



UMR-CNRS 8612 - Institut Galien Paris-Saclay

University Paris Saclay

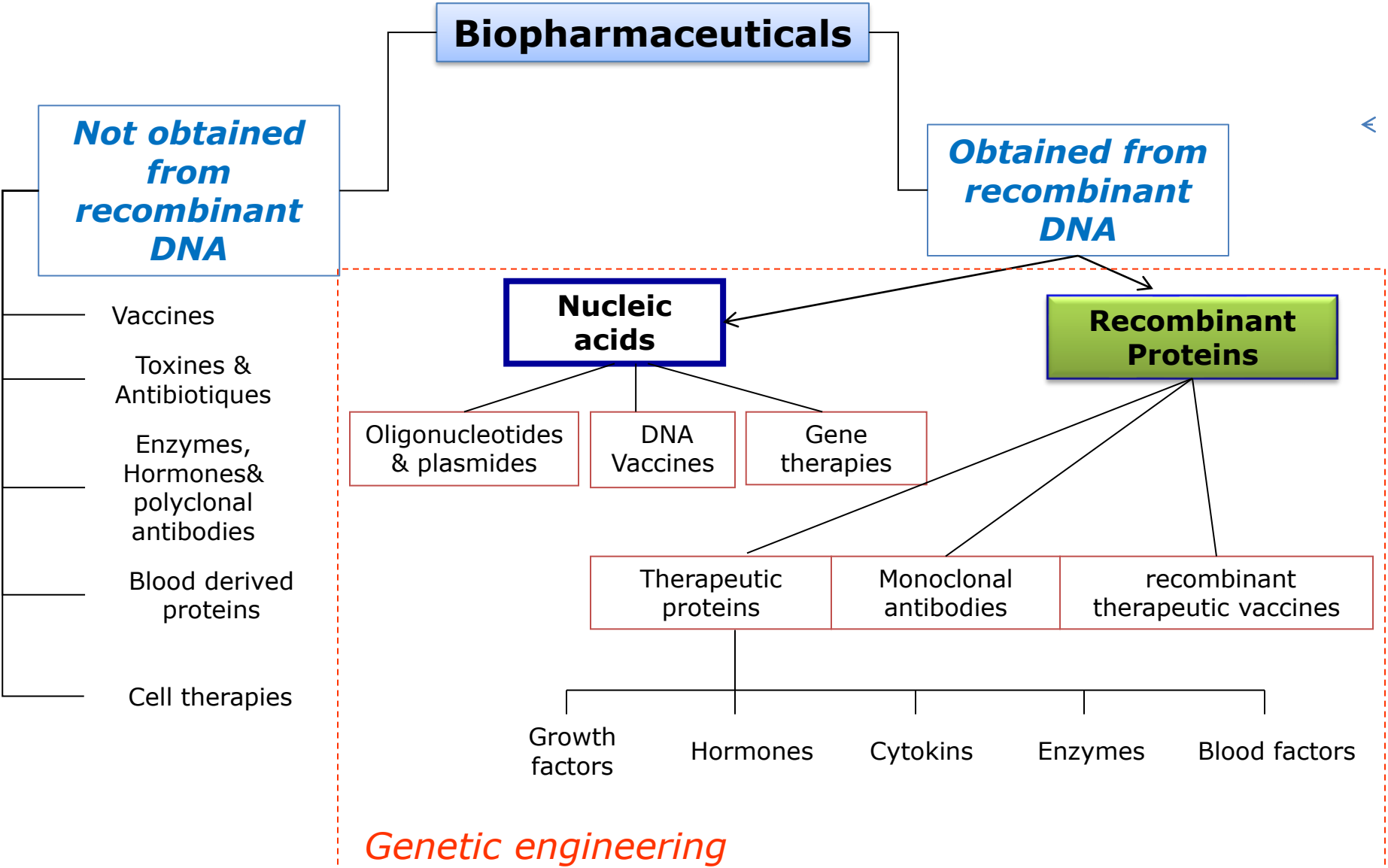
Quality control of biopharmaceuticals

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• Classification of Biopharmaceuticals

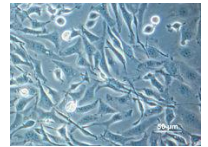


• Different sources of biopharmaceuticals

➤ **Definition:** Biopharmaceuticals are defined as pharmaceuticals manufactured by biotechnology methods, with the products obviously having **biological sources**, usually **involving live organisms or their active components**

1-Extraction : from tissues, animal or human fluids (blood, urines, cells, tissues, milk...)

2-Cell culture production : Prokaryotic cells (bacteria) or eukaryotes (fungi, insects, mammals)



- **natural secretion** : toxins or bacterial enzymes (Botulinum toxin, streptokinase ...)

- **after genetic modification** : expression of a foreign gene

3- From transgenic animals or plants

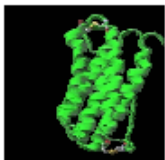


4- Live Organisms (larvae; eggs...)

• Specificities of therapeutic proteins

Therapeutic proteins differ greatly from chemically synthesized molecules by:

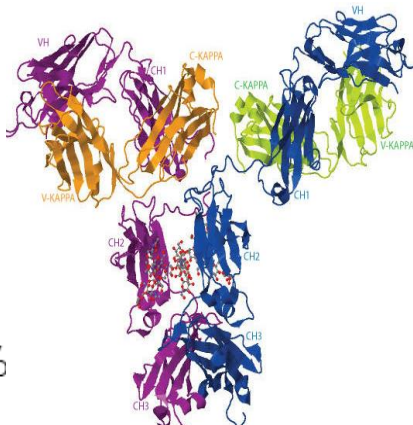
- Their size



Interferon alfa,
165AA, MW: 19 625 Da



Aspirin,
MW: 180 D

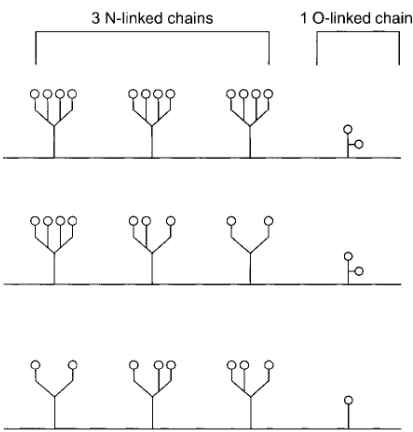


Monoclonal antibody
150kDa

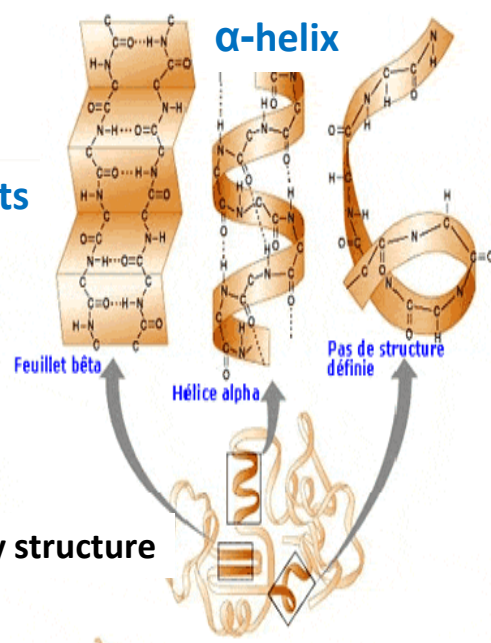
- High structural complexity which is mandatory for their activity

