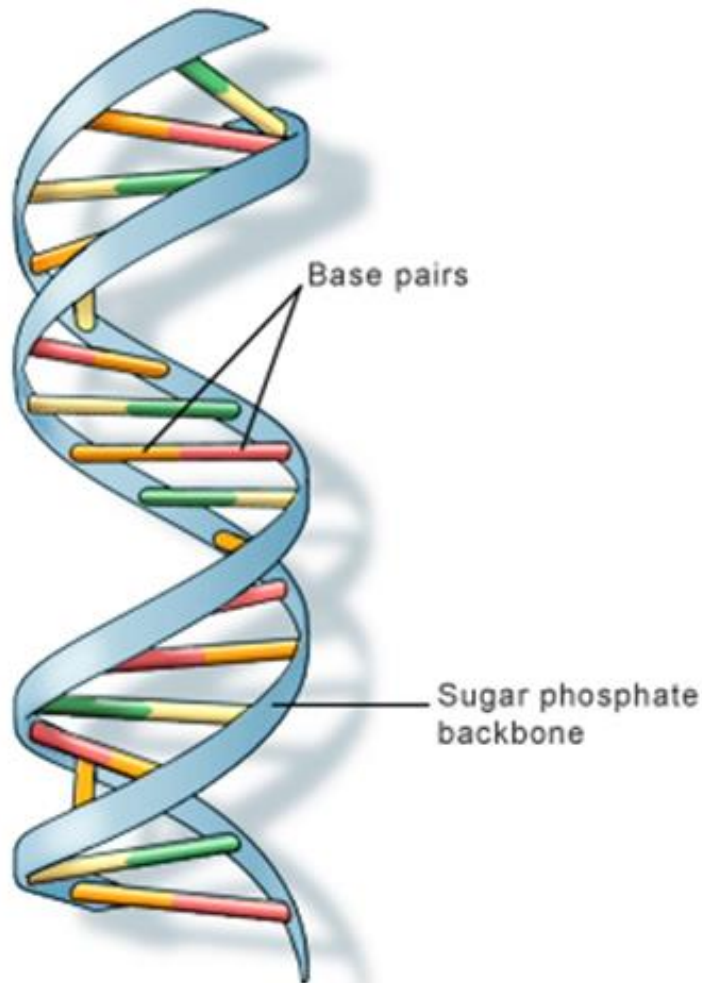
Antibodies 150 kDa



Aspirine 0,18 kda

mRNA ~400 kDa

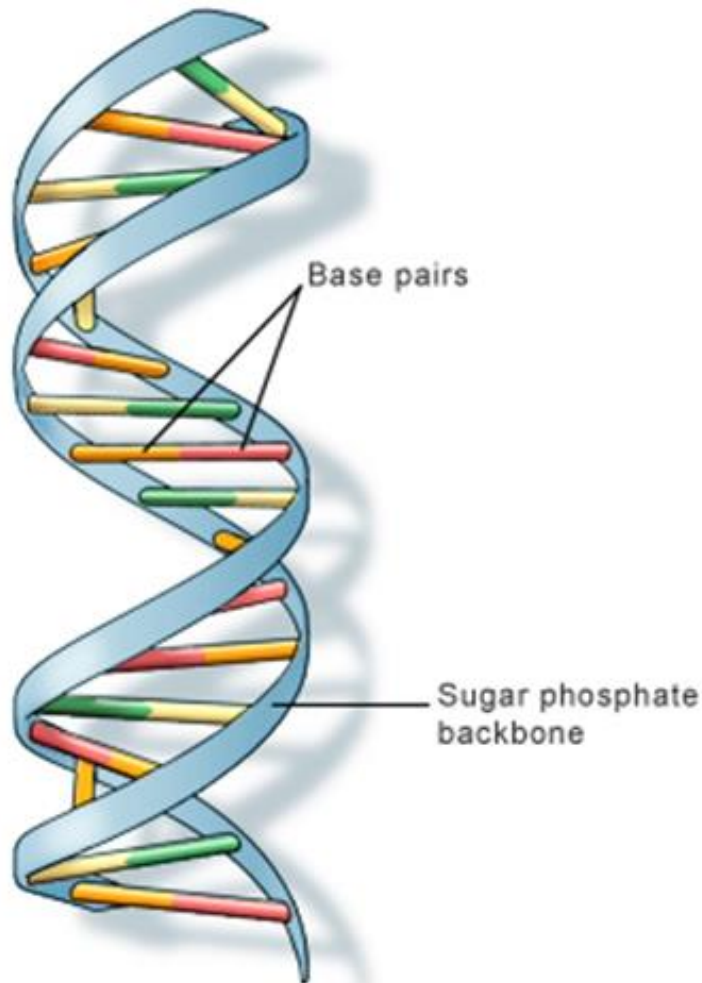
Antibodies 150 kda



U.S. National Library of Medicine



- Low stability
- High molecular weight
- Negative charge
- Hydrophilicity

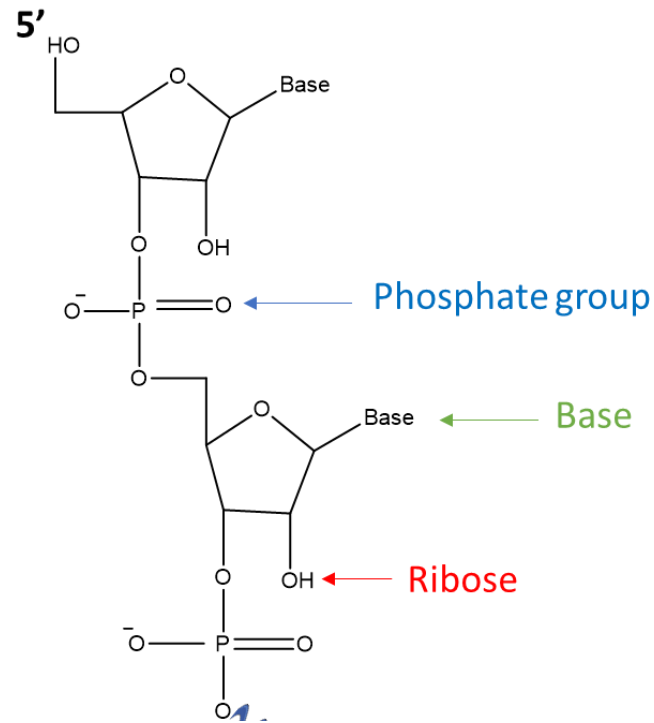


U.S. National Library of Medicine



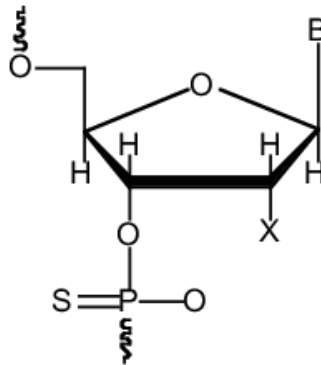
- Low stability
- High molecular weight
- Negative charge
- Hydrophilicity

