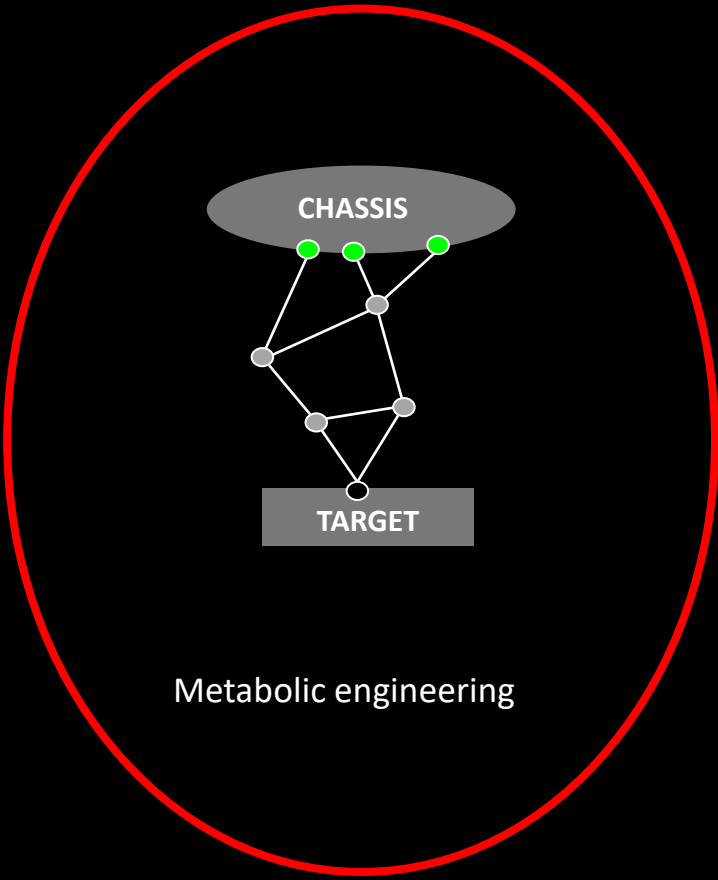


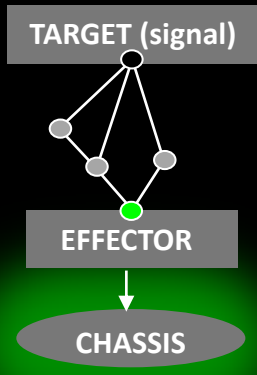
Produce, sense and compute with metabolism

Jean-Loup Faulon - Jean-Loup.Faulon@inrae.fr - <https://jfaulon.com>

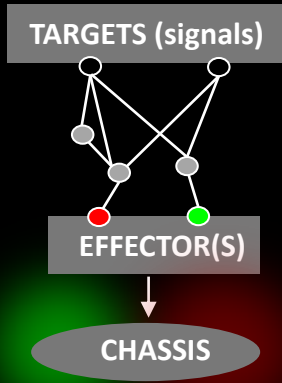
Engineering metabolic and genetic networks in vitro (cell-free) and in vivo (E. coli)



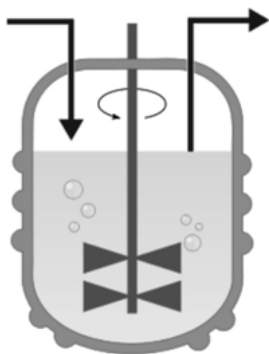
Metabolic engineering



Biosensor engineering



Information processing
biocomputing



Many tools that do not necessarily follow FAIR principles

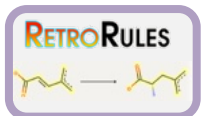


EBERHARD KARLS UNIVERSITÄT TUBINGEN

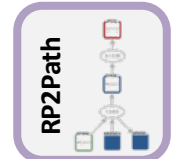


web service

EPFL



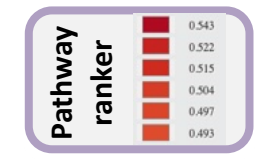
INRAE



web service WEIZMANN INSTITUTE OF SCIENCE



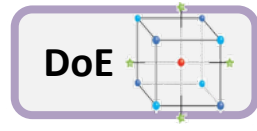
INRAE



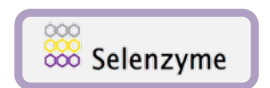
web service INRAE



web service UNIVERSITY OF LIVERPOOL



Imperial College London



web service MANCHESTER 1824



BERKELEY LAB



web service

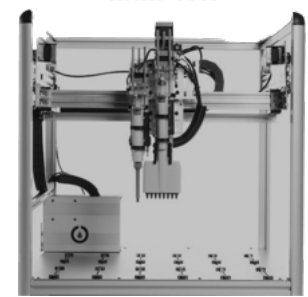
THE UNIVERSITY of EDINBURGH



web service



UNIVERSITY of WASHINGTON



Synthace



web service

Imperial College London



User-friendly
Widely used
Flexible
FAIR

— 9k+ tools
— 50 categories

https://toolshed.g2.bx.psu.edu

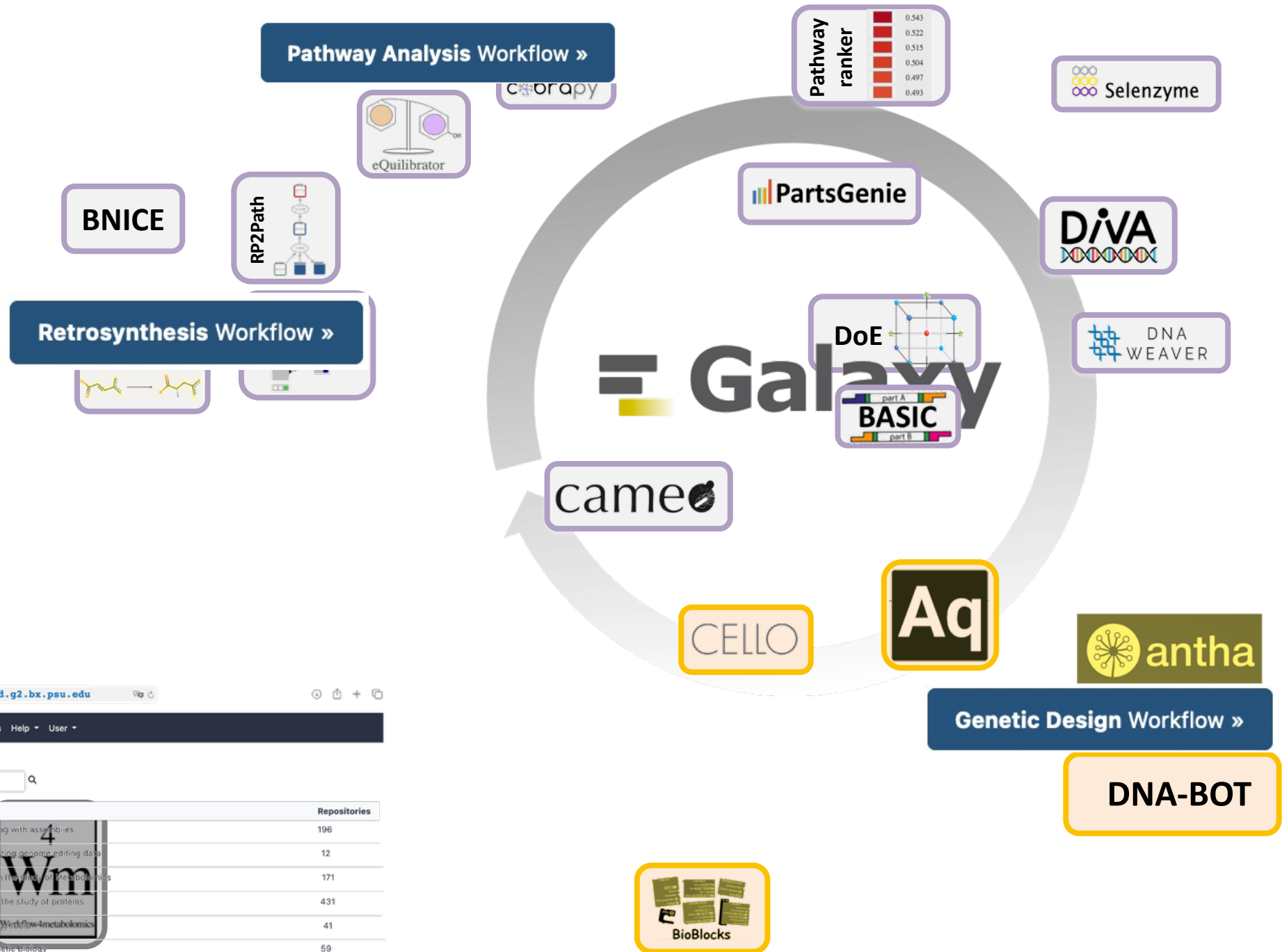
Galaxy Tool Shed Repositories Groups Help User

9425 valid tools on Apr 04, 2023

Search
Search for valid tools
Valid Galaxy Utilities
Tools
Custom datatypes
Repository dependency definitions
Tool dependency definitions
All Repositories
Browse by category
Available Actions
Login to create a repository

Repositories by Category

Name	Description	Repositories
Assembly	Tools for working with assemblies	196
Genome editing	Tools for amending genome editing data	12
Metabolomics	Tools for use in the study of metabolomics	171
Proteinomics	Tools enabling the study of proteins	431
Systems Biology	Systems biology workflow/metabolomics	41
Synthetic Biology	Tools for synthetic biology	59