- Heterogeneity

One glycoprotein = mixture of glycoforms in a given and constant proportion



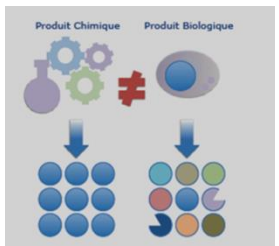
Secondary structure



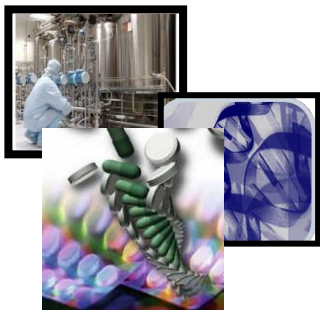
Tertiary structure



- and by the way to produce them



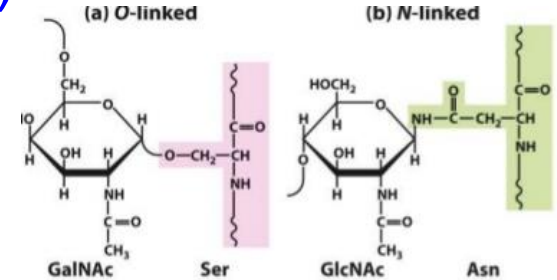
Structure/activity: Glycosylation a major post-translational modification (PMT)



N and O-glycosylation

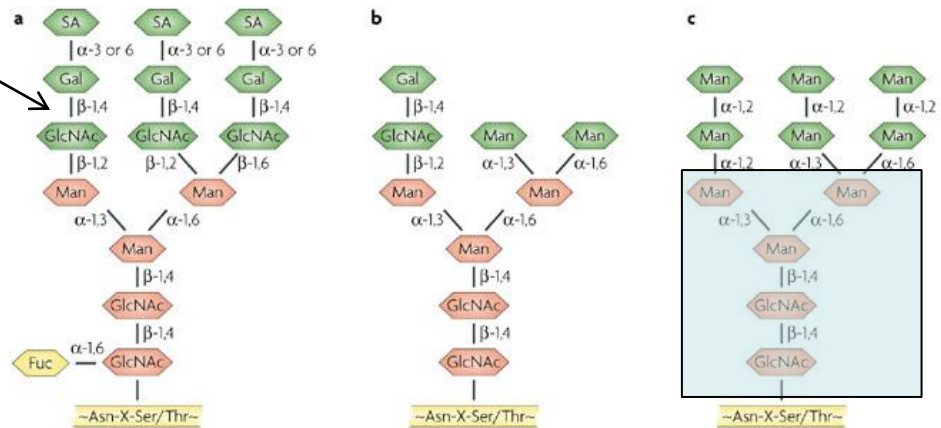
N-glycosylation: Asn (consensus sequence: Asn-X-Ser/thr)

O-glycosylation: Ser; Thr



N-glycans: 3 types

antenna



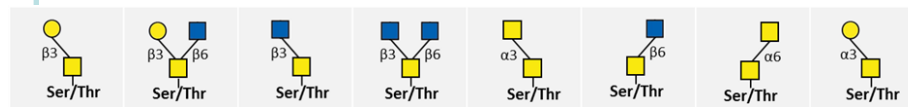
Complex

hybrid

oligomannose

balzarini ;Nature Reviews Microbiology 5, 583-597 (2007)

O-glycans



● Galactose (Gal)
■ N-acetylgalactosamine (GalNAc)
■ N-acetylglucosamine (GlcNAc)

- Many cores
- Shorter
- GalNAc

Why glycosylation is important for biopharmaceuticals?

- Solubility and stability
- Folding
- Bioactivity:
 - Cell recognition (ligand/receptor)
 - Pharmacokinetics
 - in-vivo clearance
 - Antigenicity

• **Quality control of recombinant proteins**

- is mandatory to ensure safety and efficiency
- The active product is heterogeneous and structurally complex
- needs several complementary techniques to address :
 - Purity
 - Identity
 - Dosage
 - Stability
 - Activity

• **At which step?**

- In-process

In-process tests are performed at critical decision-making steps and at other steps where data serve to confirm consistency of the process during the production of either the drug substance or the drug product.

- Purified protein
- Formulated product
- Compounding product (hospital)

• Quality control - Objectives

- QC testing:
- To assess and ensure the safety and efficacy of the therapeutic protein
 - To verify lot-to-lot consistency

Identity test

- To establish that the product batch contains the correct therapeutic protein
- Biological activity, immunogenicity

Dosage

- To determine accurately the concentration of the purified protein / in the final product
- To ensure that the correct amount of active pharmaceutical is delivered to the patient

Purity

- A biopharmaceutical product must be free of contaminating substances



- Impact the performance of the protein
- Cause unwanted and serious side effects (immunogenicity)

Stability

- To evaluate the susceptibility of the protein toward different stresses
- To define optimal storage conditions

• Why the evaluation of the desired product is a real challenge?

A protein:
a complex and heterogenous mixture

Purity profile

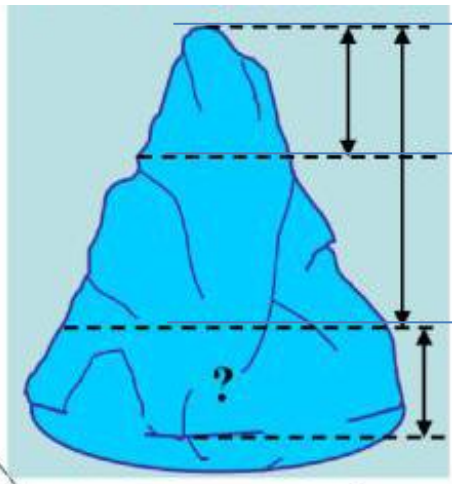
polypeptide
variants

PTM variants

Impurity profile

Product related impurities

Process related impurities



Lot release tests

Characterization

Desired protein

Natural
heterogeneity

« unwanted »
heterogeneity

• What are the impurities?

Process-related impurities:

Expected to be removed during the downstream process

- **Cell-derived**

Host cell proteins, nucleic acids, endotoxins...

- **Cell-culture medium derived**

Inducers, antibiotics, enzymes, serum, bacteria...

- **Downstream derived**

Chemicals and biochemicals, inorganic salts, solvents, residual components

Product-related impurities:

- **Related forms and variants**

Deamidation, oxidation, isomerisation, racemization, glycated, altered PTMs....