→ **Need modification and/
or vectorisation!**

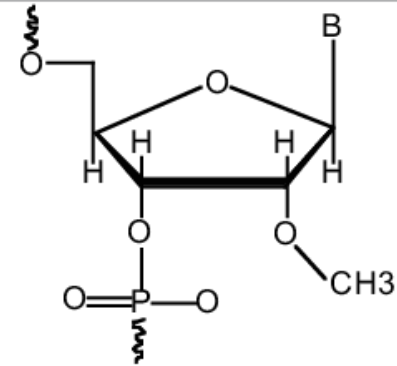


BACKBONE STRUCTURE

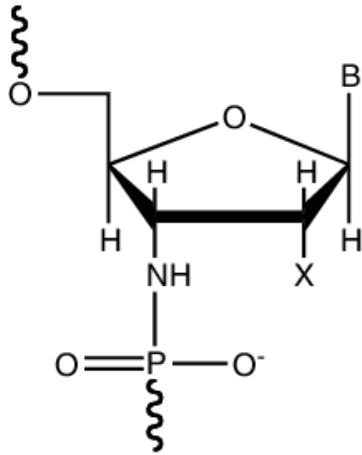
SUGAR RING



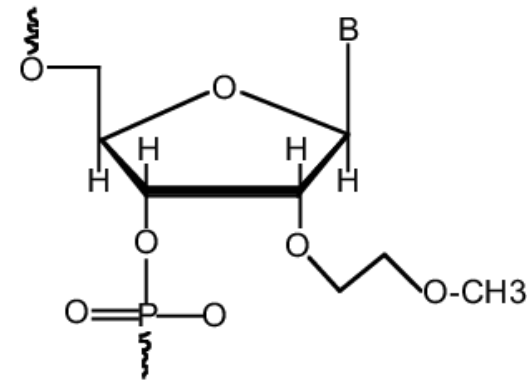
Phosphorothioate (PS)



2' *O*-Methyl (2' *O*-Me)



N'3 Phosphoramidate (NP)



2' *O*-Methoxyethyl (MOE)

Viral Vectors

Strengths

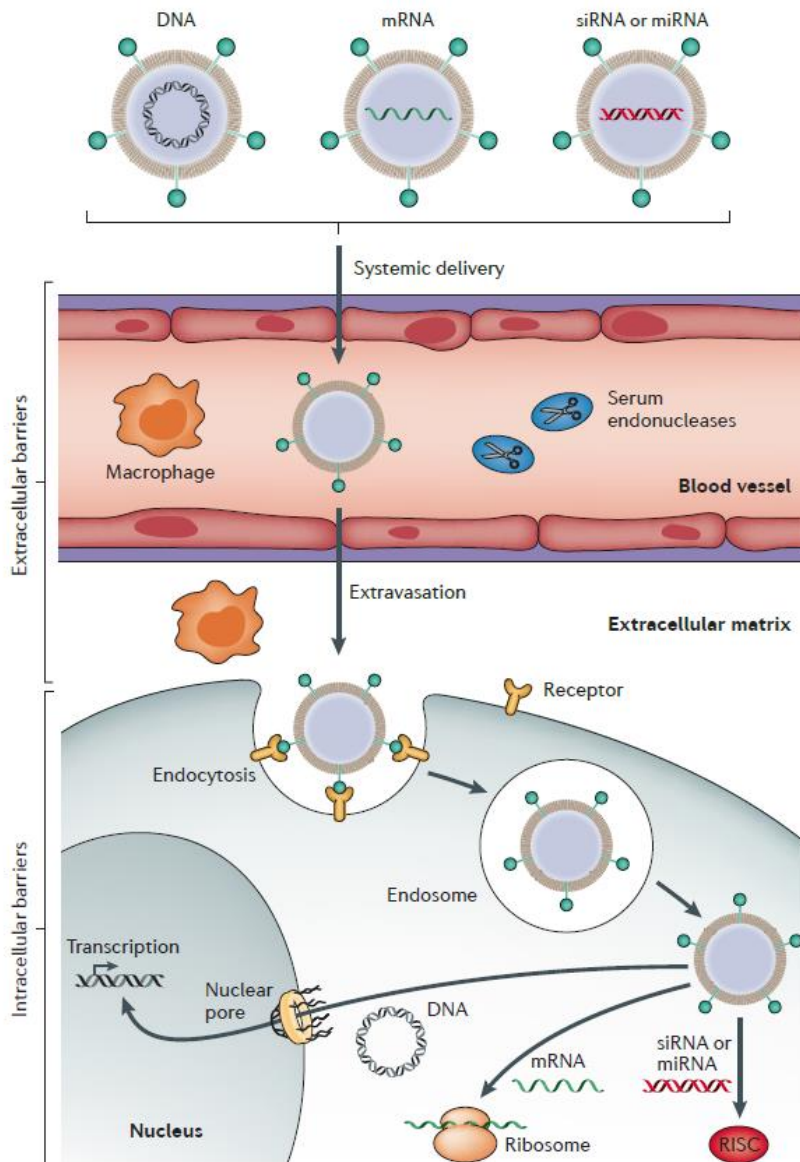
- High transfection efficiency
- Natural tropism (ability to infect different cells)
- Evolved mechanisms for endosomal escape
- Natural transportation mechanism of DNA into nucleus

Weaknesses

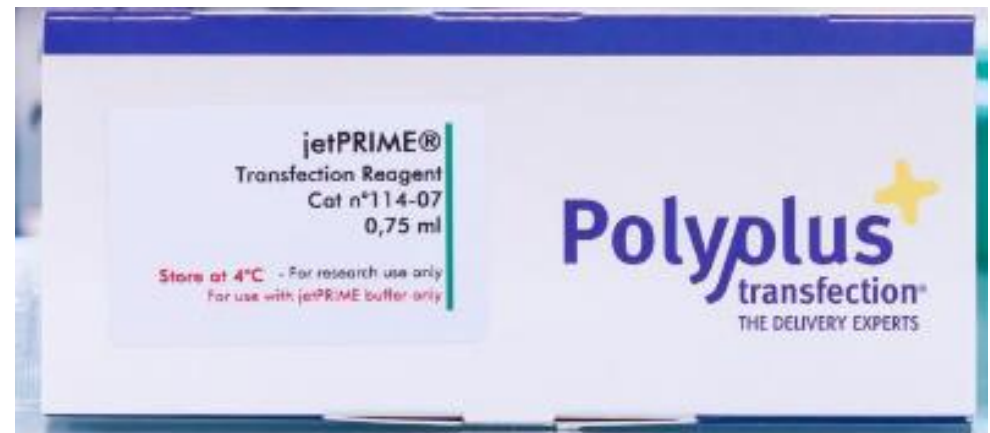
- Strong immune reactions against viral proteins prohibit multiple administrations
- Possibility of chromosomal insertion and protooncogene activation
- Complicated synthesis process
- Limitation on gene size
- Toxicity, contamination of live virus

Design criteria

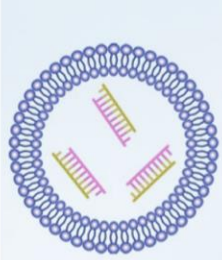
?