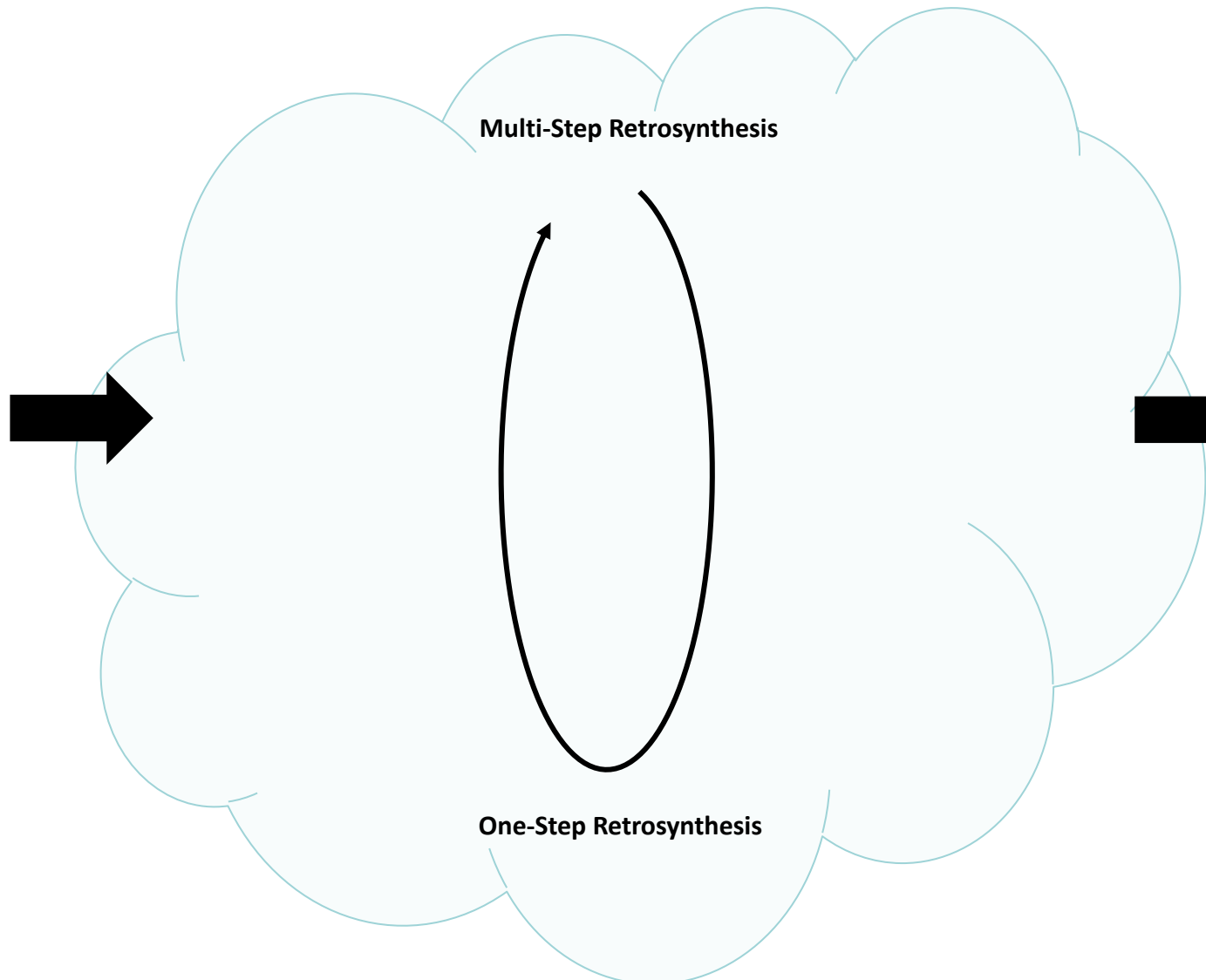
galaxy-synbiocad.org



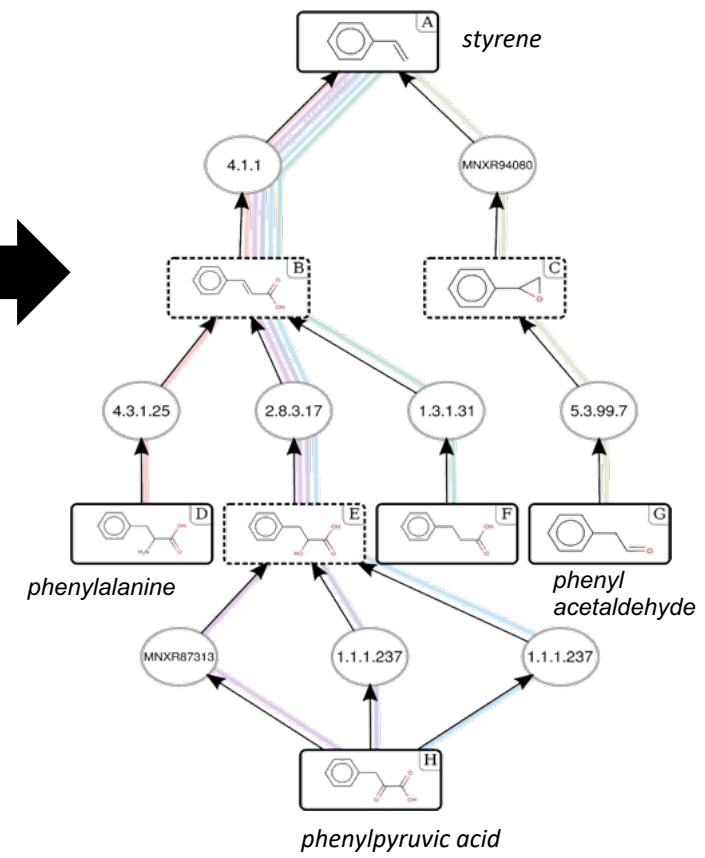
Reaction database
USPTO
MetaNetX

a target molecule
c1ccccc1C=C
styrene

Available molecules
C1=CC=CC=C1
Available molecules in a host strain



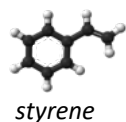
A retrosynthesis tree/graph



Reaction database



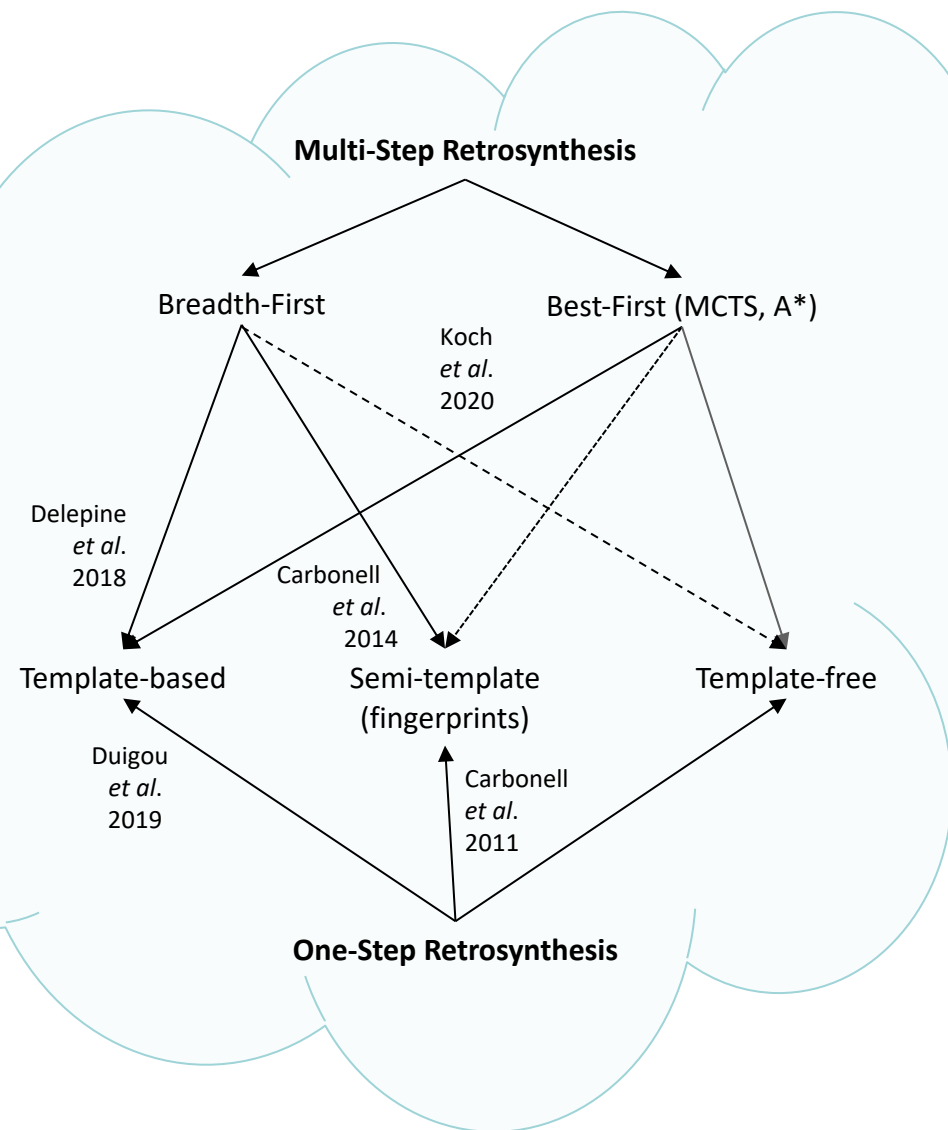
a target molecule



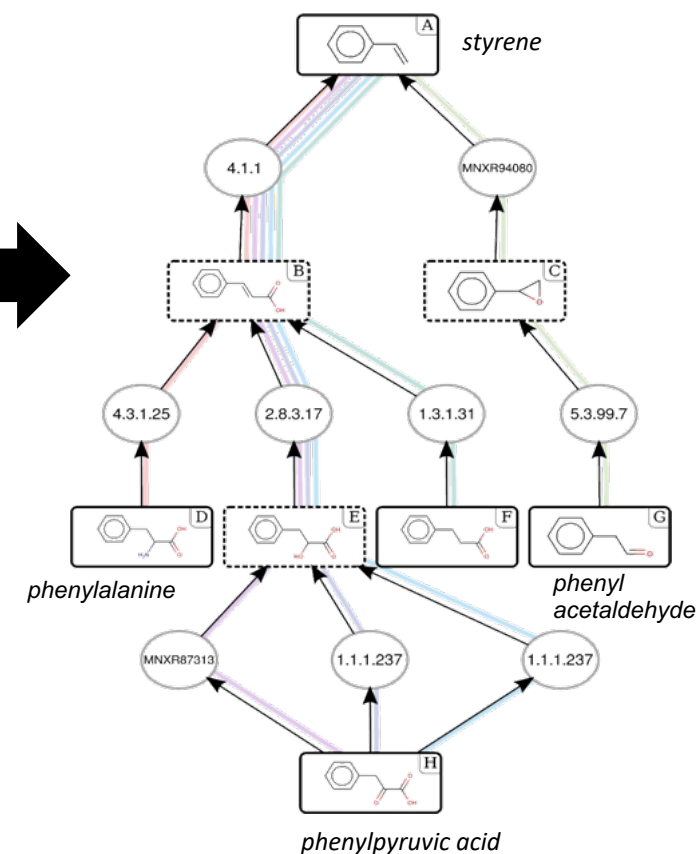
Available molecules



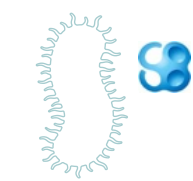
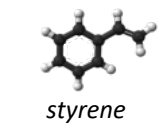
Available molecules in a host strain



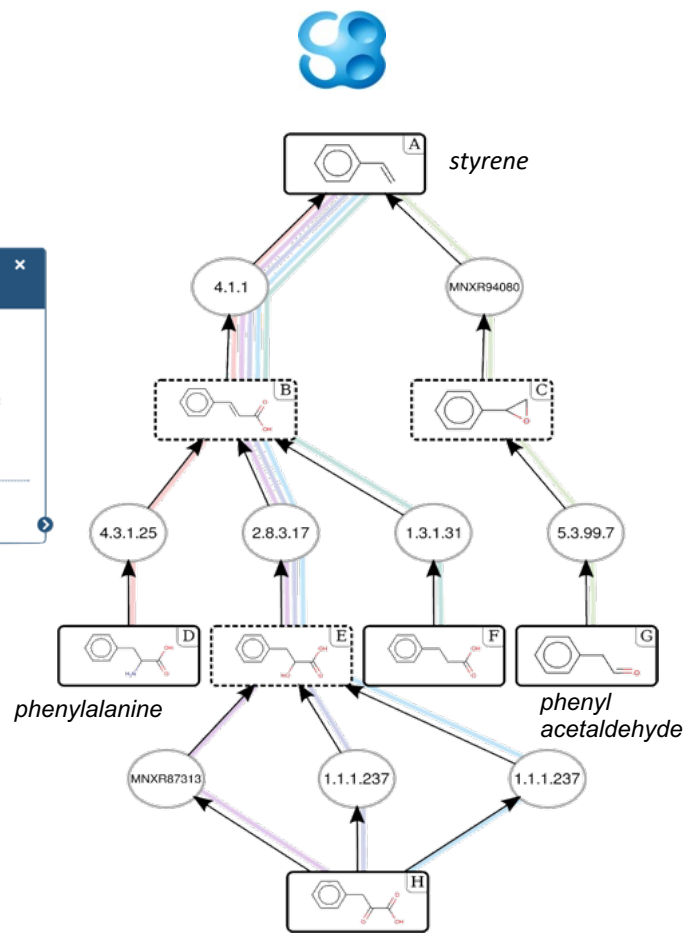
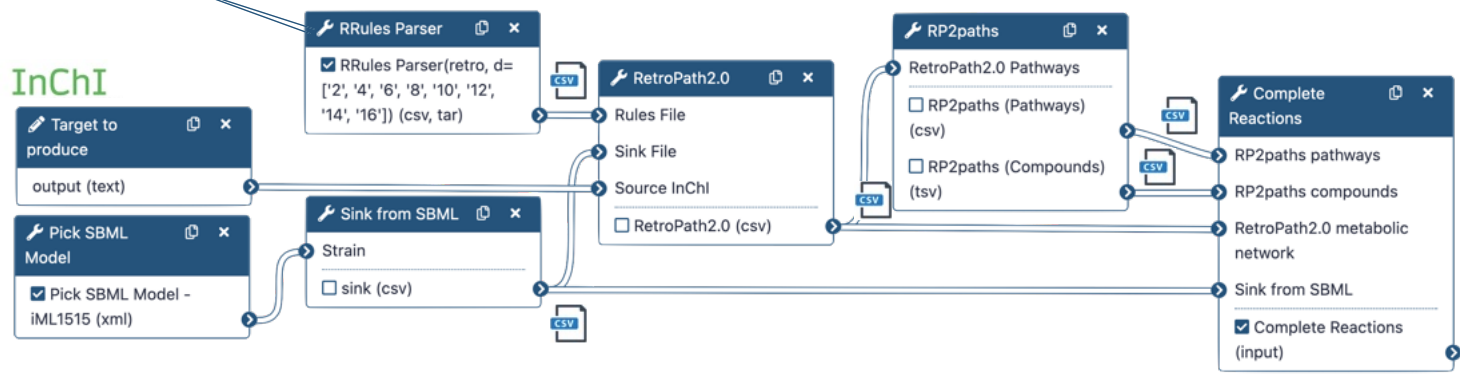
A retrosynthesis tree/graph



- Carbonell *et al.* *BMC Sys. Biol.*, 2011
- Carbonell *et al.* *Nucleic Acids Res.*, 2014
- Delepine *et al.* *Metabolic Eng.*, 2018
- Duigou *et al.*, *Nucleic Acids Res.*, 2019
- Koch *et al.* *ACS Synth Biol.*, 2020