- **Truncated forms, fragments;**

- **Dimers, oligomers, aggregates,**

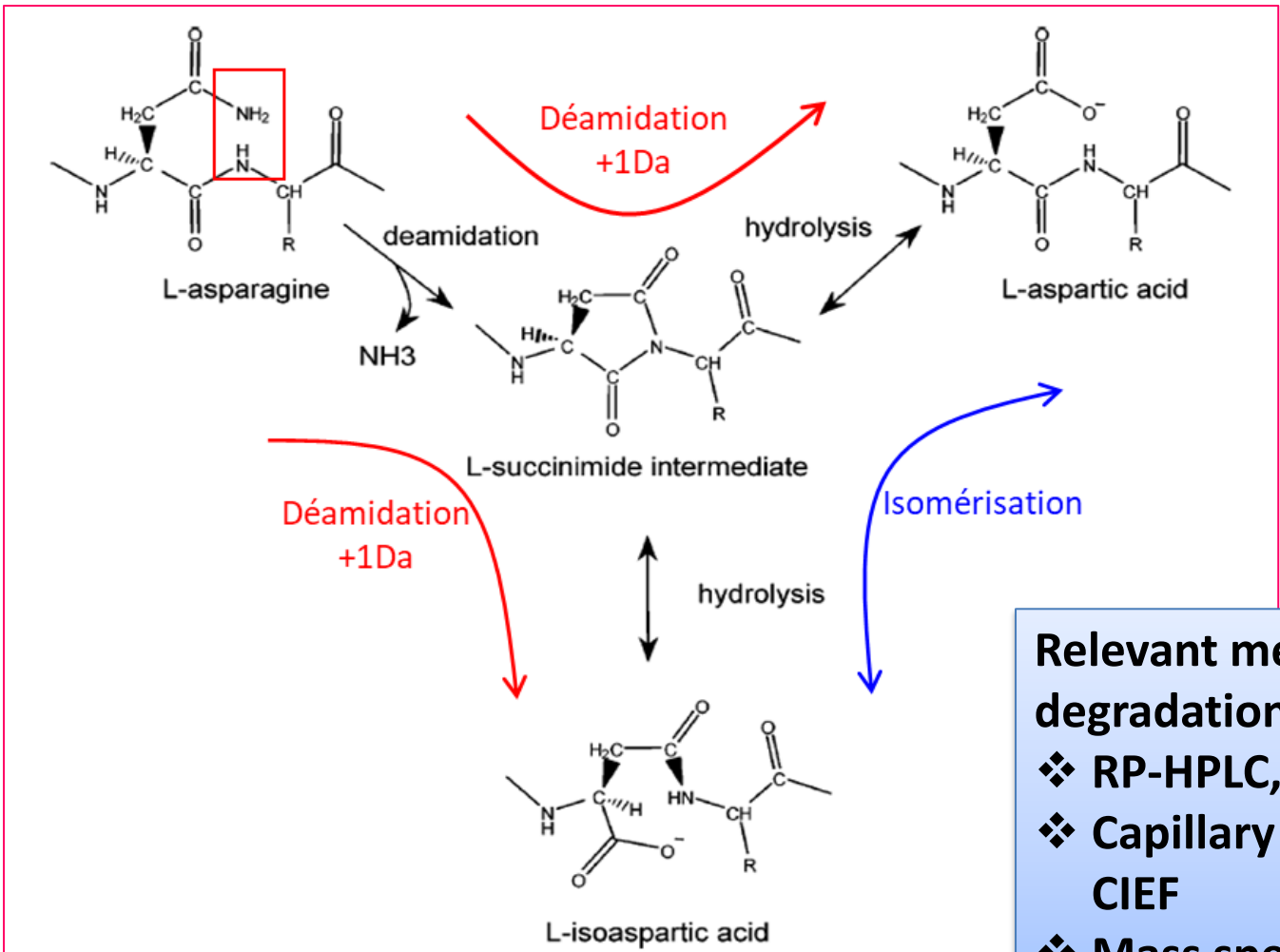
- **Misfolded or unfolded forms**

- **Cys linked variants, scrambling.**

• Chemical Degradation pathways – charge variants

Deamidation (Gln, Asn), Isomerization (Asp)

- Very frequent during production, formulation, storage
- Catalyzed by high temperatures and ionic strengths, at neutral and basic pHs



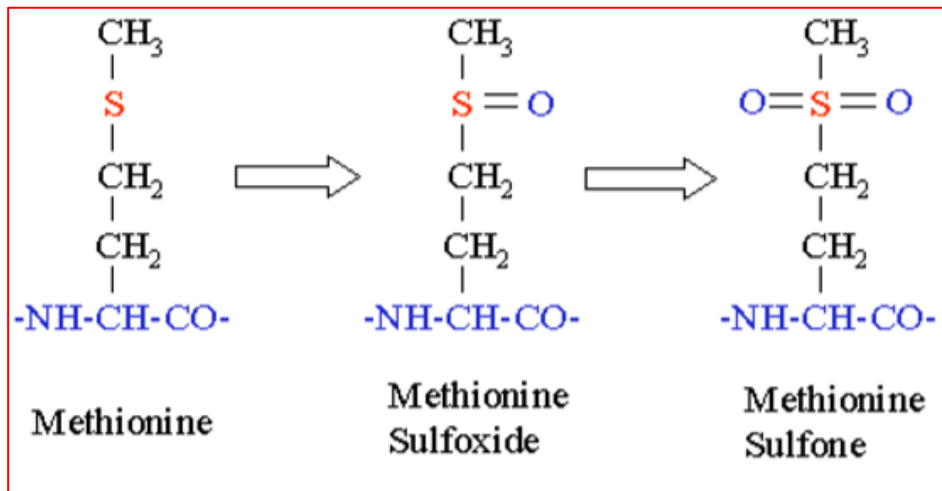
Relevant methods to detect these degradations:

- ❖ RP-HPLC, IEX-HPLC
- ❖ Capillary electrophoresis: CZE, CIEF
- ❖ Mass spectrometry

Chemical Degradation pathways – size variants

Oxidation Met, Cys, (Thr, Phe, Lys)

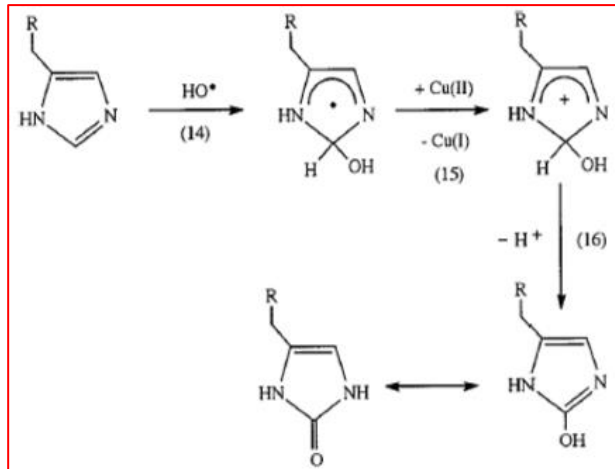
- Very frequent during purification, formulation, storage
- Site specific, metal catalyzed (Met, Cys)
- pH catalyzed (except Met)



Relevant methods:

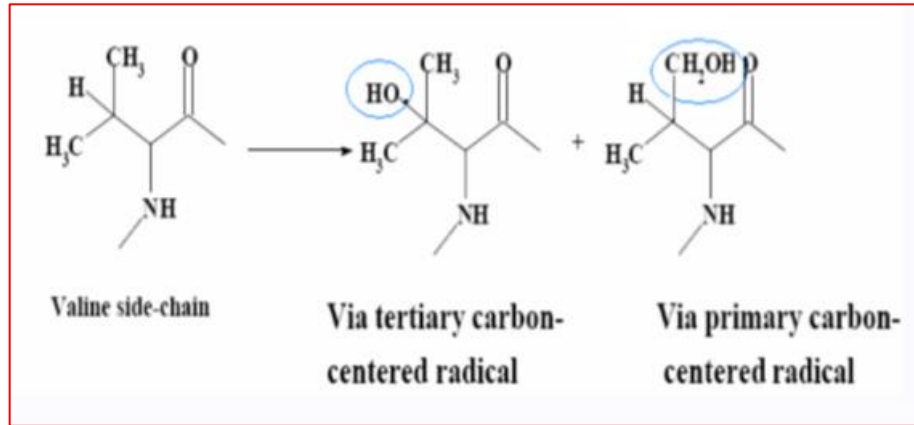
- ❖ RP-HPLC, IEX
- ❖ Capillary electrophoresis
- ❖ Mass spectrometry

His oxidation catalyzed by metal



SCHEME 1. Formation of 2-oxo-His.