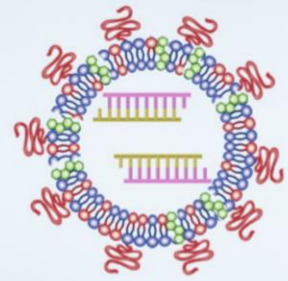
- Packaging of large DNA/RNA quantities
- Easy administration
- Serum stability
- Targetability to specific cell types
- Inexpensive synthesis
- Easy purification
- Robustness/stability
- Cell Internalization
- Endolysosomal escape
- Nuclear transport
- Efficient unpackaging
- Infection of non-dividing cells
- Safety
- Non-toxic
- Non-immunogenic
- Non-pathogenic



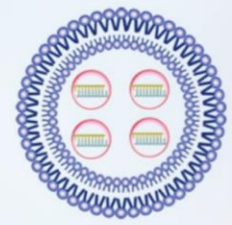
Lipid-based nanoparticles



Liposome



LNP

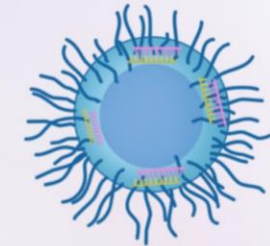


Lipopolyplex

Polymer-based nanoparticles



Polymeric NPs

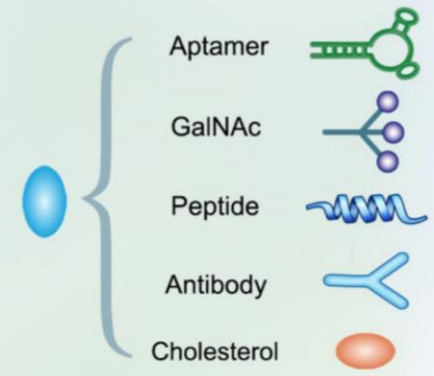


Polymeric micelles

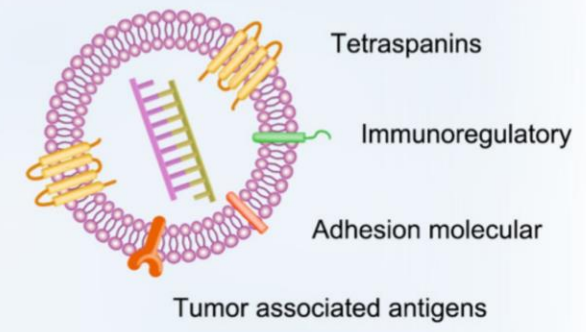


Dendriplex

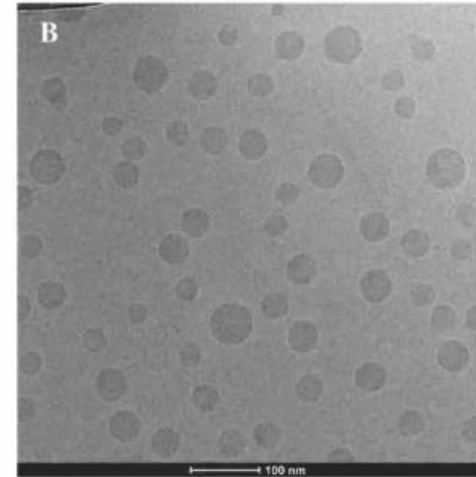
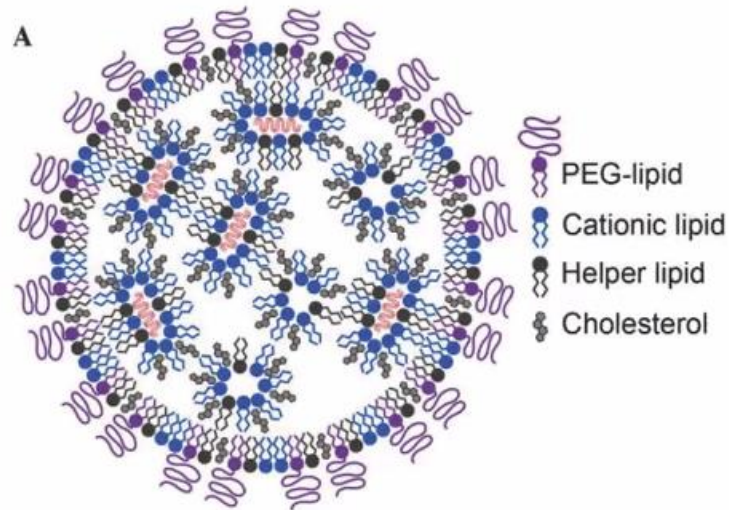
siRNA-ligand conjugates



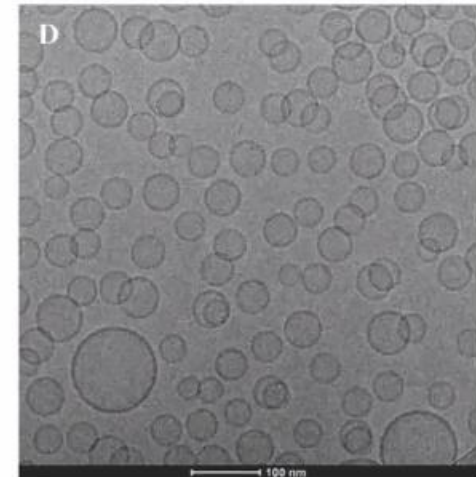
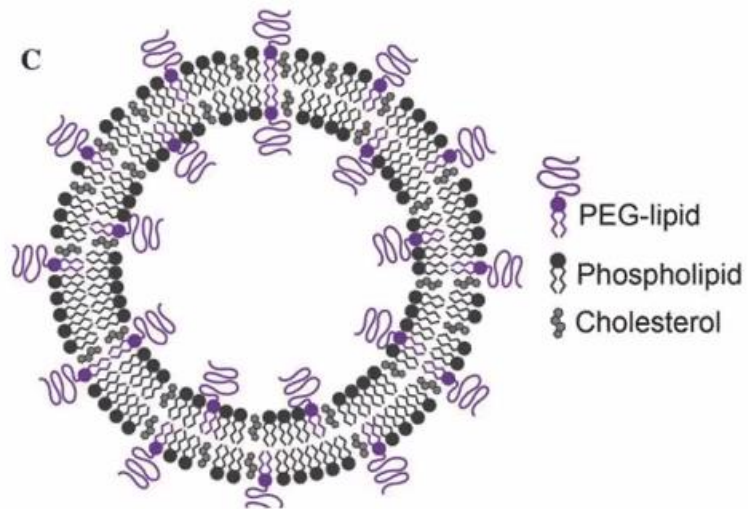
Exosomes