SMARTS
InChI



Available molecules in a host strain

Nucleic Acids Research

RetroRules: a database of reaction rules for engineering biology

Thomas Duigou, Melchior du Lac, Pablo Carbonell, Jean-Loup Faulon

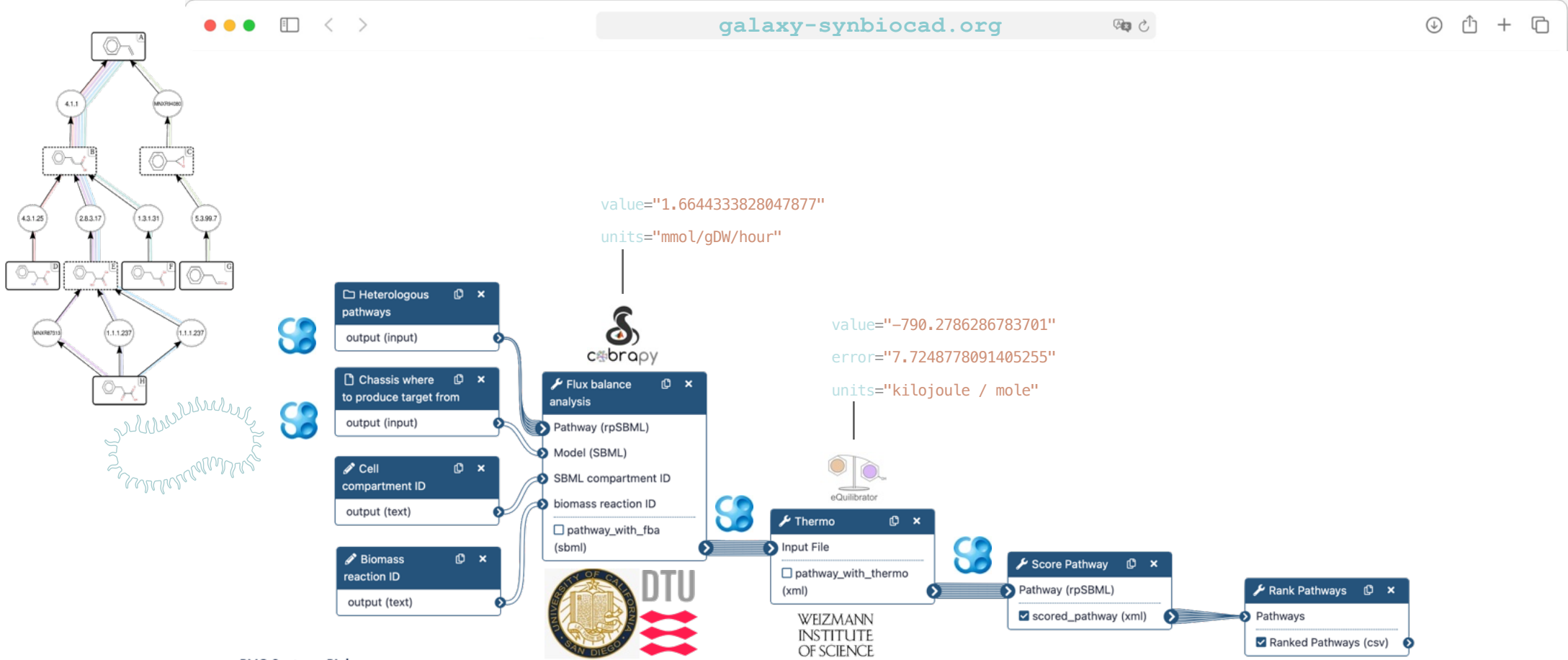
Nucleic Acids Research, Volume 47, Issue D1, 08 January 2019, Pages D1229–D1235, <https://doi.org/10.1093/nar/gky940>



Metabolic Engineering
Volume 45, January 2018, Pages 158-170

RetroPath2.0: A retrosynthesis workflow for metabolic engineers

Baudoin Delépine, Thomas Duigou, Pablo Carbonell, Jean-Loup Faulon



BMC Systems Biology

Software | [Open Access](#) | Published: 08 August 2013

COBRAPy: COntstraints-Based Reconstruction and Analysis for Python

All Ebrahim, Joshua A Lerman, Bernhard O Palsson & Daniel R Hyde

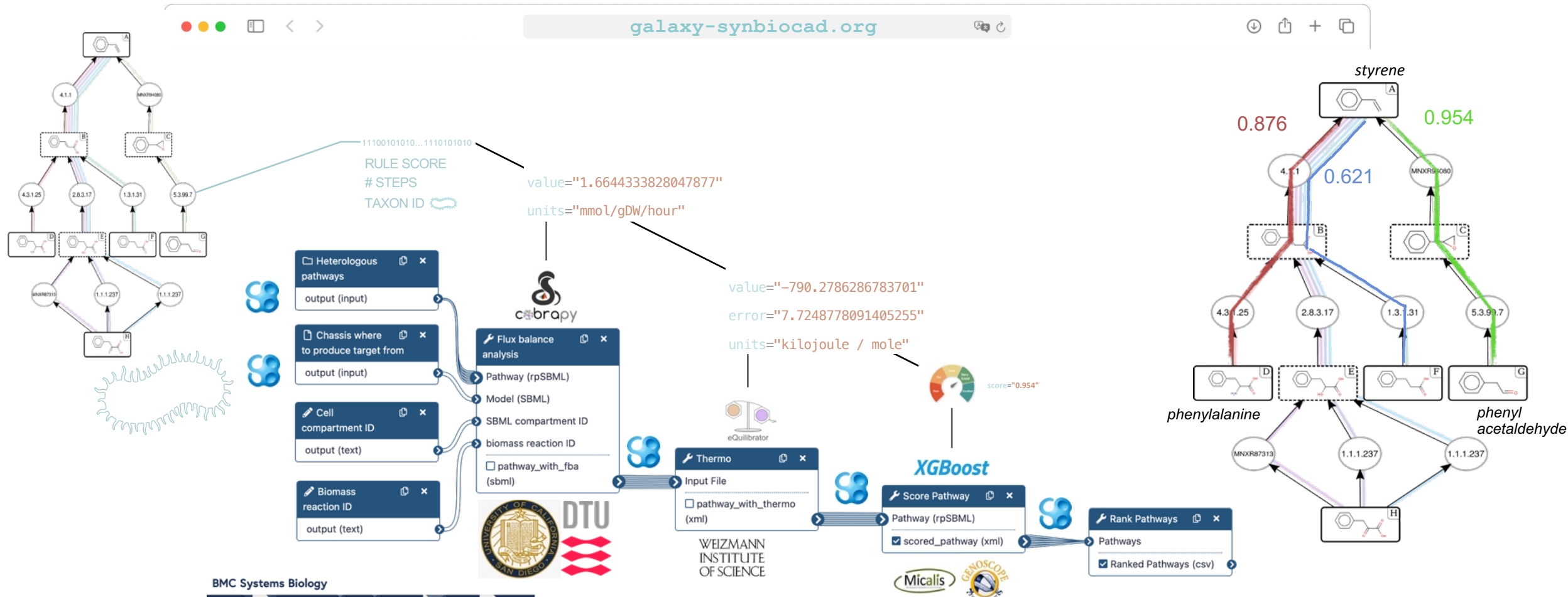
BMC Systems Biology 7, Article number: 74 (2013) | [Cite this article](#)

PLOS COMPUTATIONAL BIOLOGY

Consistent Estimation of Gibbs Energy Using Component Contributions

Elaad Noor, Hulda S. Haraldsdottir, Ron Milo, Ronan M. T. Fleming

Published: July 11, 2013 | <https://doi.org/10.1371/journal.pcbi.1003098>



BMC Systems Biology

Software | [Open Access](#) | [Published: 08 August 2013](#)

COBRaPy: COstraints-Based Reconstruction and Analysis for Python

[Ali Ebrahim](#), [Joshua A Lerman](#), [Bernhard O Palsson](#) & [Daniel R Hyde](#)

BMC Systems Biology 7, Article number: 74 (2013) | [Cite this article](#)

PLOS COMPUTATIONAL BIOLOGY

Consistent Estimation of Gibbs Energy Using Component Contributions

[Elad Noor](#) [Huida S. Haraldsdóttir](#) [Ron Milo](#) [Ronan M. T. Fleming](#)

Published: July 11, 2013 • <https://doi.org/10.1371/journal.pcbi.1003098>

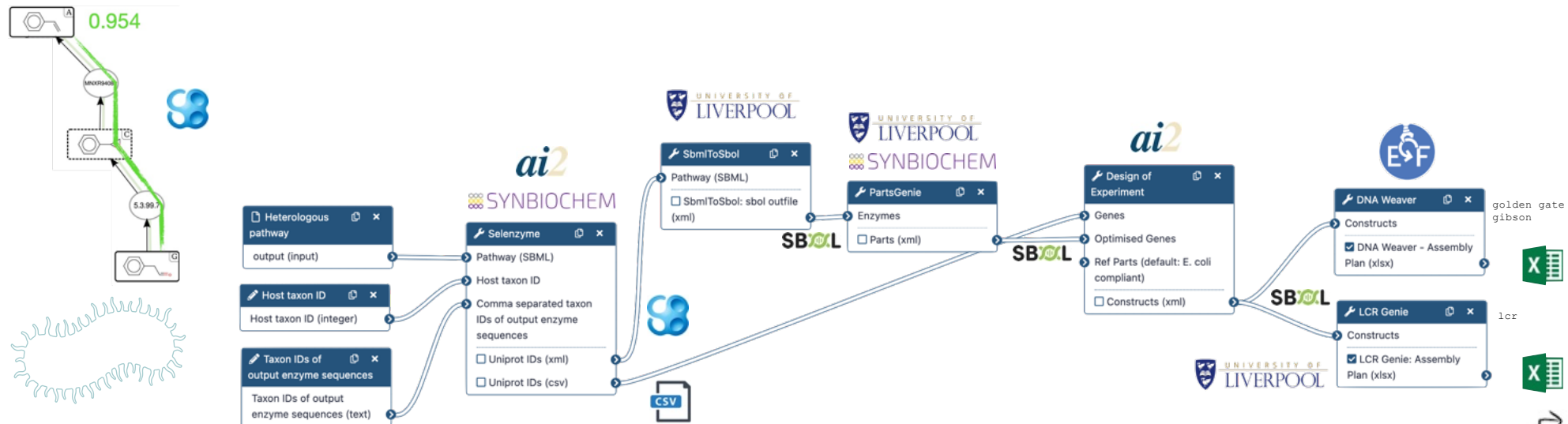
Nucleic Acids Research

XTMS: pathway design in an eXTended metabolic space

[Pablo Carbonell](#) [Pierre Parutto](#), [Joan Herisson](#), [Shashi Bhushan Pandit](#), [Jean-Loup Faulon](#)

Nucleic Acids Research, Volume 42, Issue W1, 1 July 2014, Pages W389–W394, <https://doi.org/10.1093/nar/gku362>

phenylpyruvic acid



OptBioDes: optimal design for the SynBio toolchain

P. Carbonell, R. Breiting, J.-L. Faulon, and the SYNBOCHEMA team
 IWBD, Cambridge (UK), 2019

Selenzyme: enzyme selection tool for pathway design

Pablo Carbonell, Jerry Wong, Neil Swainston, Eriko Takano, Nicholas J Turner, Nigel S Scrutton, Douglas B Kell, Rainer Breiting, Jean-Loup Faulon

Bioinformatics, Volume 34, Issue 12, 15 June 2018, Pages 2153–2154,

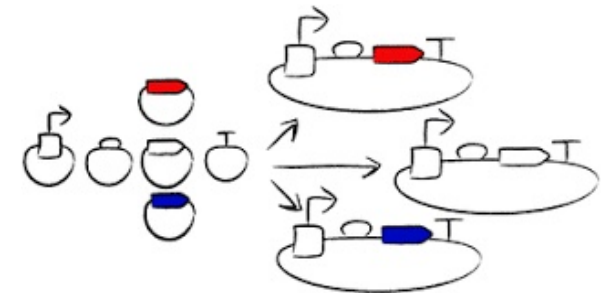
PartsGenie: an integrated tool for optimizing and sharing synthetic biology parts