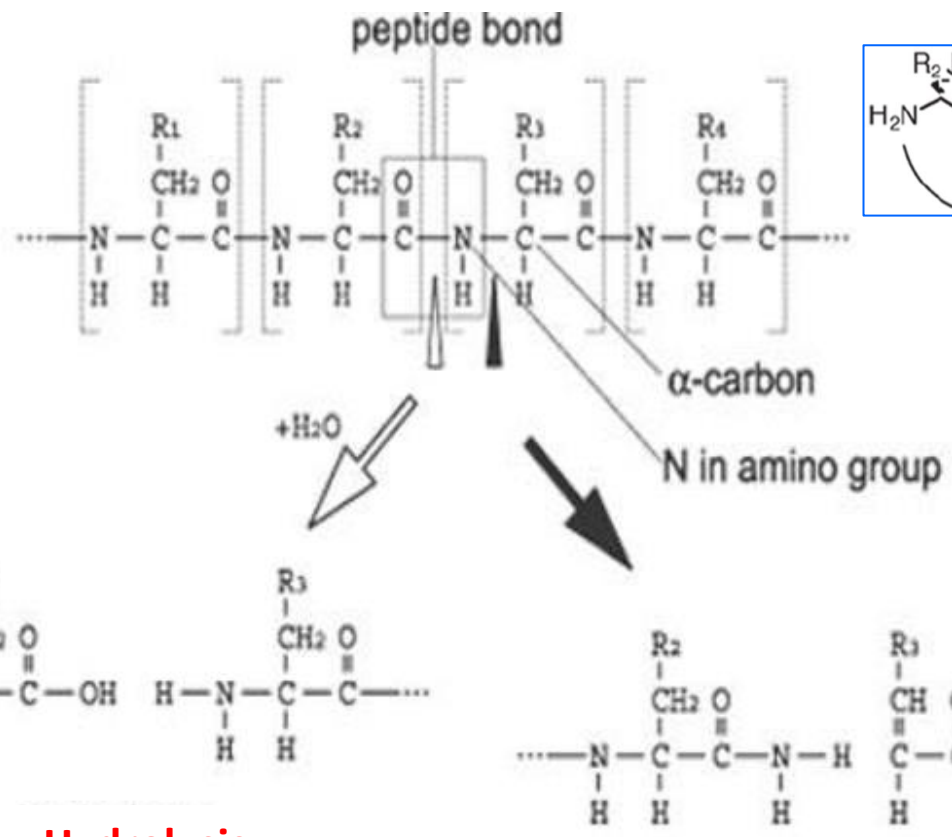
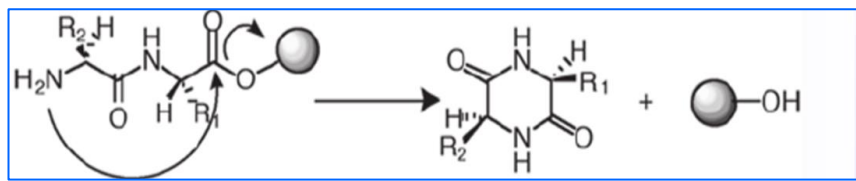
Aliphatic amino acid oxidation



Chemical degradation pathways : fragments / oligomers

Formation of a diketopiperazine (DKP)

N-Term



**Covalent Oligomerisation
Exchange of disulfide bond**

Hydrolysis

Fragmentation of the peptide binding

β Elimination

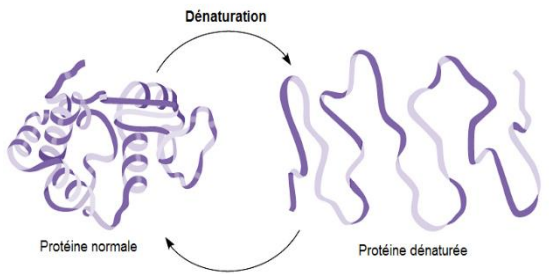
Cys, Ser, Thr, Phe, Lys

Relevant methods:

- ❖ RP-HPLC, size exclusion chromatography (SEC)
- ❖ Capillary gel electrophoresis (CGE)
- ❖ Mass spectrometry

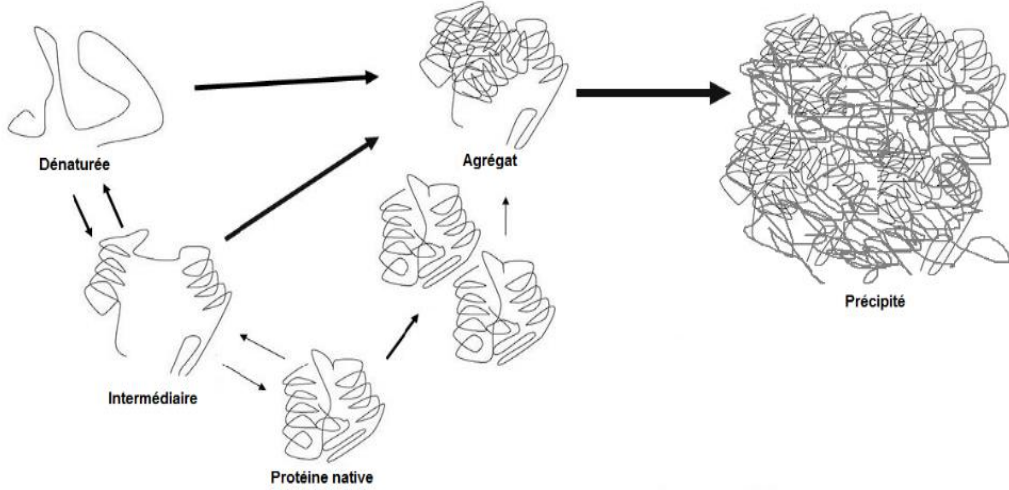
Physical degradation pathways : aggregates, oligomers

➤ Denaturation



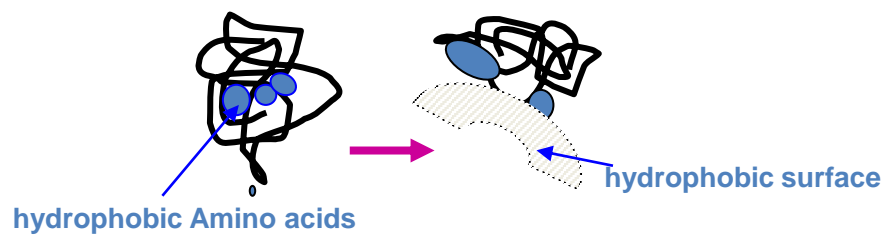
Critical parameters : Temperature, solvents, buffer pH and ionic strenght, chaotropic agents