lipid nanoparticle



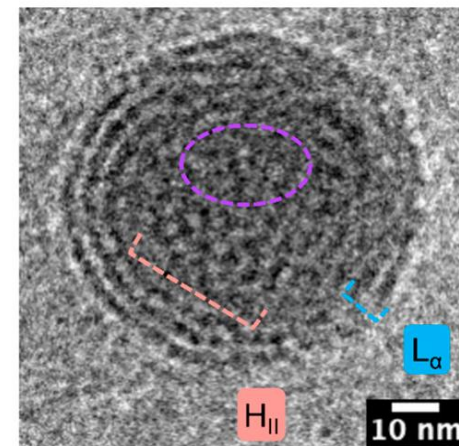
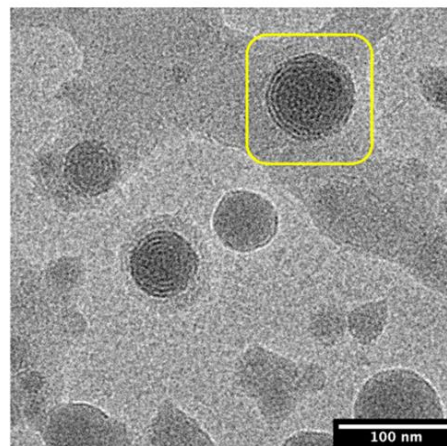
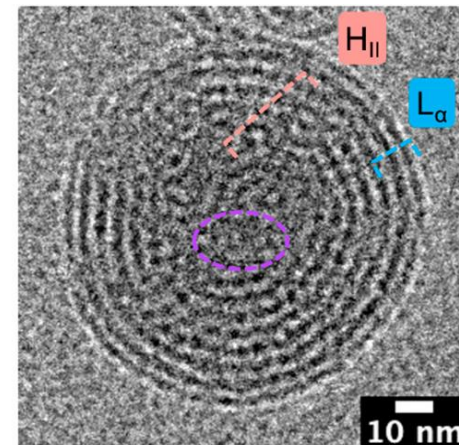
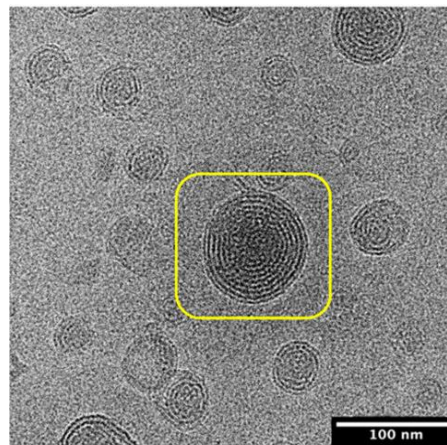
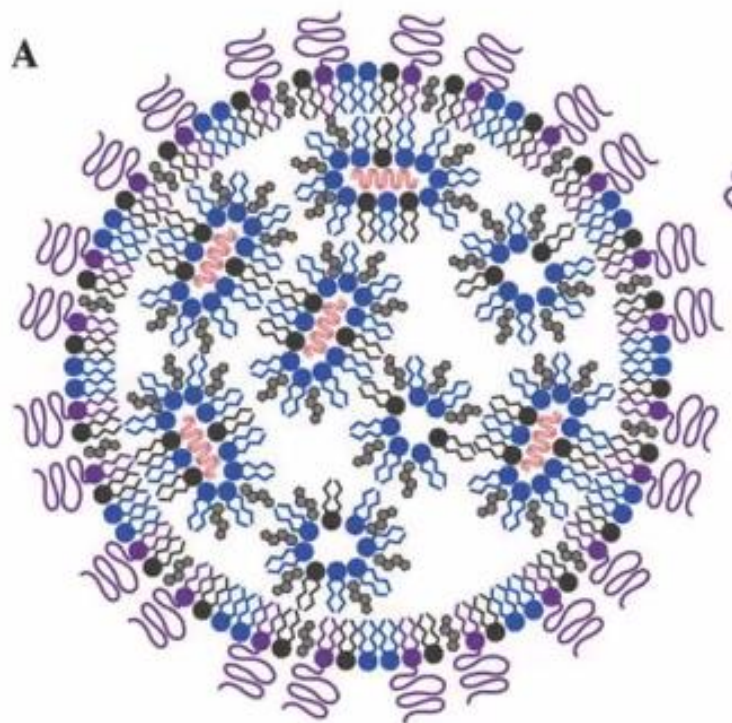
liposome

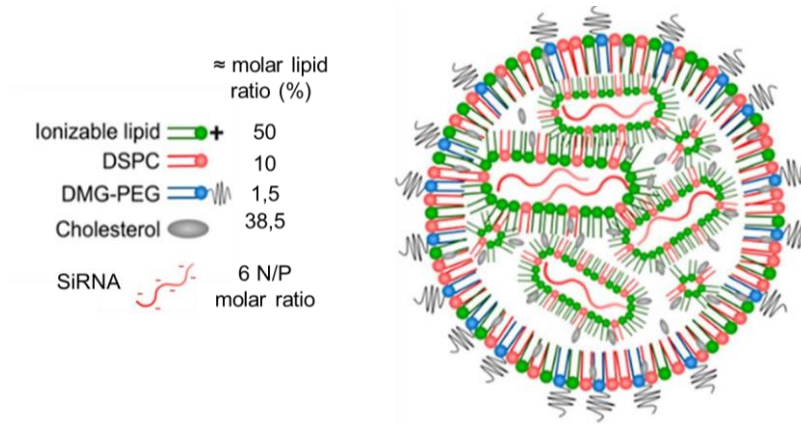


ACS Nano 2018, 12, 4787–4795

<http://nano.petr cigler.cz>

From a Presentation by Petr Cigler



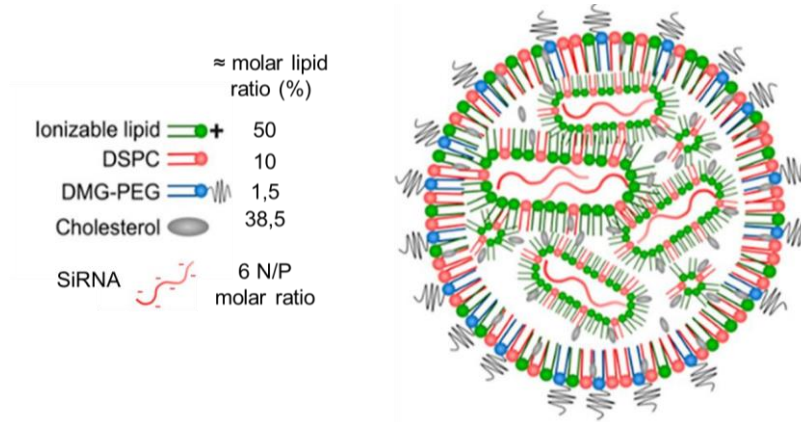



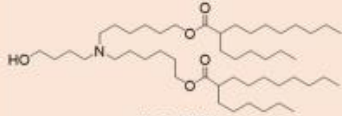
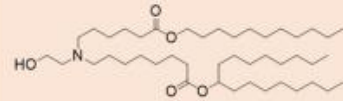
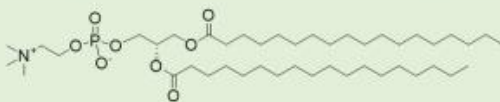
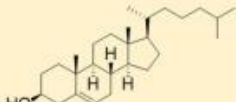
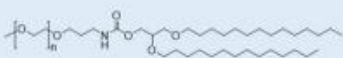