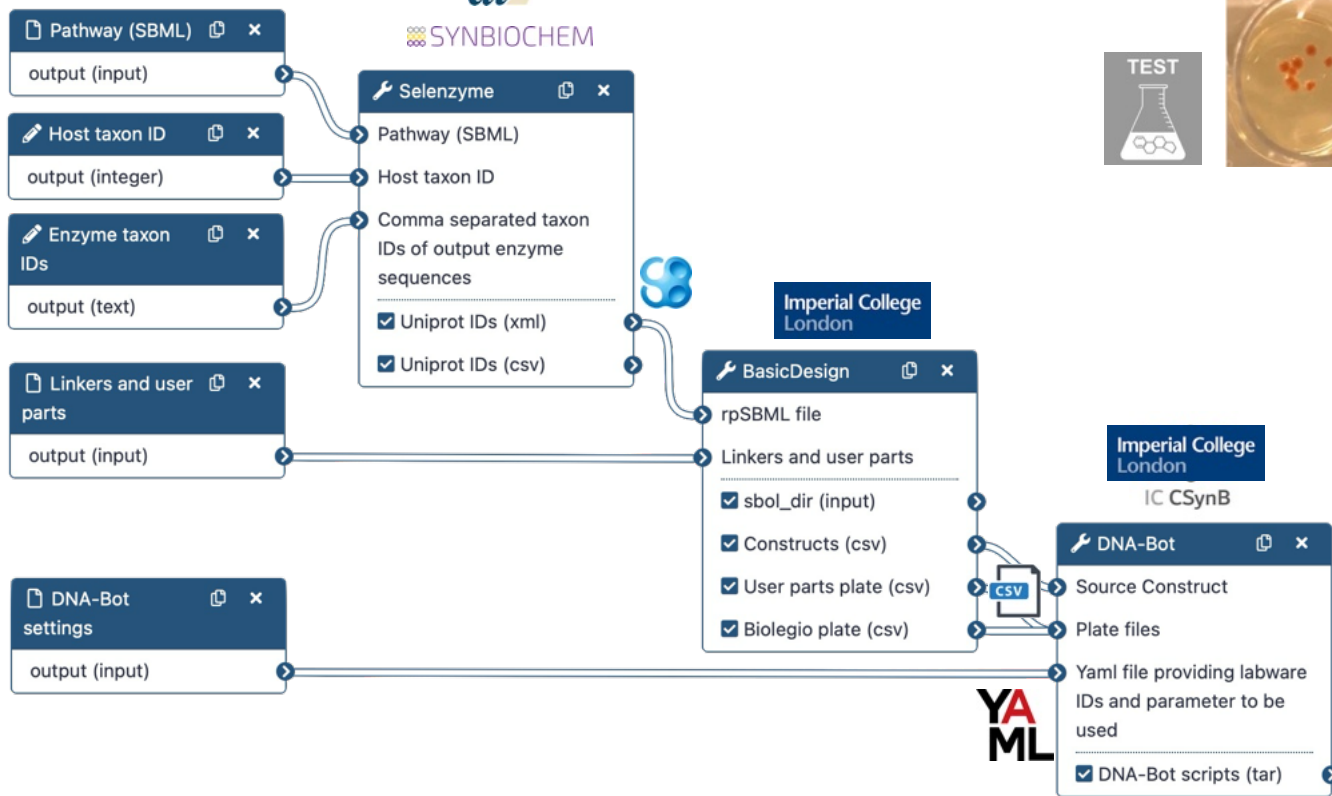
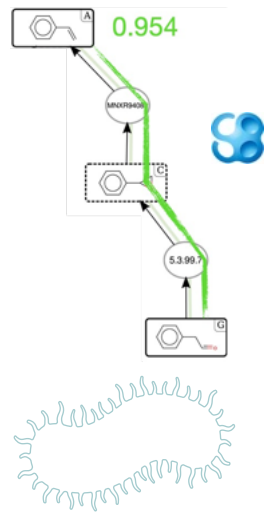
Neil Swainston, Mark Dunstan, Adrian J Jervis, Christopher J Robinson, Pablo Carbonell, Alan R Williams, Jean-Loup Faulon, Nigel S Scrutton, Douglas B Kell

Bioinformatics, Volume 34, Issue 13, 01 July 2018, Pages 2327–2329,

DNA-Weaver: optimal DNA assembly strategies via supply networks and shortest-path algorithms

V. Zulkower and S. Rosser
 IWBD, Cambridge (UK), 2019





ACS Synthetic Biology Letter
pubs.acs.org/synbio

BASIC: A New Biopart Assembly Standard for Idempotent Cloning Provides Accurate, Single-Tier DNA Assembly for Synthetic Biology

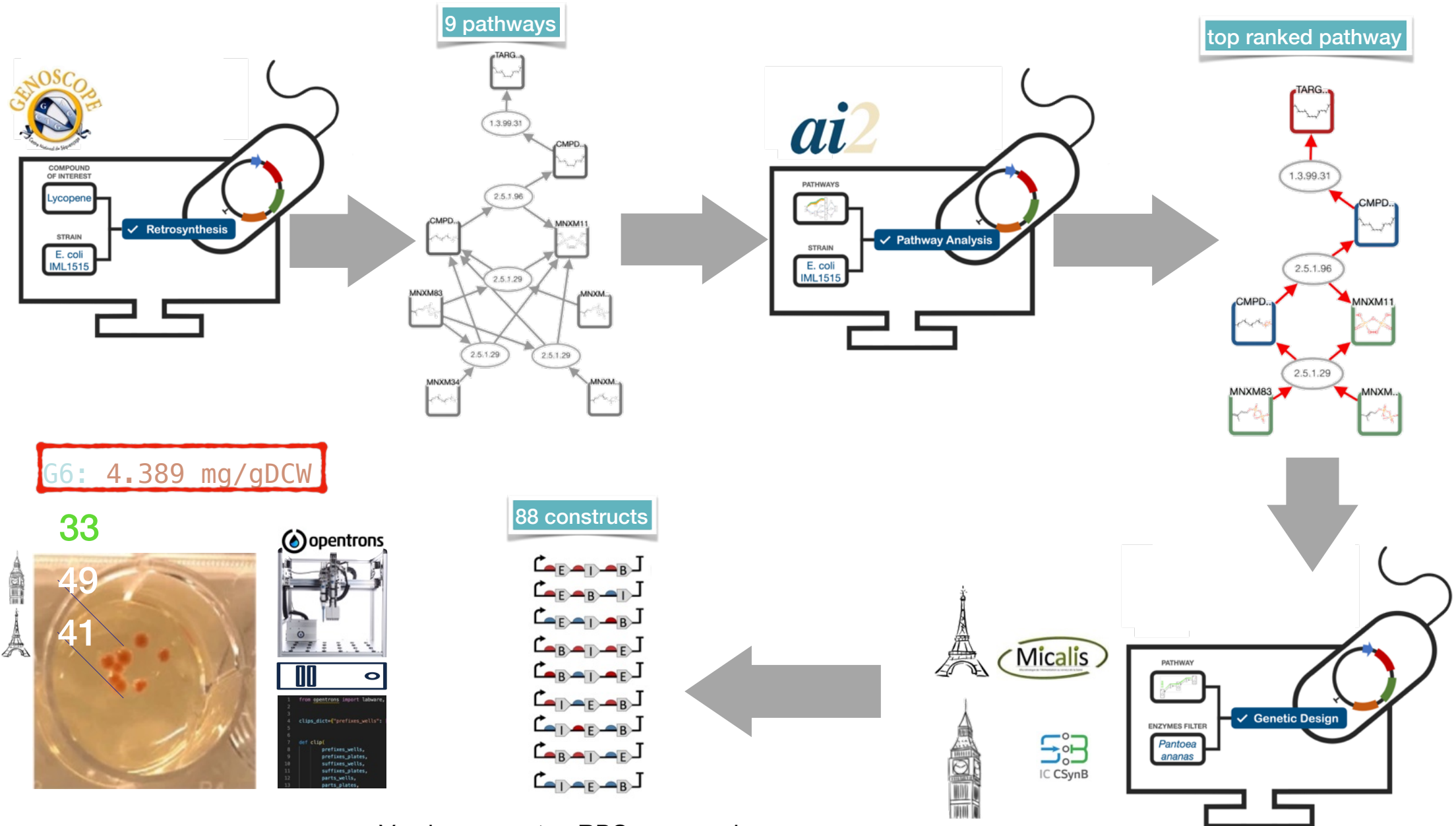
Marko Storch,^{†,||} Arturo Casini,^{†,||} Ben Mackrow,[†] Toni Fleming,[‡] Harry Trehwhitt,[†] Tom Ellis,[§] and Geoff S. Baldwin^{*†}

DNA-BOT: a low-cost, automated DNA assembly platform for synthetic biology

Marko Storch, Matthew C Haines, Geoff S Baldwin Author Notes

Synthetic Biology, Volume 5, Issue 1, 2020, ysaa010,

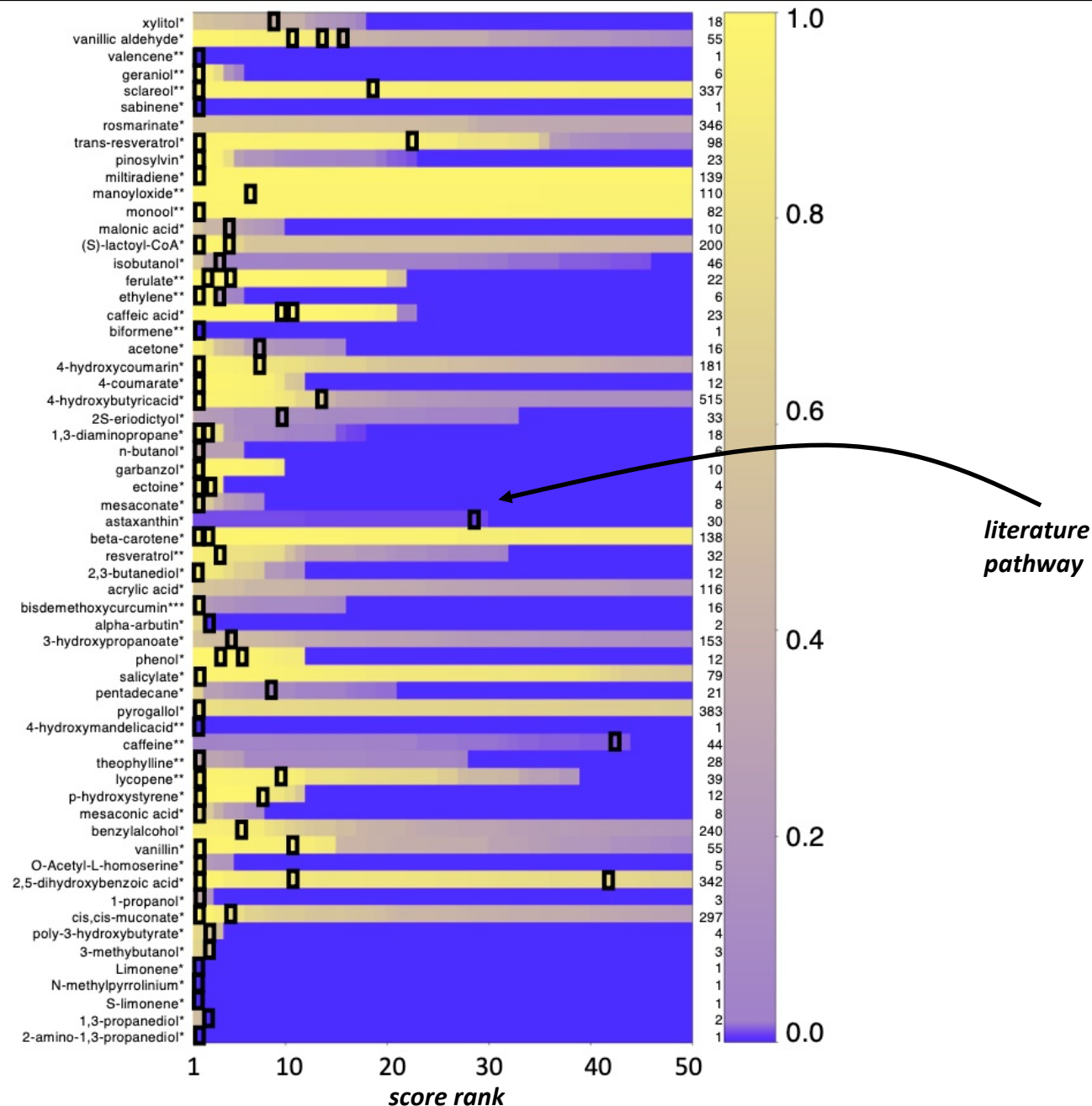
Benchmarking with lycopene production in *E. coli*



Varying promoter, RBS, gene order

Benchmarking with literature and expert users

- Compiled a list of 80 literature pathways for various compounds and strains
- Run workflow of the same compounds and strains, generated ~8000 pathways
- Rank each pathway using the ML scored trained on expert validation trial
- 83% (94%) success rate in retrieving the literature pathways among the top 10 (50) workflow generated pathways



Synthetic Biology

<https://www.youtube.com/watch?v=B1qJKWOe1PU>

Synthetic Biology is the engineering of biology : the deliberate (re)design and construction of novel biological and biologically based parts, devices and systems to perform new functions for useful purposes, that draws on principles elucidated from biology and engineering. By applying these principles to living systems, Synthetic Biology overcomes mimicry and optimisation-led research and introduces a rationale and systematic approach to the construction and (re)design. Synthetic Biology is at the intersection of engineering, bioscience, chemistry, and information technology. The goal of synthetic biology is to extend or modify the behavior of organisms and engineer them to perform new tasks. In this section, the tutorials shows how to design metabolic pathways for producing the desired chemical targets.











You can view the tutorial materials in different languages by clicking the dropdown icon next to the slides () and tutorial () buttons below.