➤ Agrégation, precipitation



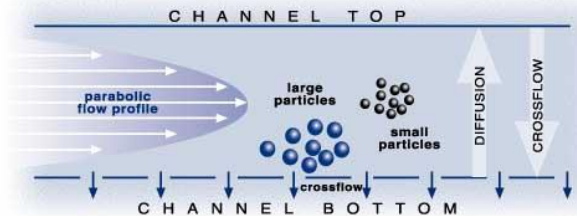
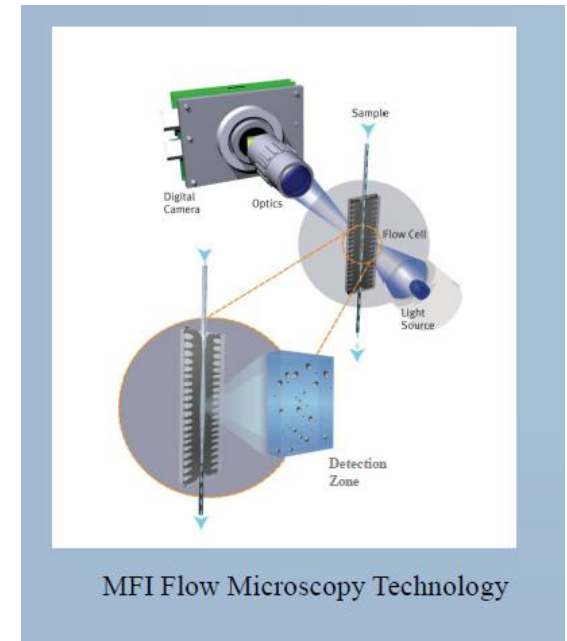
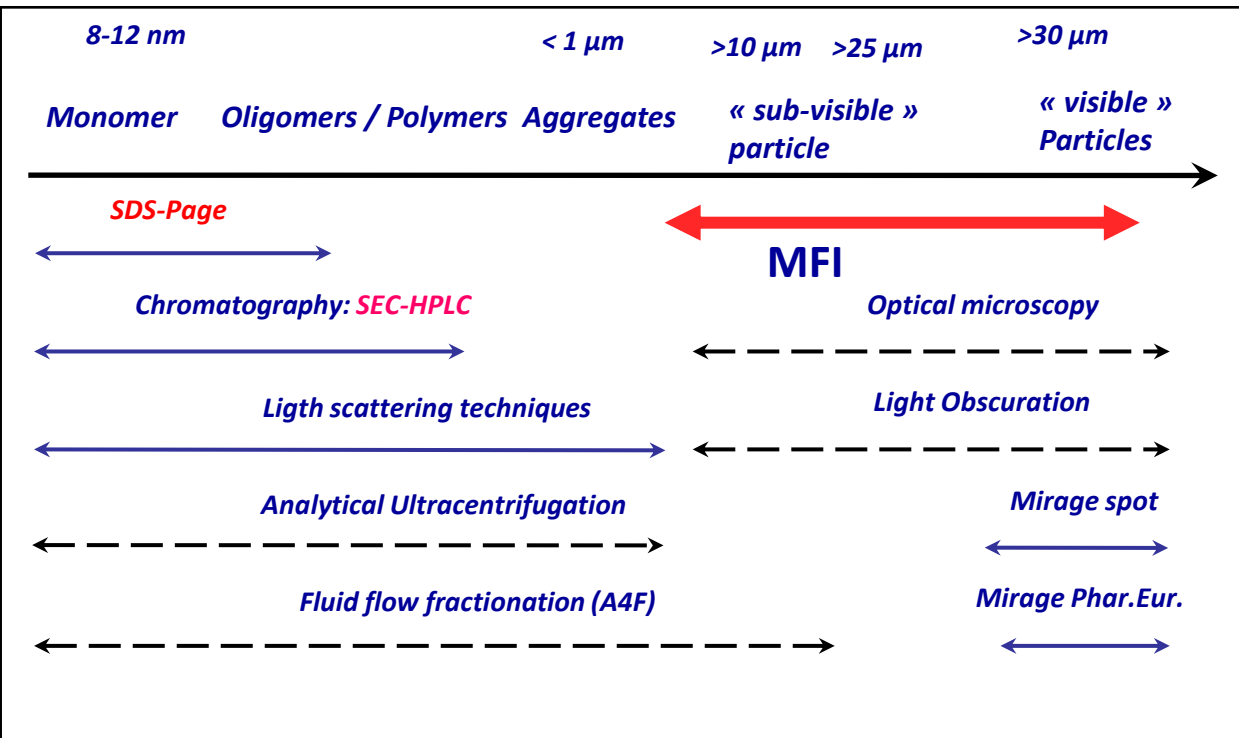
Factors that favor the agrégation:
High concentration of proteins
Stirring, freezing freeze-drying, thawning....

➤ Adsorption



Aggregates and particules detection

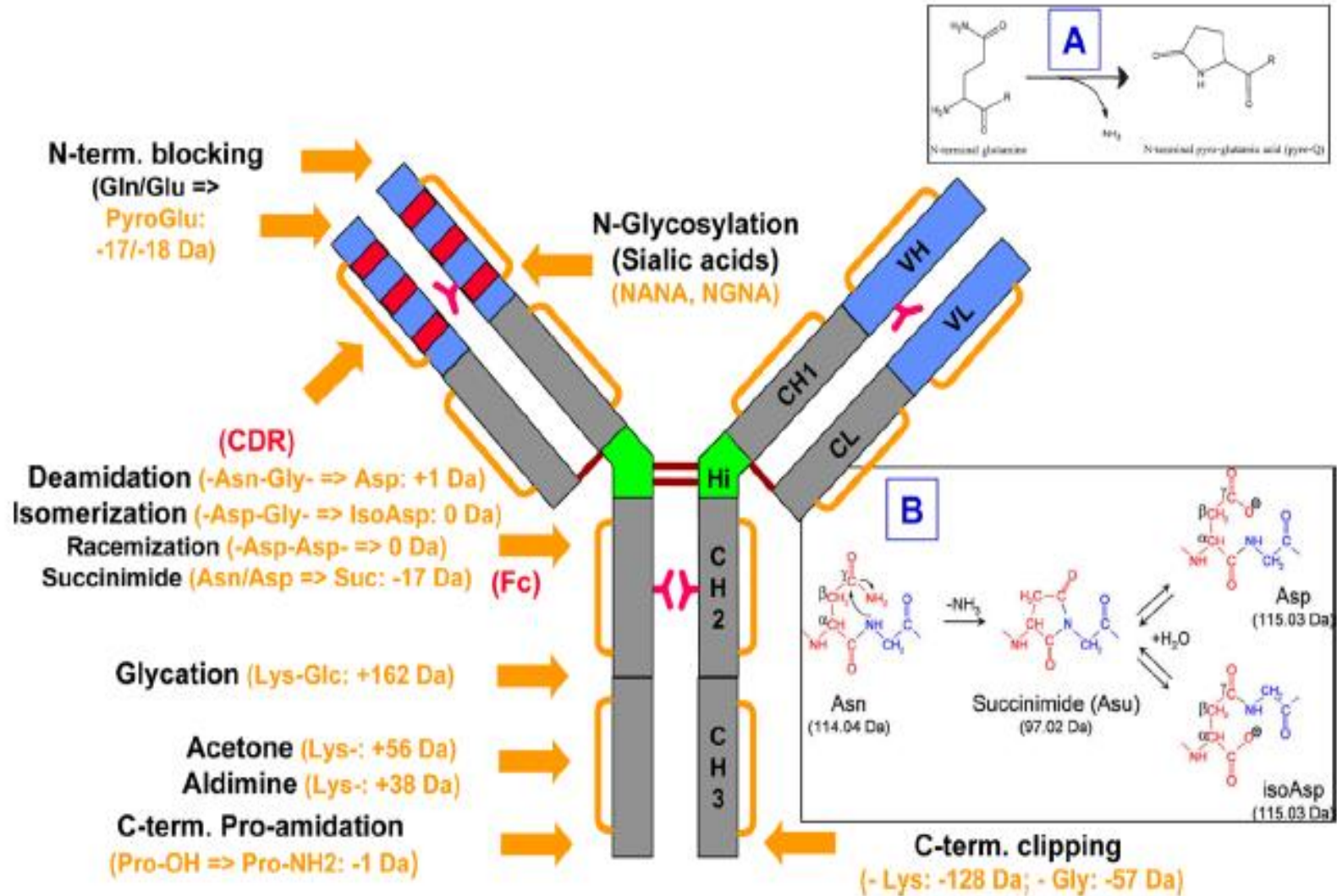
- Aggregates and particles are impurities among others , they can generate undesirable side effects (immunogenicity).
- Analytical techniques to be used depend on their size



Assymetrical field flow fractionation (A4F)

How many microvariants are we looking for?