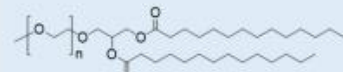
**Active ingredient
(Company)**

**Patisiran Onpattro
(Alnylam)**

**Tozinameran Comirnaty
(Pfizer/BioNTech)**

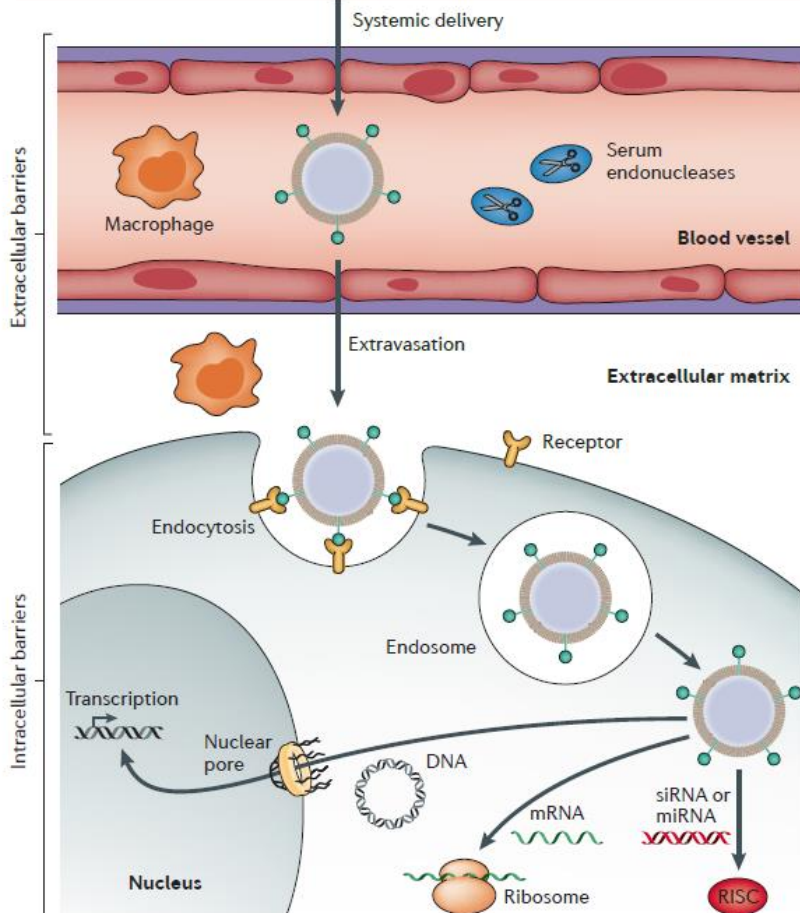
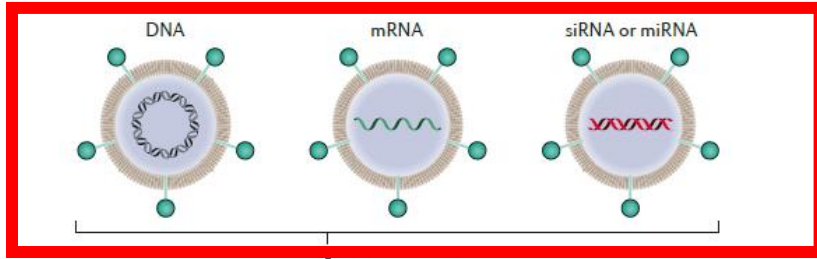
**Elasomeran Spikevax
(Moderna)**



Active ingredient (Company)	Patisiran Onpattro (Alnylam)	Tozinameran Comirnaty (Pfizer/BioNTech)	Elasomeran Spikevax (Moderna)
Ionizable lipid	 DLin-MC3-DMA	 ALC-0315	 SM-102
Phospholipid	 DSPC		
Sterol	 Cholesterol		
PEG-lipid	 PEG ₂₀₀₀ -C-DMG	 ALC-0159	 PEG ₂₀₀₀ -DMG