Requirements

Before diving into this topic, we recommend you to have a look at:

- [Introduction to Galaxy Analyses](#)

Material

Lesson	Slides	Hands-on	Recordings	Input dataset	Workflows
Introduction to Synthetic Biology					
Designing plasmids encoding predicted pathways by using the BASIC assembly method					
Generating theoretical possible pathways for the production of Lycopene in E.Coli using Retrosynthesis tools					

SynBioCAD tools can be connected to many others in the ToolShed



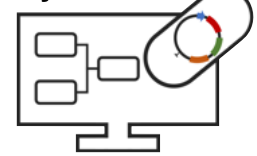
DATA AND PROCESS MANAGEMENT

ACTIVE LEARNING

Tool Shed	Nbr Tools
Machine Learning	62



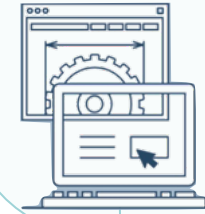
Galaxy SynBioCAD



Tool Shed	Nbr Tools
Synthetic Biology	59

DESIGN TOOLS

STRAIN DESIGN



Tool Shed	Nbr Tools
Systems Biology	41



nature communications

Explore content ▾ About the journal ▾ Publish with us ▾

nature > nature communications > articles > article

Article | Open Access | Published: 29 August 2022

The automated Galaxy-SynBioCAD pipeline for synthetic biology design and engineering

Joan Hérisson, Thomas Duigou, Melchior du Lac, Kenza Bazi-Kabbaj, Mahnaz Sabeti Azad, Gizem Buldum, Olivier Telle, Yorgo El Moubayed, Pablo Carbonell, Neil Swainston, Valentin Zulkower, Manish Kushwaha, Geoff S. Baldwin & Jean-Loup Faulon

TEST DATA ACQUISITION



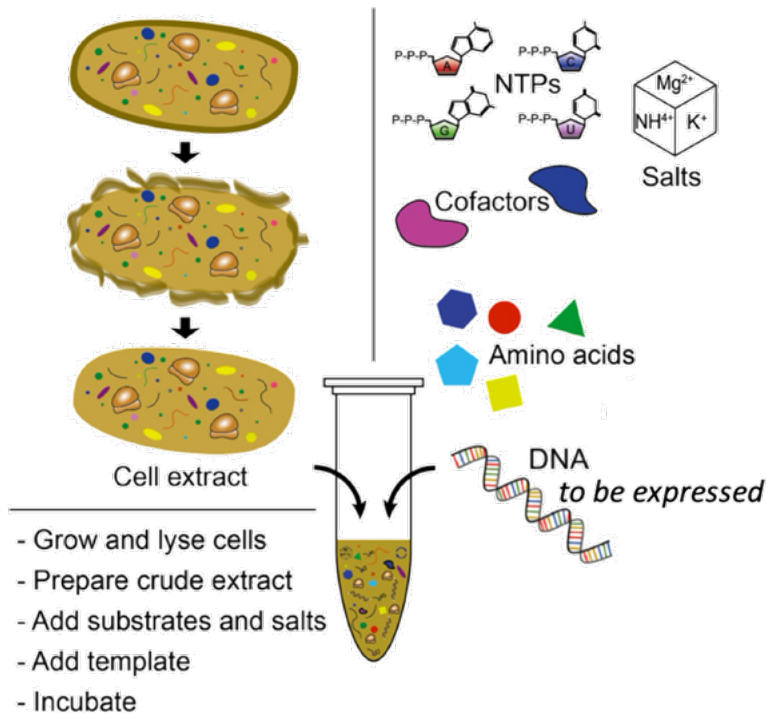
Tool Shed	Nbr Tools
Flow cytometry	45
Proteomics	431
Metabolomics	171

ROBOT DRIVERS



Active learning to optimize productivity in cell-free systems

Lysate-based cell-free systems (TXTL)



Reference composition
Sun Z.Z. et al. J. Vis. Exp. 2013

Component	Concentration			
Mg-glutamate (mM)	0.4	1.2	2	4
K-glutamate (mM)	8	24	40	80
Amino Acid (mM)	0.15	0.45	0.75	1.5
tRNA (mg/ml)	0.02	0.06	0.1	0.2
CoA (mM)	0.026	0.078	0.13	0.26
NAD (mM)	0.033	0.099	0.165	0.33
cAMP (mM)	0.075	0.225	0.375	0.75
Folinic Acid (mM)	0.0068	0.0204	0.034	0.068
Spermidine (mM)	0.1	0.3	0.5	1
3-PGA (mM)	3	9	15	30
NTP (mM)	0.15	0.45	0.75	1.5

Combinatorial space = 4^{11}
= 4 194 304 compositions



- Can we improve protein production without increasing the price of cell-free reaction?
- Can we provide efficient predictions of protein production *in vitro*?
- Can we highlight the critical parameters involve in protein production *in vitro*?

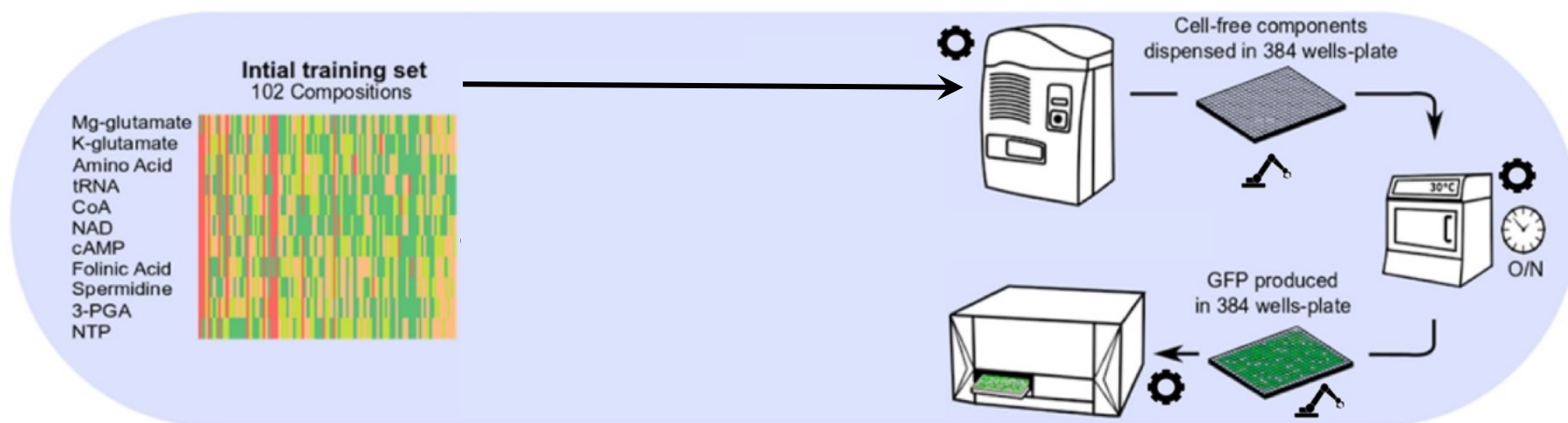
Active learning to optimize productivity in cell-free systems

- Set up an initial batch sampling the space of possible compositions



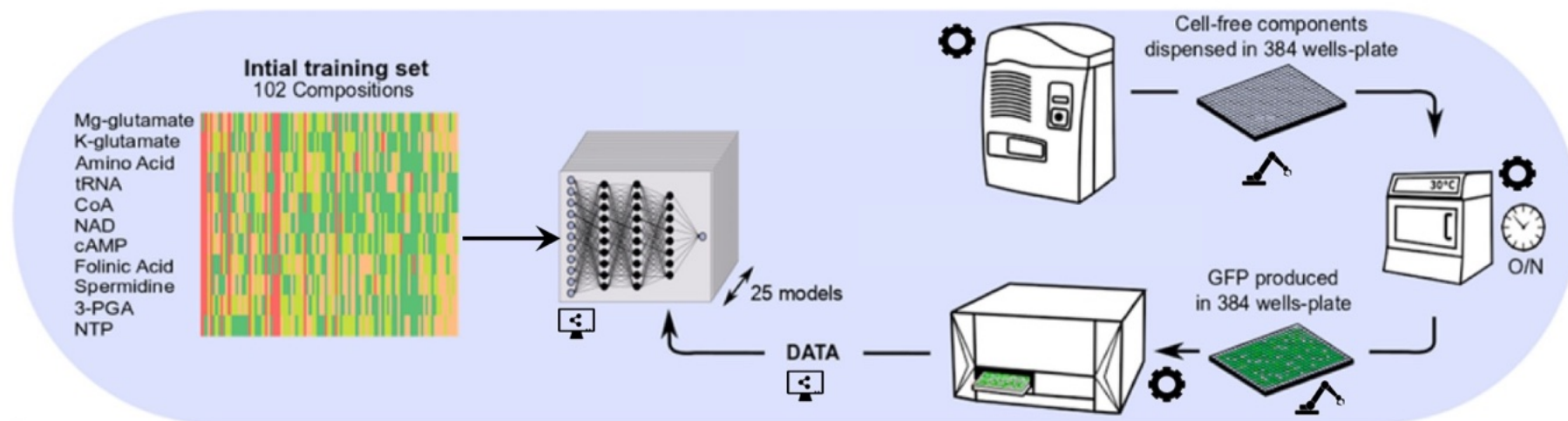
Active learning to optimize productivity in cell-free systems

- Set up an initial batch sampling the space of possible compositions
- **Measure yield level though fluorescence**



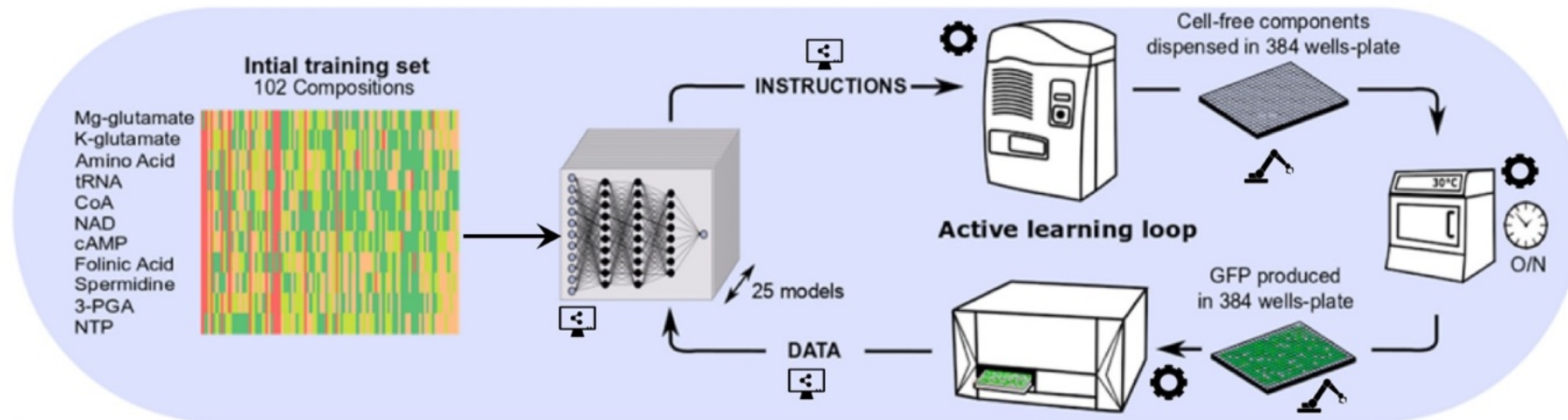
Active learning to optimize productivity in cell-free systems

- Set up an initial batch sampling the space of possible compositions
- Measure yield level through fluorescence
- **Develop a Neural Network model predicting yield from composition**



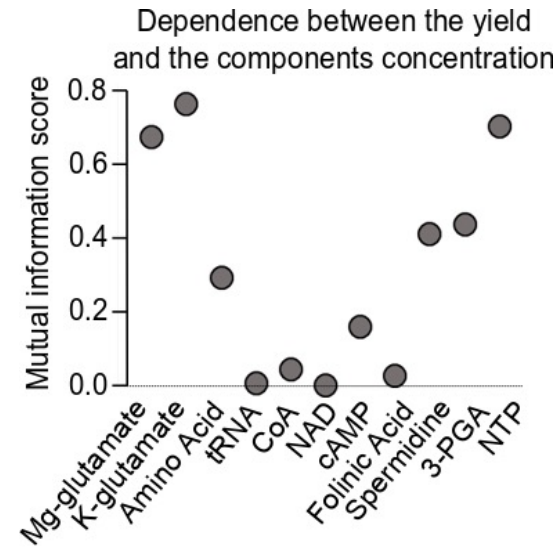
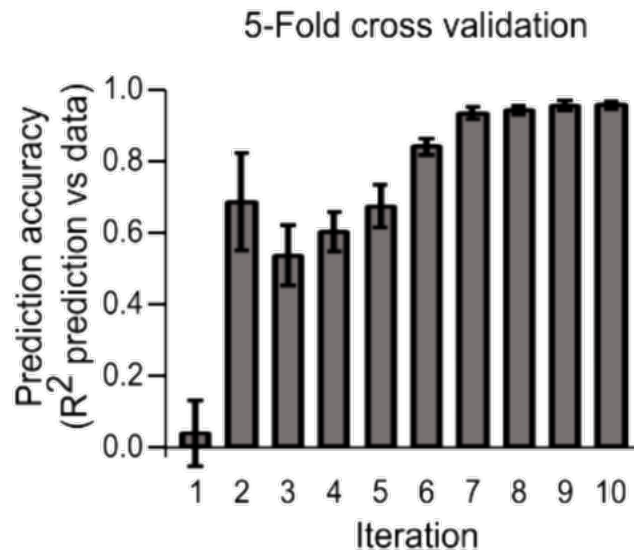
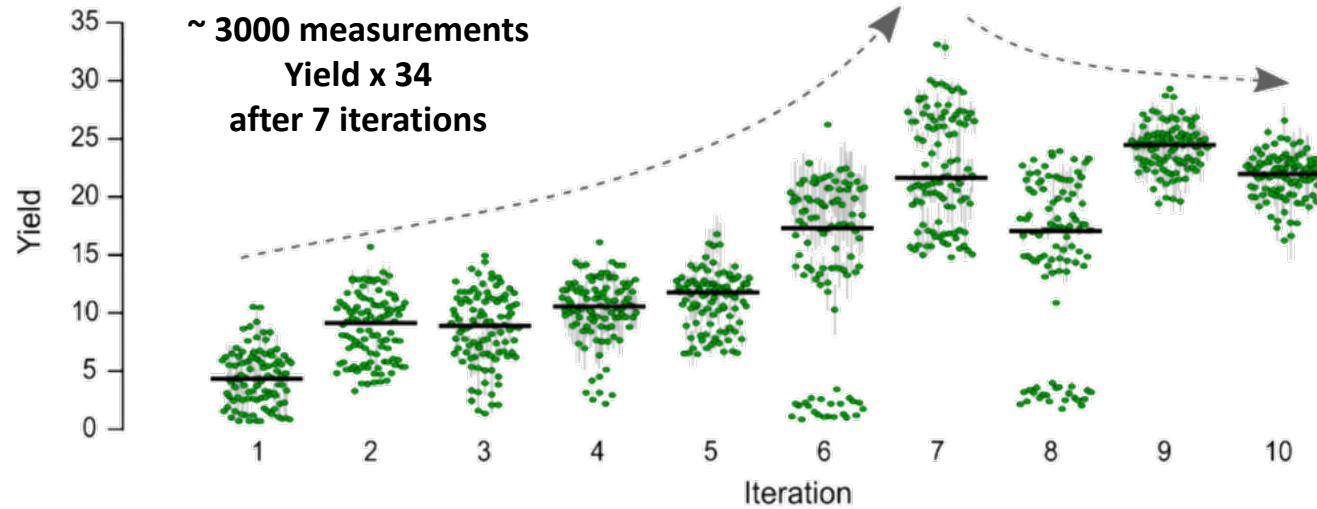
Active learning to optimize productivity in cell-free systems