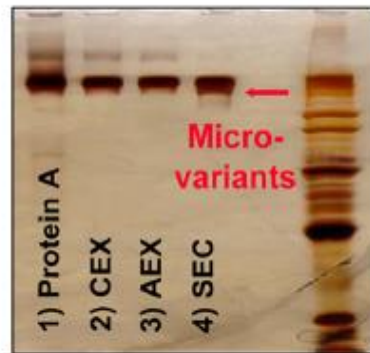
Exemple of a monoclonal antibody



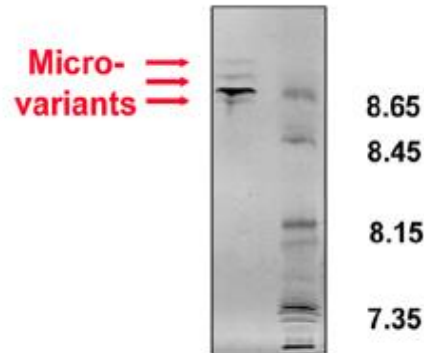
Complementary techniques are required for a QC

- Electrophoresis-based techniques (gel electroph., CE, CE-MS...)
- Liquid Chromatography techniques (all modes/MS)
- Spectrophotometric techniques: Circular dichroism, Fluorescence and UV absorbance, IR/FT, DSC, Light diffusion...

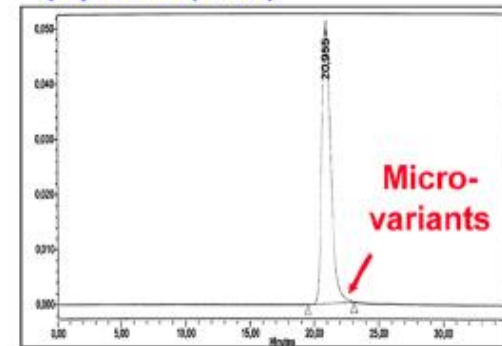
(A) SDS-PAGE (size)



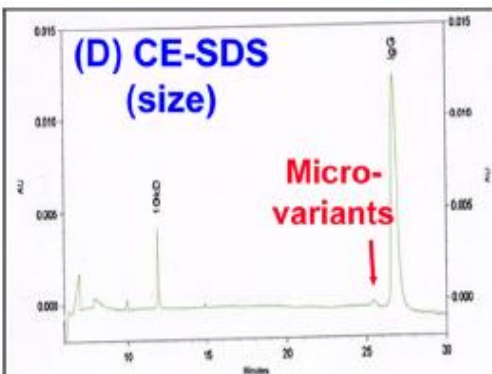
(B) IEF (charge)



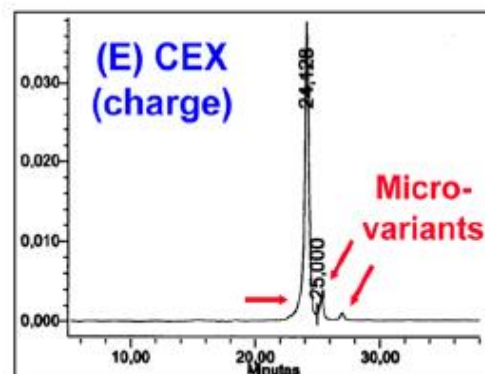
(C) SEC (size)



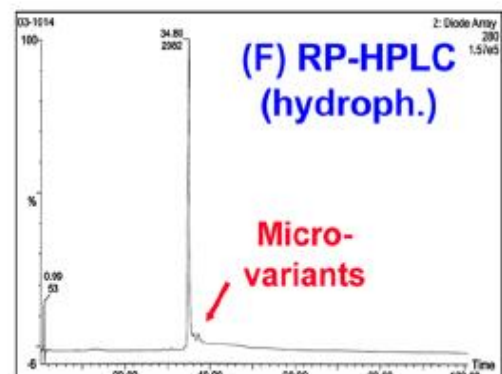
(D) CE-SDS (size)



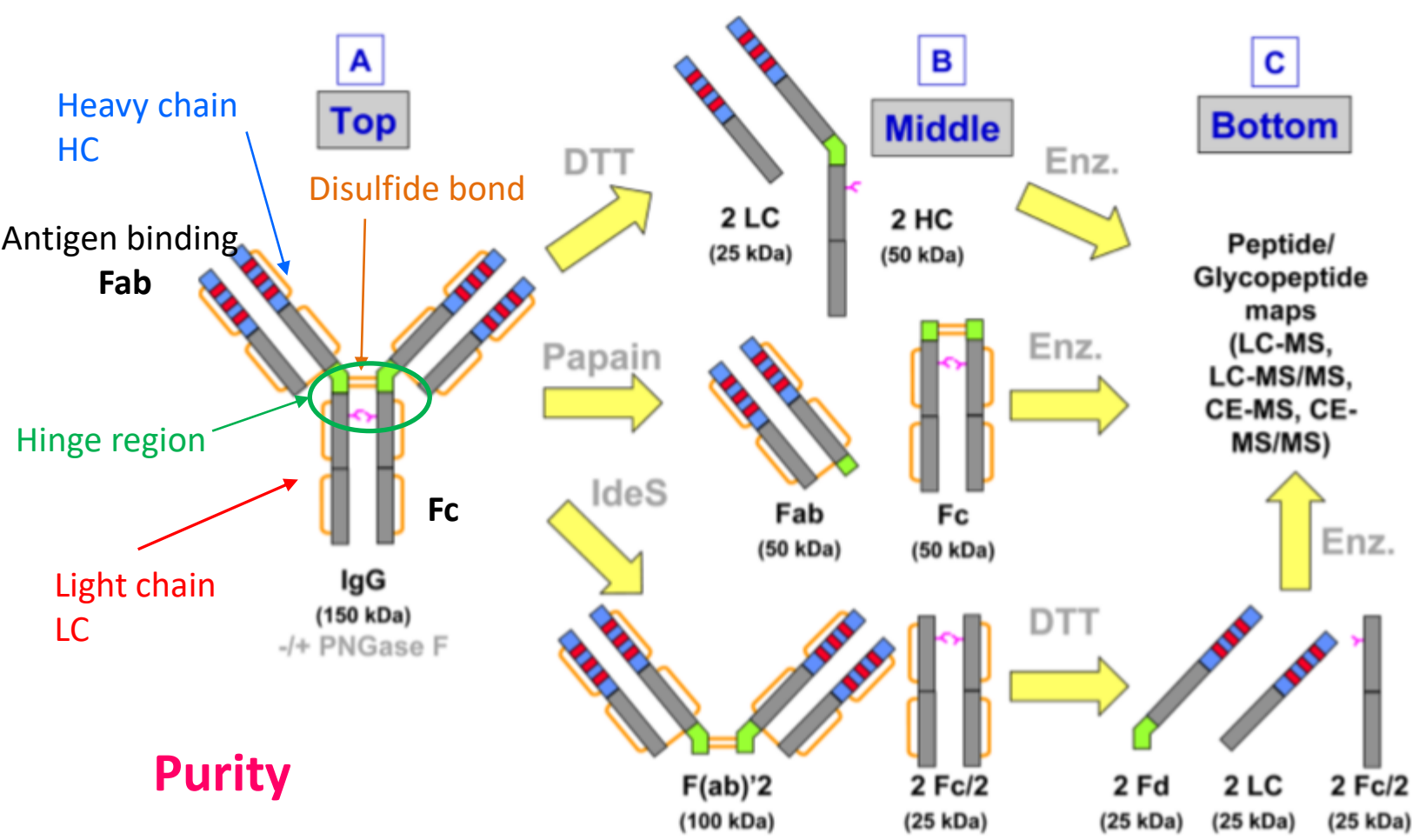
(E) CEX (charge)