Natural barriers

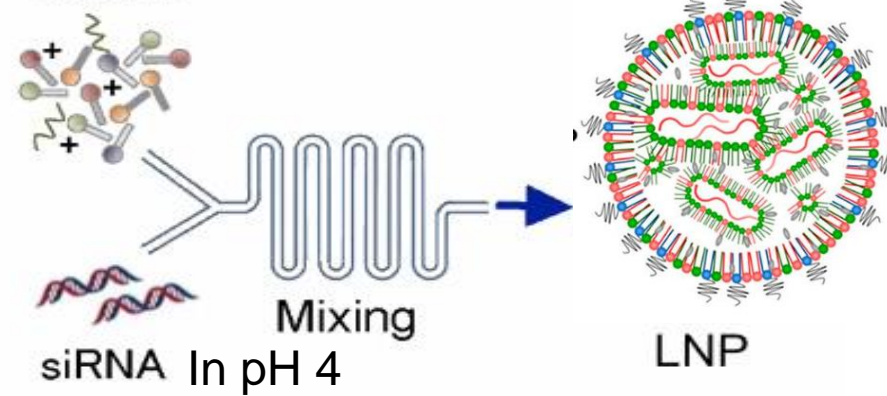
Physiological barriers



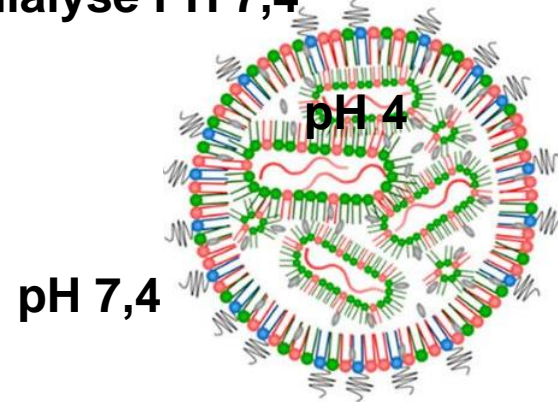
LNP

2) mixing

Lipids in
Ethanol

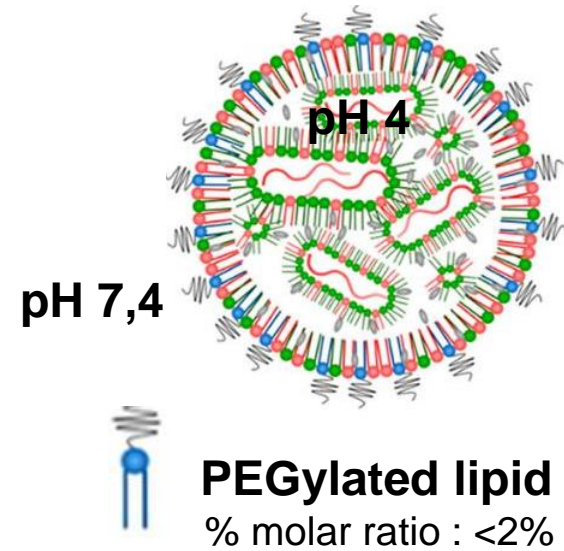
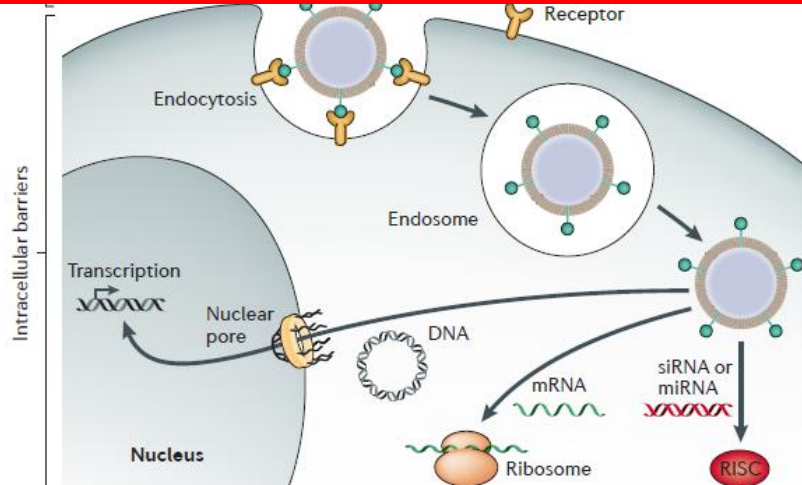
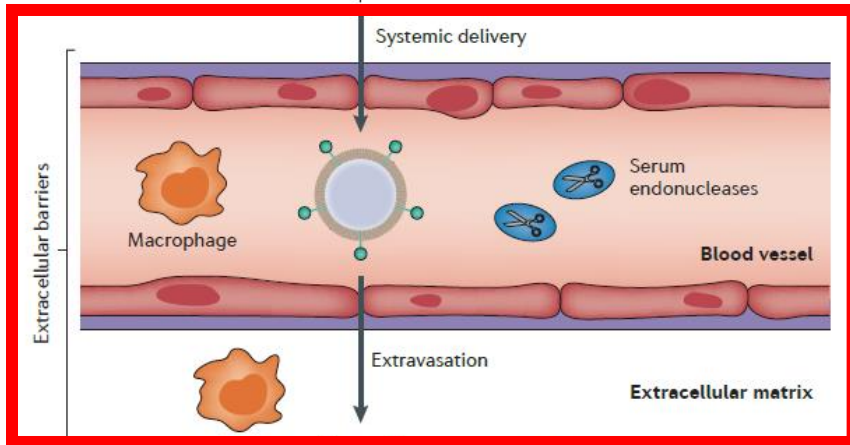
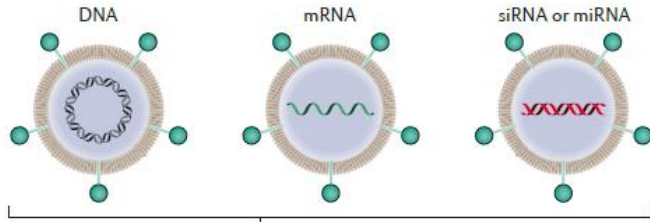


2) dialyse PH 7,4



Natural barriers

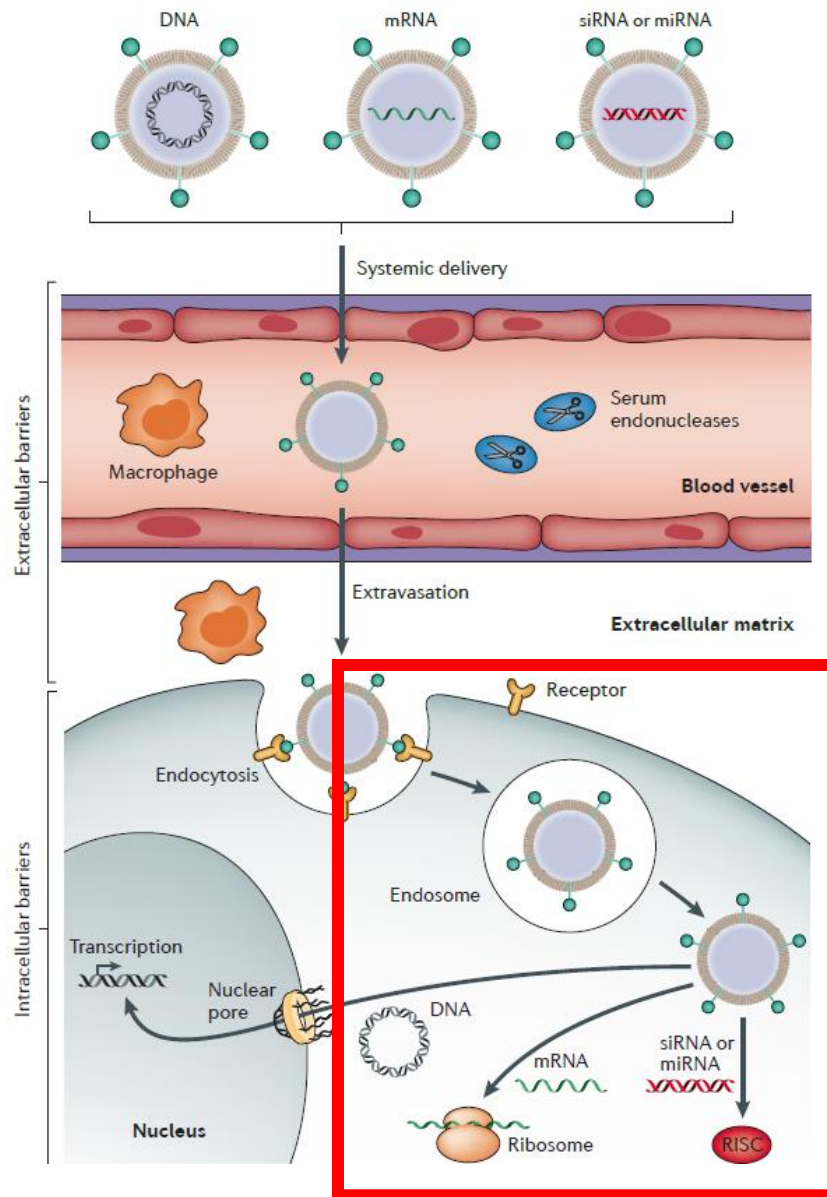
Physiological barriers



- Promotes colloidal stability
- Prevents protein aggregation
- Modulate size, transfection efficiency