- Set up an initial batch sampling the space of possible compositions
- Measure yield level through fluorescence
- Develop a Neural Network model predicting yield from composition
- **Use the model to predict the yield for each composition not yet tested**
- **Select next batch of compositions to be measured based on exploitation vs. exploration (UCB formula)**
- Repeat

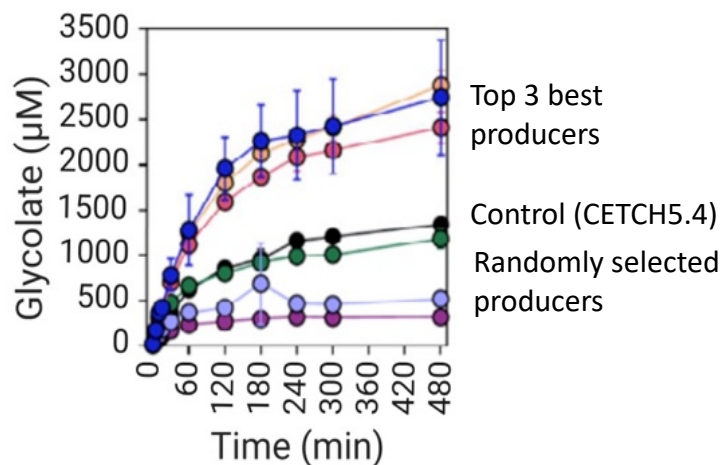
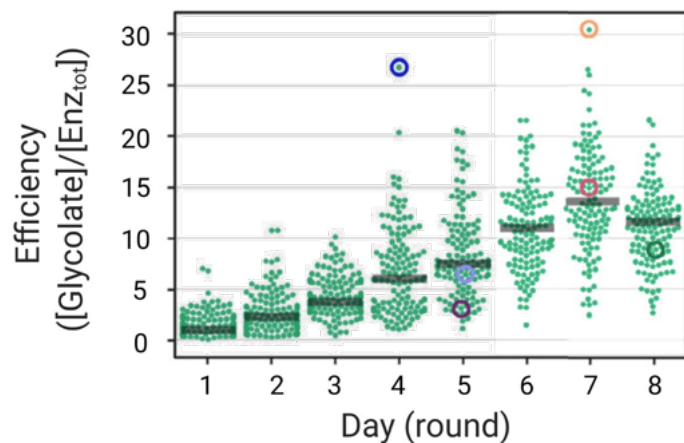
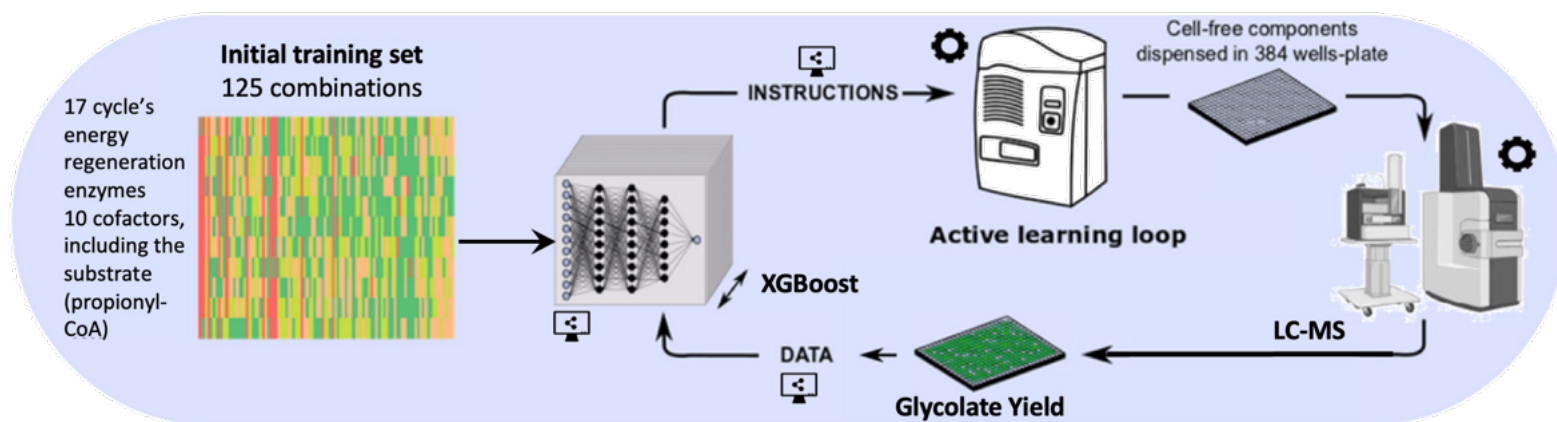
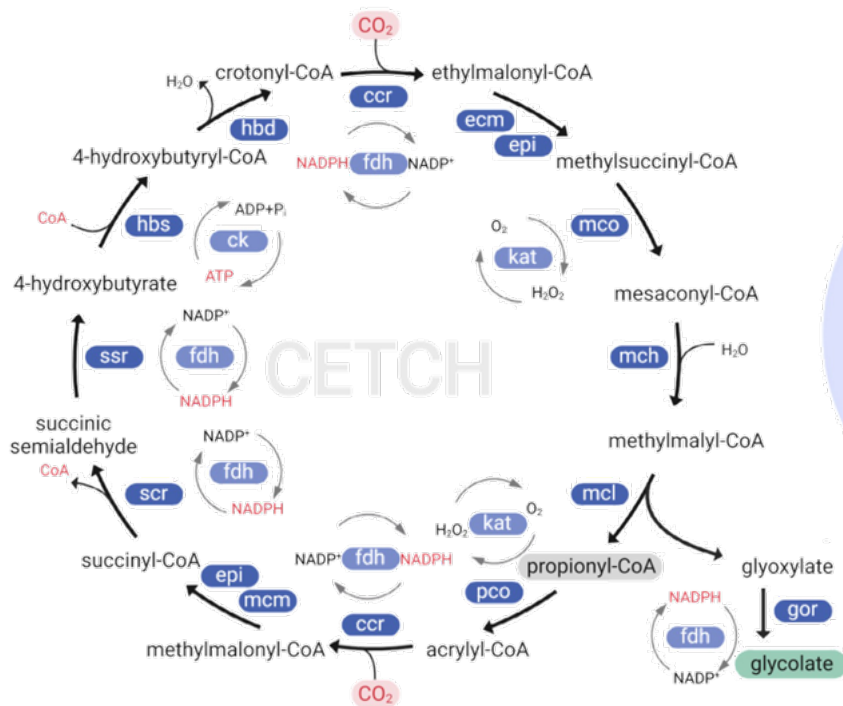


* UCB: Take the compositions having the top values $\mu + \sqrt{2} \sigma$, where μ is the mean predicted yield and σ the standard dev
Picking high μ values favor exploitation picking high σ values favor exploration

Active learning to optimize productivity in cell-free systems

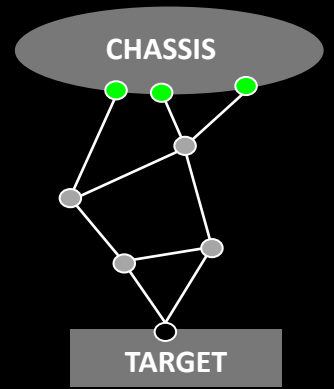


Active learning to optimize metabolic pathways in cell-free systems

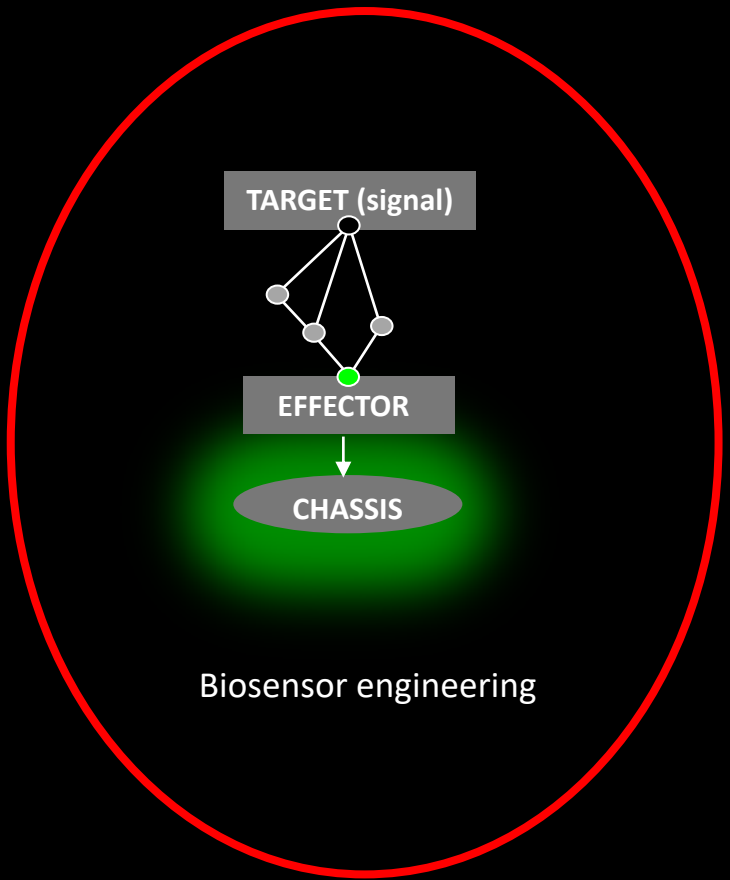


6x more efficient than the best in vitro CO₂-fixing system described to date (CETCH 5.4, Schwander *et al.* *Science* 2016)

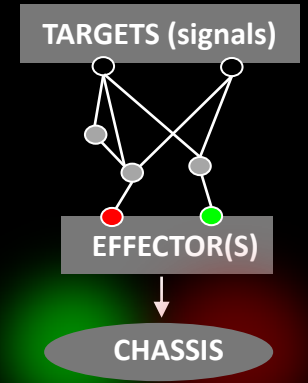
Engineering metabolic and genetic networks in vitro (cell-free) and in vivo (E. coli)



Metabolic engineering



Biosensor engineering



Information processing
biocomputing

Designing biosensing circuits

Detectable Metabolites DB

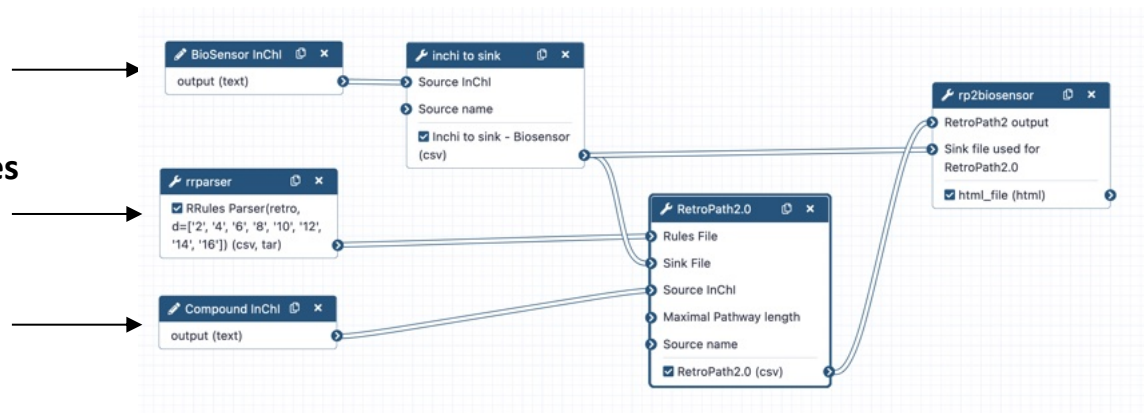
(Koch et al. DIB 2018)