(F) RP-HPLC (hydroph.)



3 Approaches for mAbs analysis



Top down, Middle up, Bottom up

DTT: dithiothreitol

Middle up approach to detect Isomerisation and succinimide by HIC-chromatography

HIC: Hydrophobic interaction chromatography

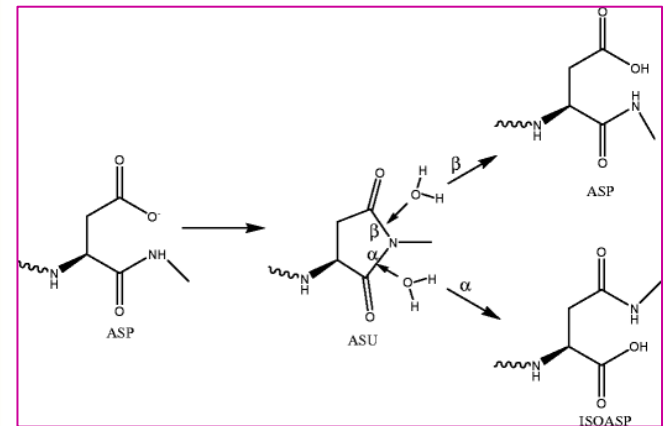
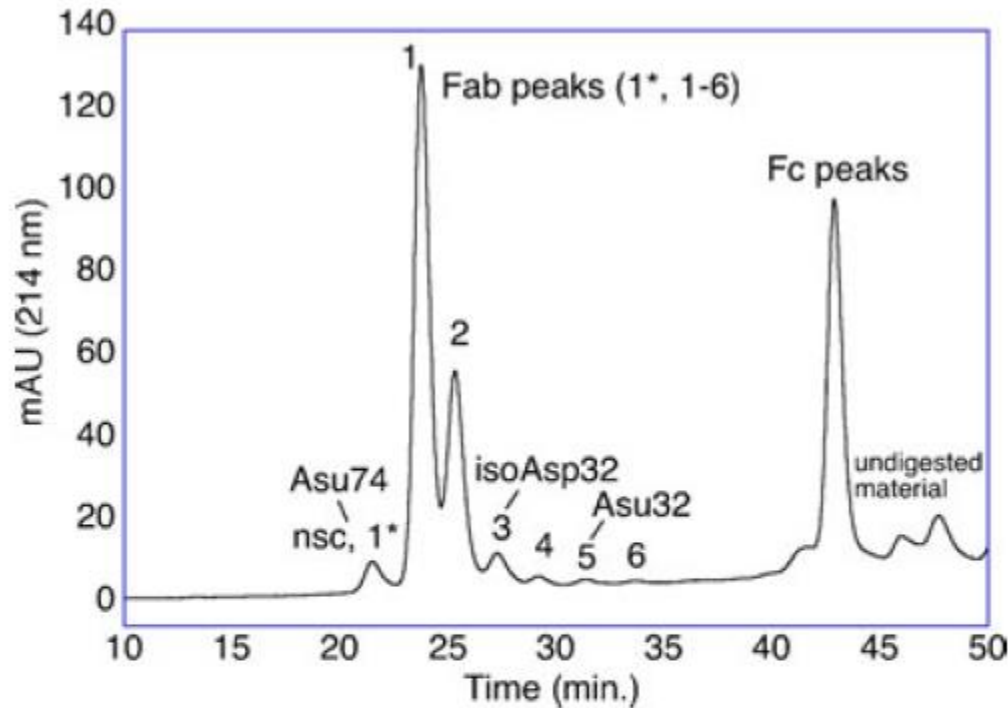


Fig. 1 HIC elution profile of a papain digested MAb1 sample. Peak 1* corresponds to Asu74; nsc=non-specific clip due to papain cleavage, coelutes with peak 1*; Peak 2 is Asp32 with a free thiol group; Peak 3 corresponds to isoAsp32; and Peak 5 corresponds to Asu32.

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Identity test

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Purity

- A biopharmaceutical product must be free of contaminating substances

Dosage

- To determine accurately the concentration of the purified protein / in the final product
- To ensure that the correct amount of active pharmaceutical is delivered to the patient

Stability

- To evaluate the susceptibility of the protein toward different stresses
- To define optimal storage conditions

Impurities and methods to control the purity

1-Proteines from the Host cell : non spécific methods required

- SDS-PAGE/Coomassie blue staining
- «host cell proteins» HCP tests

2- Degraded proteins : resolutive techniques

- RP-CLHP , IEX (deamidated, oxidized forms...), HIC (isomerisation)
- isoelectric focusing (IEF), Capillary zone electrophoresis (glycoforms, deamidated, oxidized forms....)
- Size exclusion chromatography (fragments, agrégates)
- capillary gel electrophoresis (fragments, agrégates, non glycosylated sites...)
- hydrophobic interaction chromatography (HIC) : misfolding , unfolding:
- techniques specific for aggregates and particles detection

3-DNA

- Hybridization method using radiolabelled probes

4- Pyrogenic substances, endotoxins :