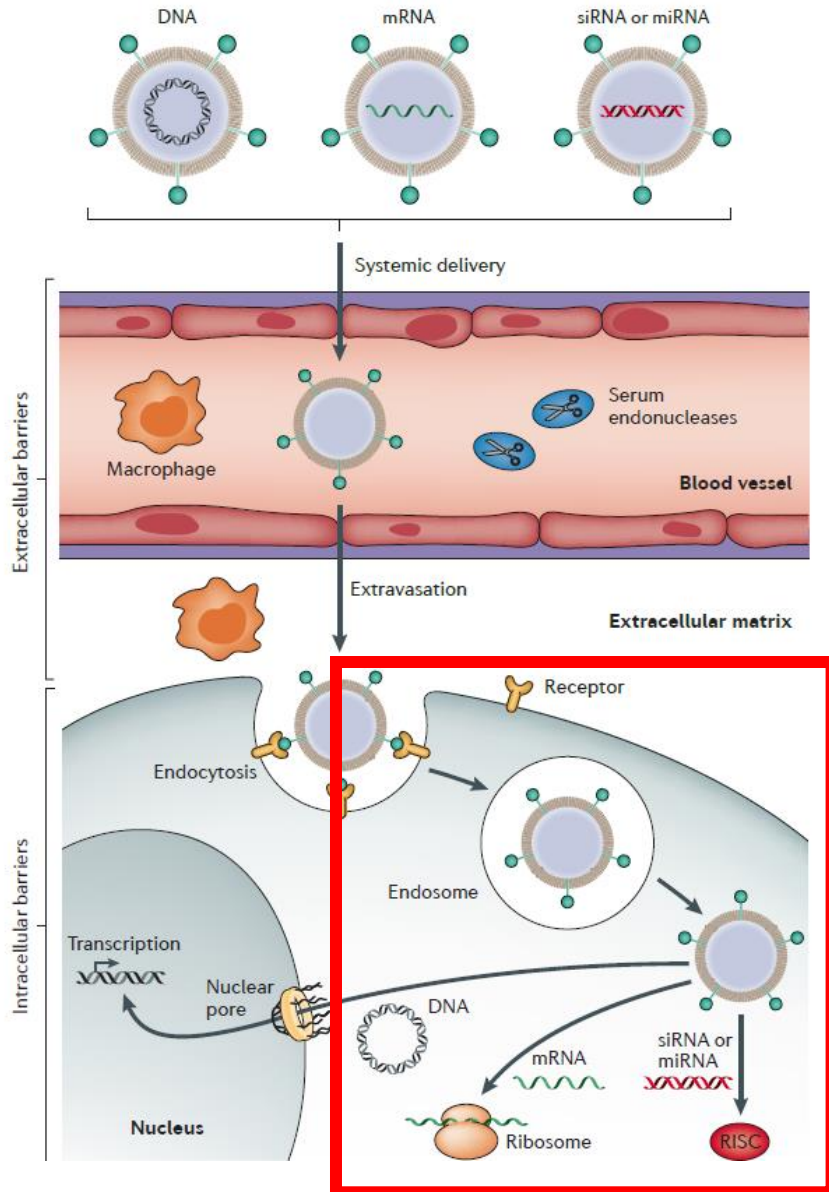
Natural barriers

Physiological barriers

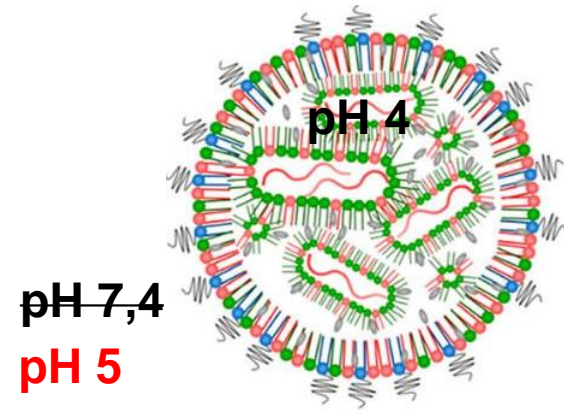


Natural barriers

Endosomal escape



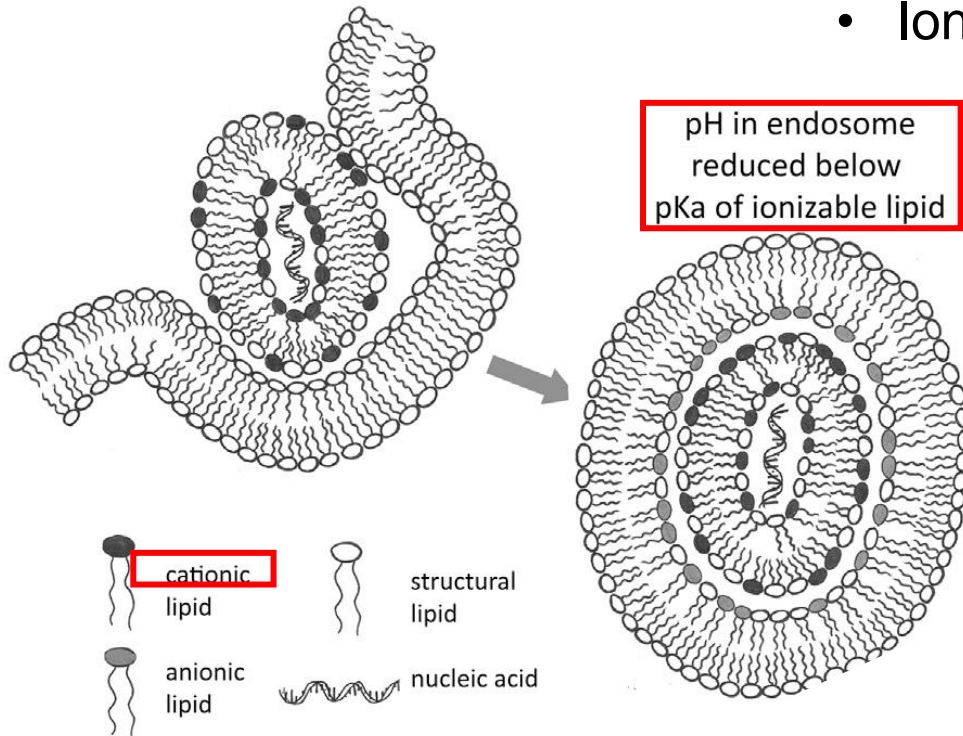
- Ionizable Cationic Lipids: pKa 6.2–6.5