Transformations rules

RetroRules.org

(Duigou et al. NAR, 2018)

Molecule to detect



nature communications

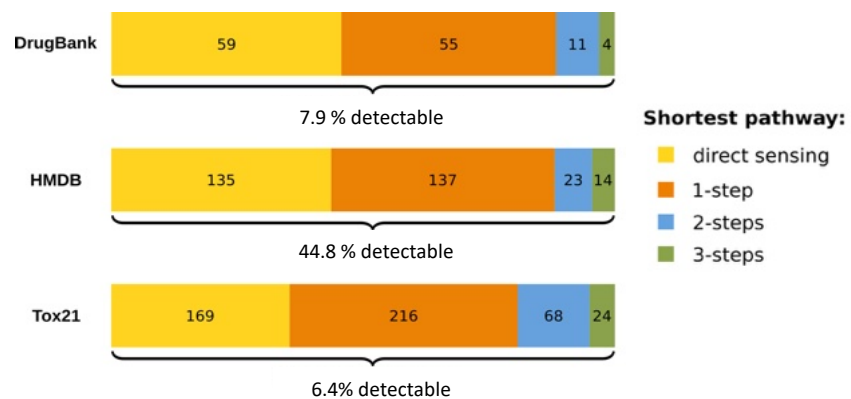
Explore content About the journal Publish with us

nature > nature communications > articles > article

Article | Open Access | Published: 29 August 2022

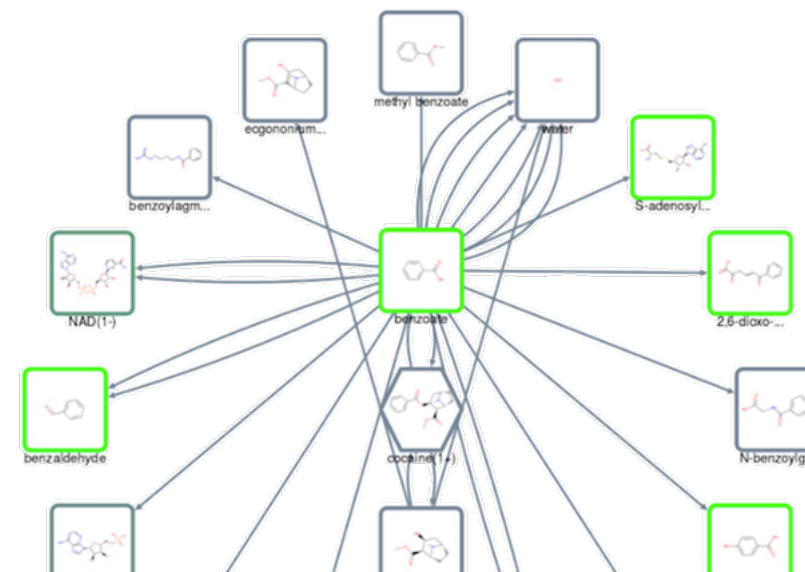
The automated Galaxy-SynBioCAD pipeline for synthetic biology design and engineering

Joan Hérisson, Thomas Duigou, Melchior du Lac, Kenza Bazi-Kabbaj, Mahnaz Sabeti Azad, Gizem Buldum, Olivier Telle, Yorgo El Moubayed, Pablo Carbonell, Neil Swainston, Valentin Zulkower, Manish Kushwaha, Geoff S. Baldwin & Jean-Loup Faulon

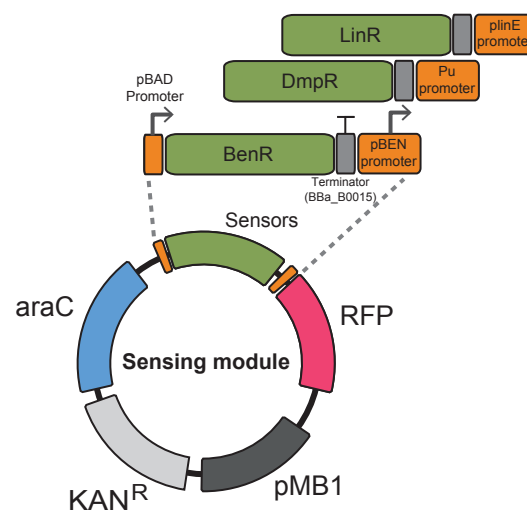
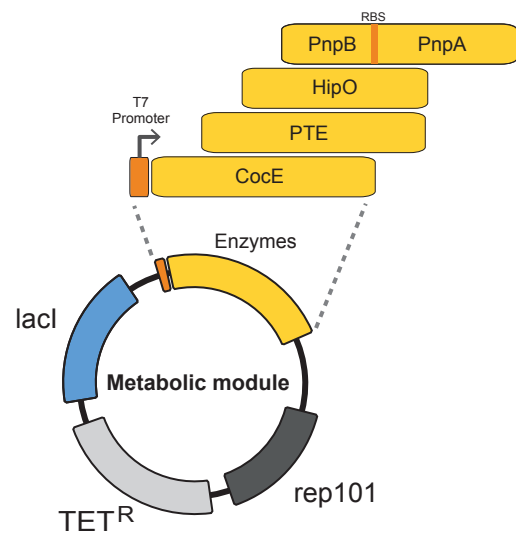
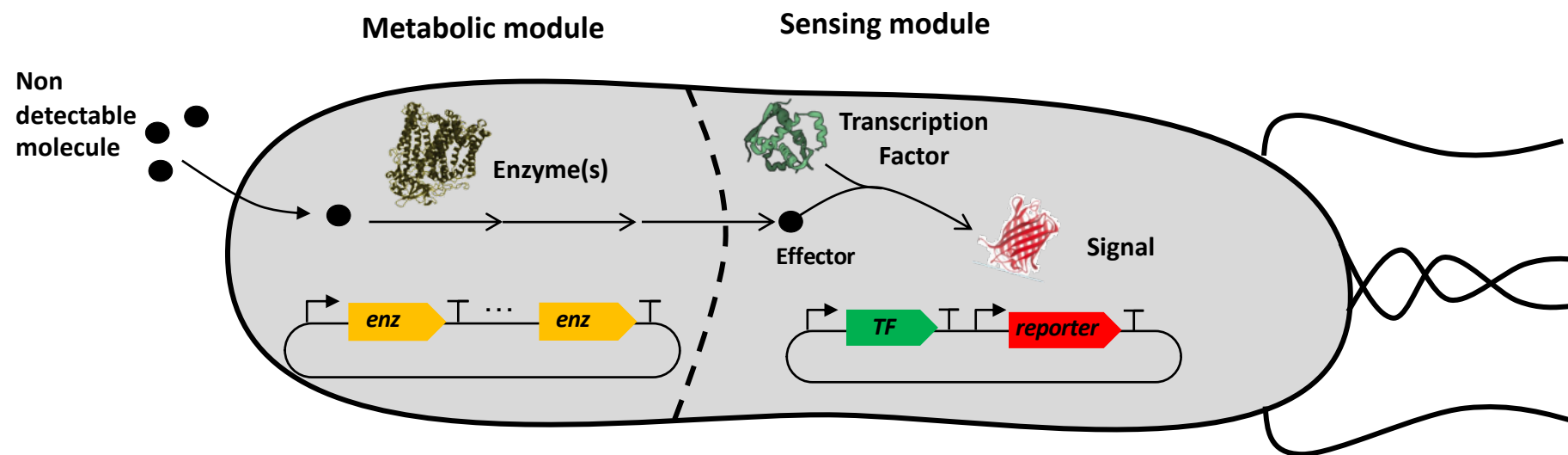


- Delepine B, et al. *NAR* 2016

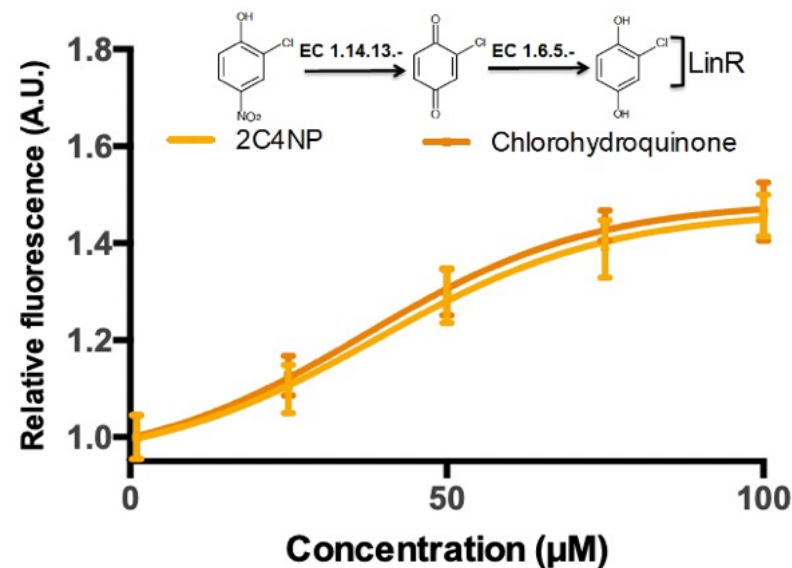
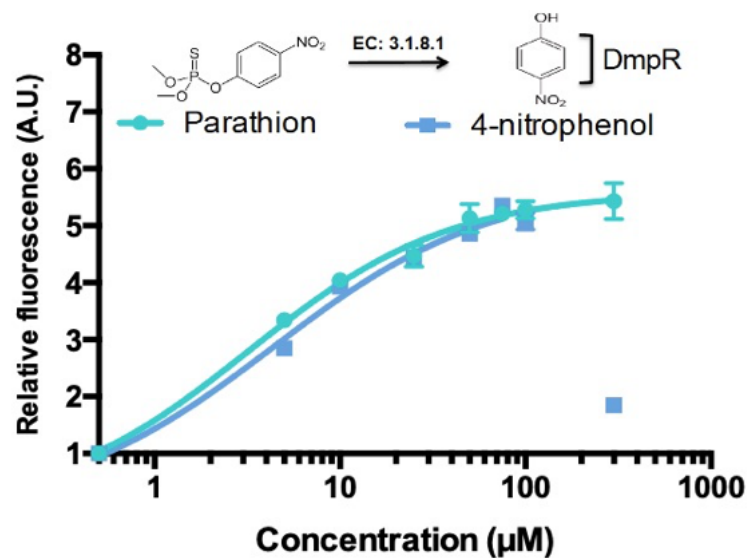
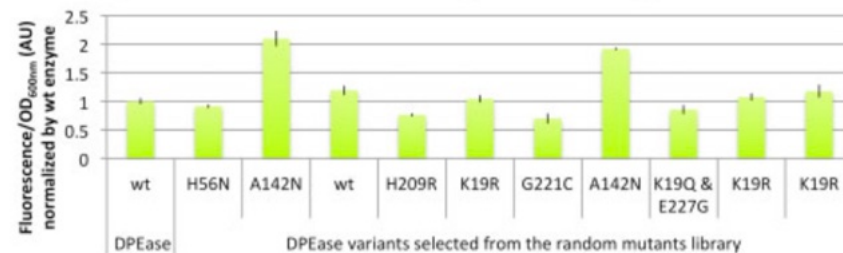
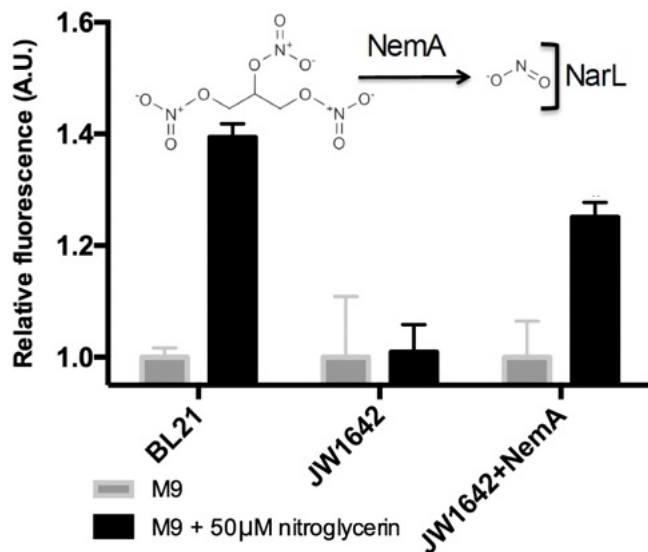
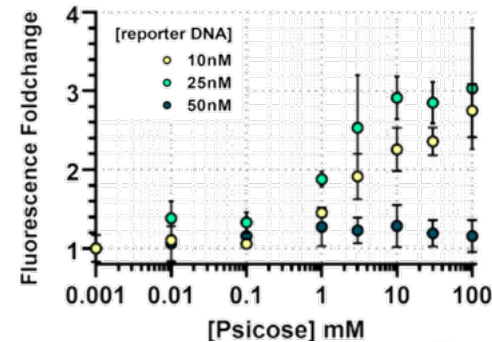
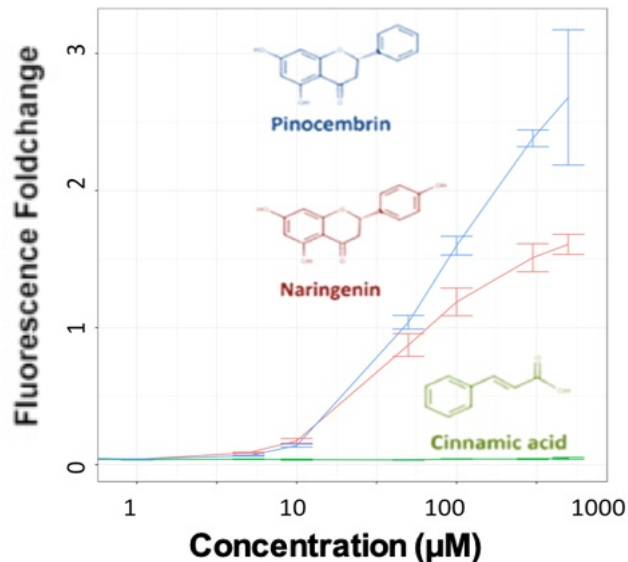
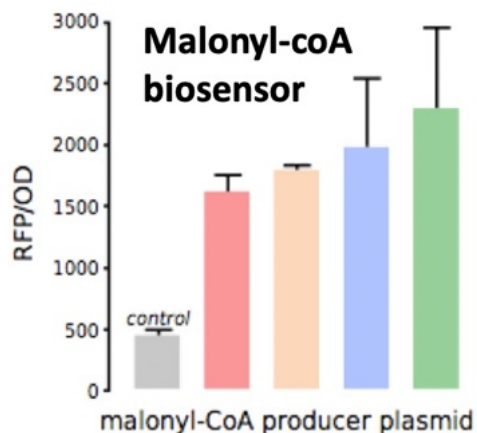
SensiPath v2.1.2