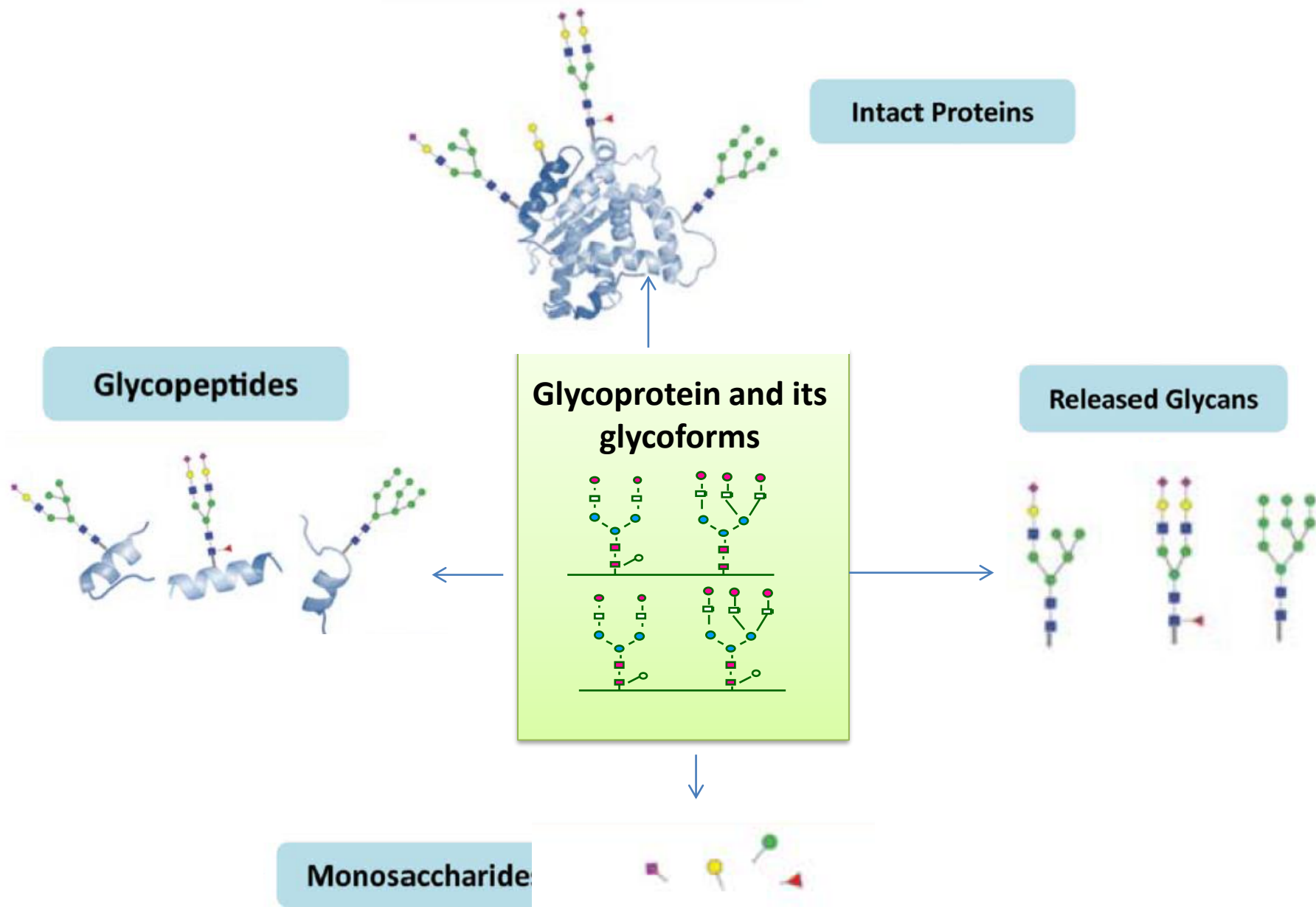
- LAL test: *in vitro* test based on activation of the complement cascade by endotoxin in *Limulus Polyphemus*
- New coming “RP-HPLC-based technique”

5- Virus

Approaches for Identity control

- **Primary sequence**
 - amino acid composition
 - automated sequencing (EDMAN)
 - peptide mapping
- **Physicochemical parameters estimation**
 - Molecular mass: SDS-PAGE, size exclusion chromatography, capillary Gel Electrophoresis mass spectrometry (MALDI HPLC-ESI-MS using ion trap or FTICR mass spectrometry)
 - Isoelectric point : gel isoelectrofocusing (IEF) or capillary isoelectrofocusing (CIEF)
 - Hydrophobicity : RP-HPLC
 - Combination of several parameters : PAGE , Capillary zone electrophoresis
- **Secondary and tertiary structure**
 - Circular dichroism
 - Infra-Red -based techniques
- **Post translationnal modifications : Glycosylation mostly**
 - Monosaccharide composition, sialic acid...
 - Glycan mapping
 - Glycopeptide mapping
 - Glycoform analysis: heterogeneity profile
- **Biological and immunological activity**

Glycosylation analysis can be performed at different levels



Dosage and activity

- **Colorimetric methods**

- *Méthodes du biuret et dérivées :*
Biuret : $\text{CuSO}_4(\text{OH}^-)$ à 540 nm \rightarrow 0,1 g/l
Lowry : + Folin-Ciocalteu (Tyr) à 650-750 nm \rightarrow 0,05 g/l
BCA : + acide bicinchoninique (inverse) à 560 nm \rightarrow 0,01 g/l
- *Méthodes par fixation de colorants :* Bradford au bleu de Coomassie G250 (H^+) à 600-650 nm \rightarrow 0,01 g/l

- **Spectrophotometry (UV at 280nm)**

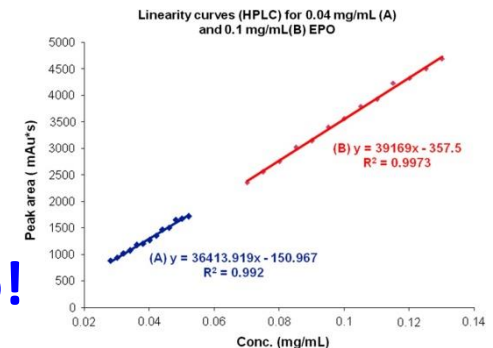
- **Immunochemical methods : ELISA..**

- **HPLC, CE techniques can be quantitative too!**

- **Biological activity:**

- Administer a known quantity of the product to a biological system (cells, animal...) and measure a quantitative response that will be transcribed into a unit of activity.
- In Vivo, Ex vivo, in vitro (cell cultures), biochemical or enzymatic assays, binding tests (SPR.....)

EPO dosage



Rane et al., 2012

• Analytical techniques for QC of common variants of biopharmaceuticals

	Modifications	Analytical techniques
Charge variants	Deamidation Oxidation Glycosylation	CEX, IEF, CIEF, CZE ; HILIC-HPLC
Size variants	Fragmentation, Aggregation	SEC-HPLC, SDS-PAGE, CGE
Conformers	Misfolding / Denaturation	HIC (not in a QC context)

Homework for the 5th of November

- Prepare **15 min** oral presentation PPT
(groups of 3/4 students; each group assisted by one teacher T1 or T2)

Group 1: Acidic and basic variants analysis of Monoclonal Antibodies by IEX-HPLC- (T1)

Group 2: Purity control of therapeutic proteins by Size Exclusion Chromatography (SEC-HPLC) (T1)

Group 3: SDS-PAGE for purity and identity controls of therapeutic proteins (T1)

Group 8: detection of chemical-Degradation variants of therapeutic proteins by RP-HPLC

Group 5: Peptide mapping of therapeutic proteins by RP-HPLC as an identity Control (T2)

Group 6: N- Glycan mapping of therapeutic glycoproteins by HILIC-HPLC (T2)

Group 7: Capillary gel electrophoresis for identity and purity control of Therapeutic proteins (T2)

Groupe 4: Control of Secondary structure of therapeutic proteins- (T2)

Oral presentation

- **3-5 slides/ 15 Min/ PPT presentation: use the template for each topic on ecampus**
 - Principle/Mechanism of the technique/approach used for this specific application
 - Type of information provided **for quality control purposes only** , Limits and advantages of the technique
 - A few examples illustrating this technique/method **for quality Control of therapeutic proteins**
 - 30 min of group works (now)
 - **BE CAREFUL : 15 min for presentation is a maximum!**
 - **Documents available on ecampus platform; USE PREFERENTIALLY the selected ARTICLES for your presentation** (figures or info...)
 - TU08- biotechnology <supporting files< one folder per topic
 - PPT presentation , to be sent by email to your teacher (T1 or T2) before the **5th of November 2023 12:00am**
 - **Presentation the 6th of November 9-12am**