Natural barriers

Endosomal escape

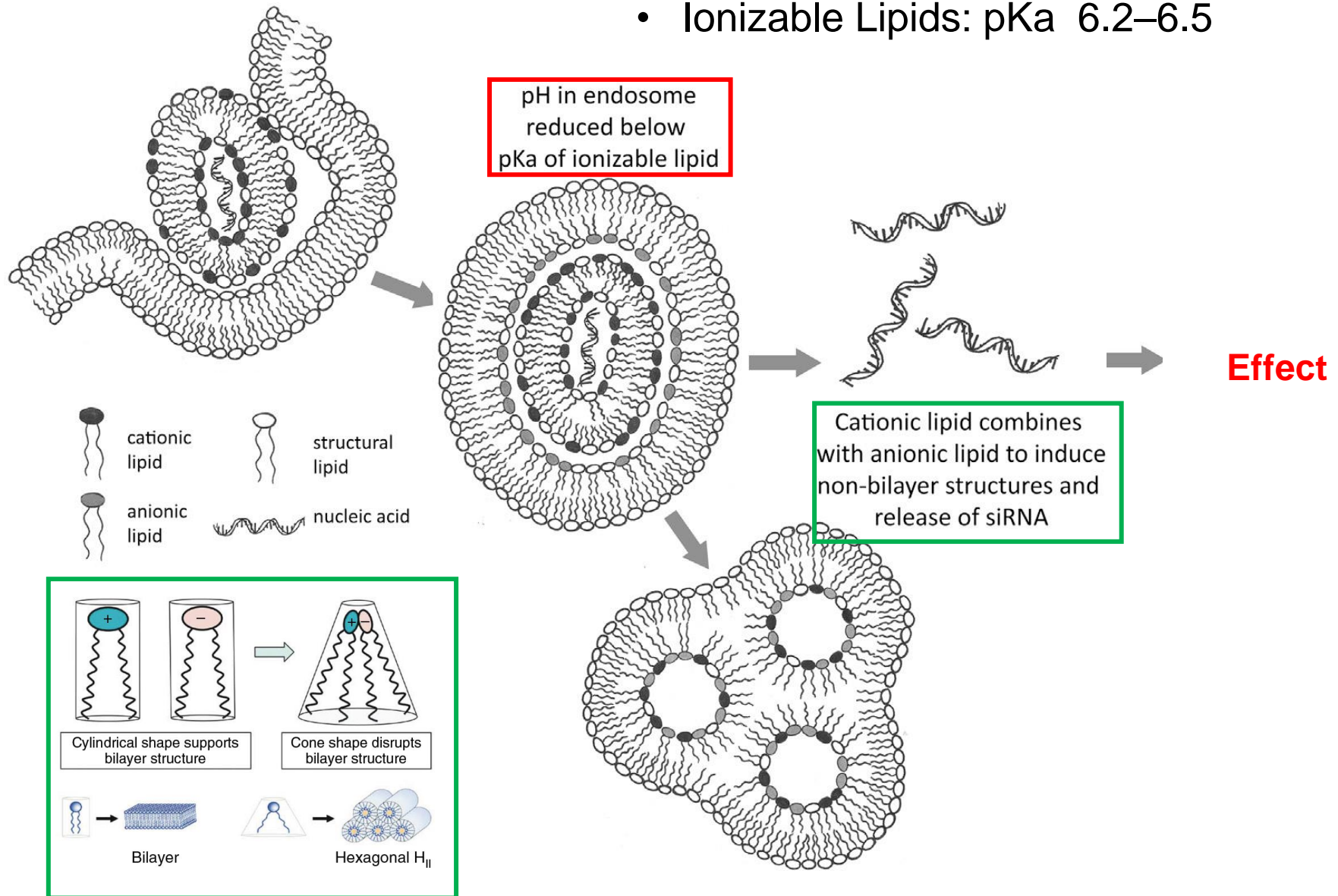
- Ionizable Lipids: pKa 6.2–6.5



Natural barriers

Endosomal escape

- Ionizable Lipids: pKa 6.2–6.5



Evolution of ionizable lipid during Patisiran development

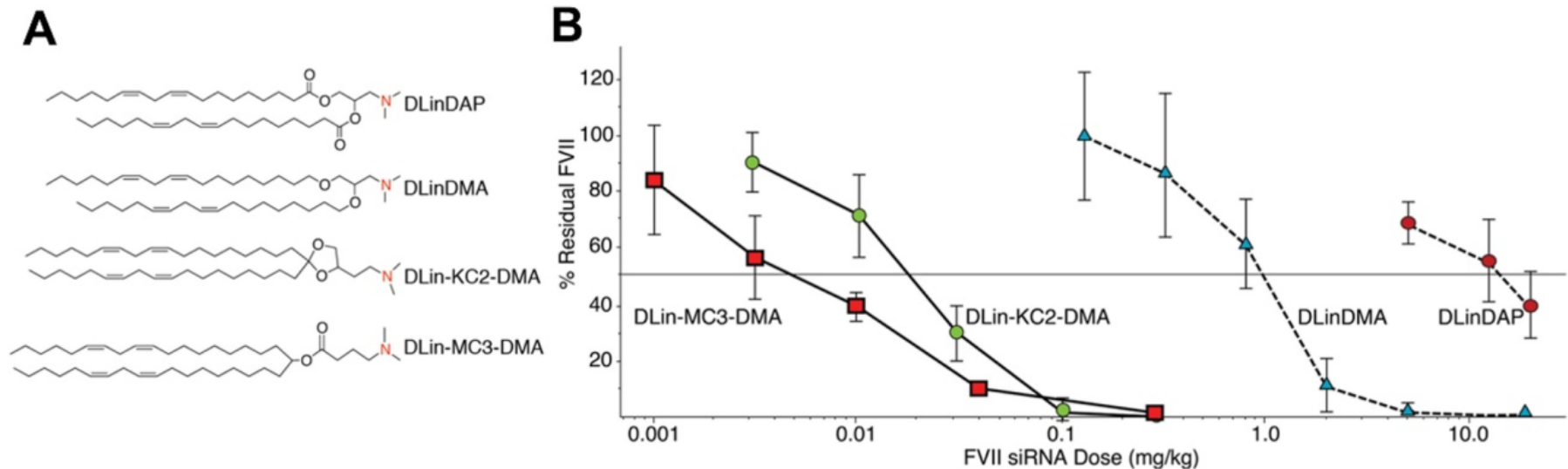


Figure 2. Evolution of ionizable cationic lipids enabling gene silencing *in vivo*. (A) DLinDMA, an ether-linked variant of DLinDAP, was the first lipid that enabled appreciable LNP-siRNA gene silencing activities *in vivo* following intravenous administration. Subsequent studies identified KC2 and MC3 lipids as potent successors with the optimal structure and pK_a necessary for siRNA delivery. (B) *In vivo* optimization of LNP-siRNA systems using the FVII mouse model. Over 300 species of ionizable cationic lipids were synthesized, formulated in LNP-siRNA, and screened in the FVII model. This led to the identification of the current “gold standard” lipid MC3, which resulted in improved LNP-siRNA potency by 4 orders of magnitude compared to DLinDAP, with no increase in toxicity. MC3 was subsequently employed in the patisiran formulation.^{4,5,17}