Engineering biosensing circuits

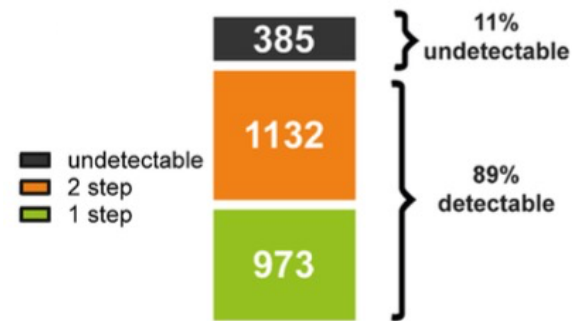


Characterizing biosensing circuits

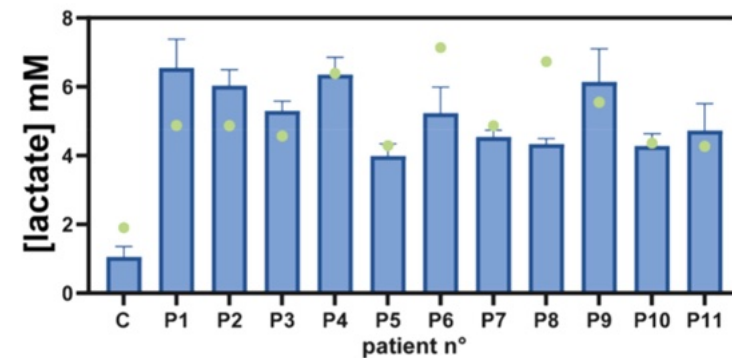
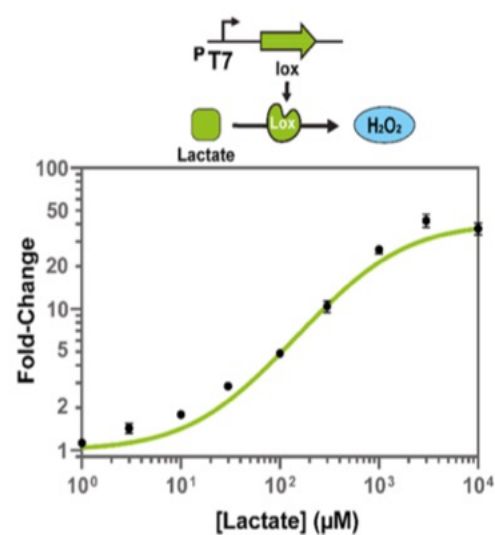
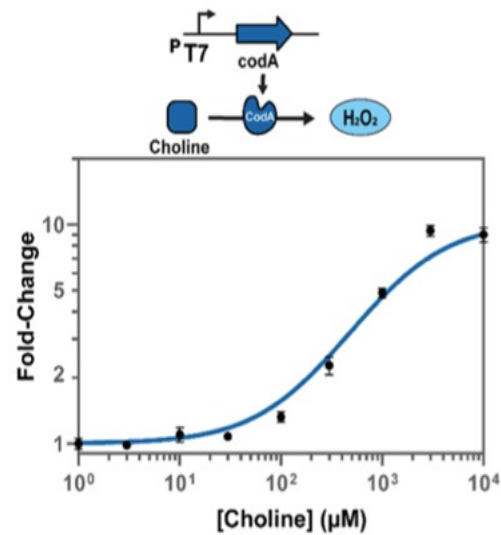
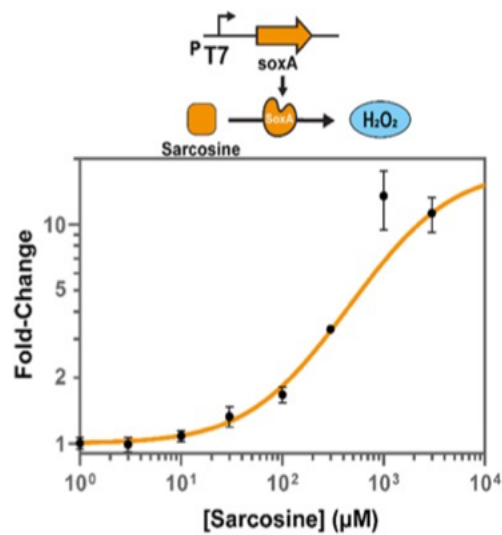
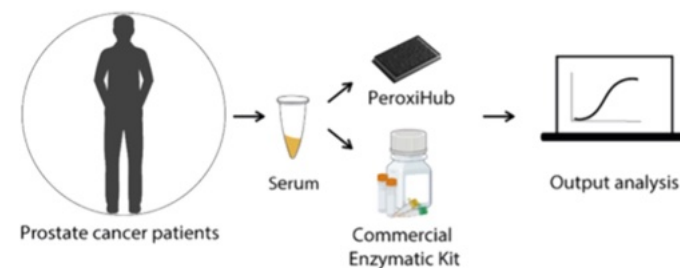


- Feher T *et al.* *Frontier in Bioeng. & Biotech*, 2015
- Libis V. *et al.* *Curr Opin Microbiol*, 2016
- Trabelsi H. *et al.* *Biotech. & Bioeng.*, 2018
- Koch M. *et al.*, *Frontiers in Bioeng & Biotech.*, 2018
- Koch M. *et al.* *Curr Opin Biotech*, 2019
- Armetta J. *et al.* *Synthetic Biology* 2019 & Pandi A. *et al.* *ACS Synth Biol*, 2019

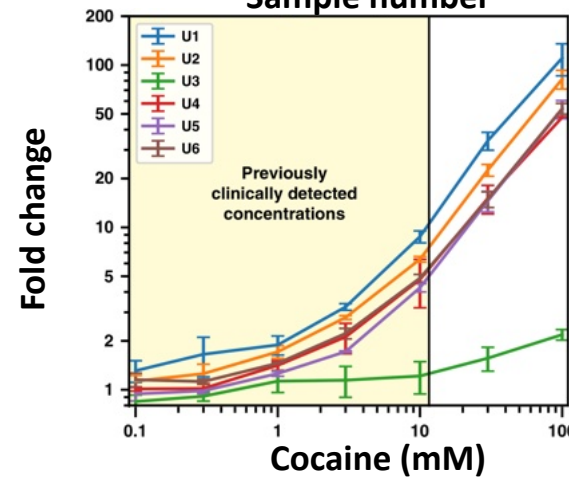
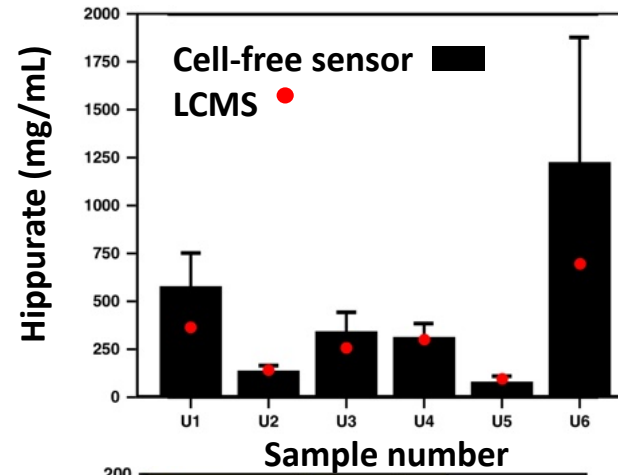
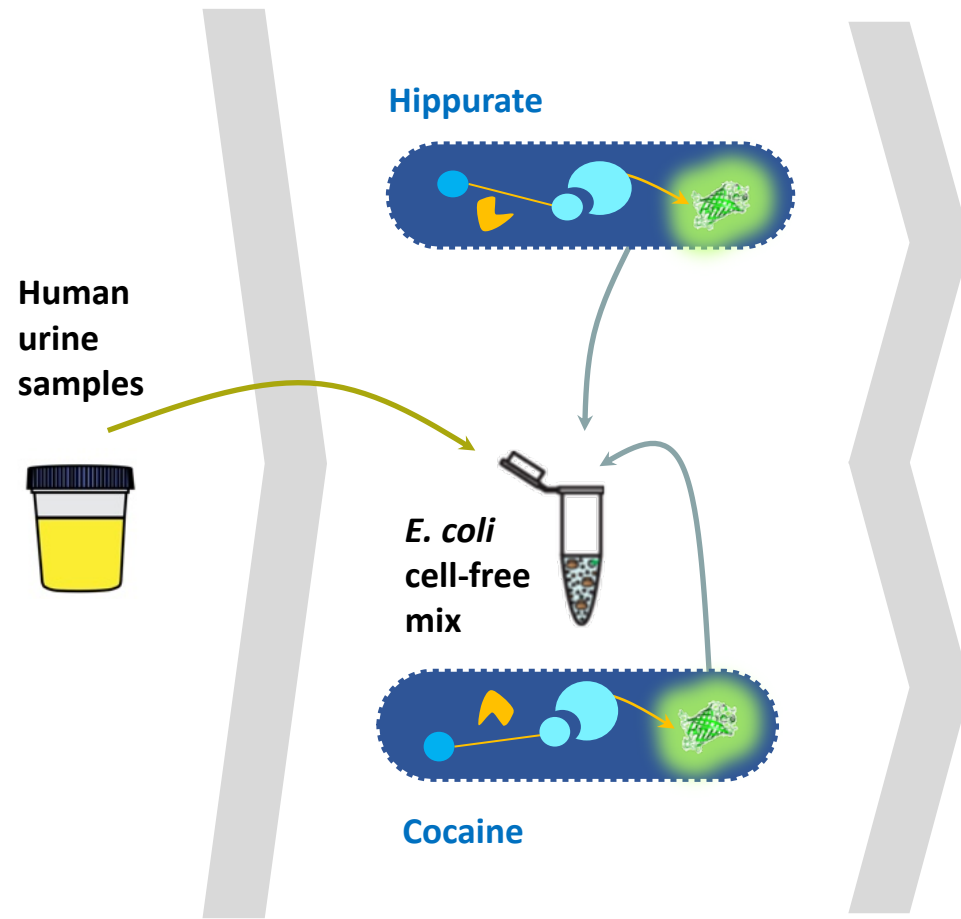
Cell-free biosensors with clinical samples



2490 biomarkers from HMDB



Cell-free biosensors with clinical samples

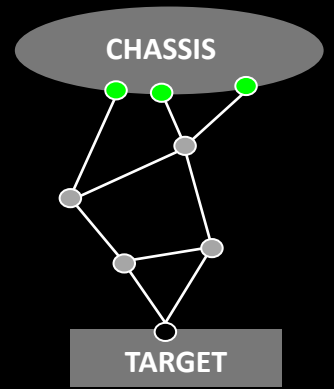


Hippurate is a degradation product of a probiotic treatment of *Phenylketonuria*

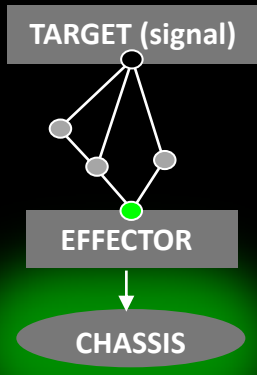


can be freeze dry

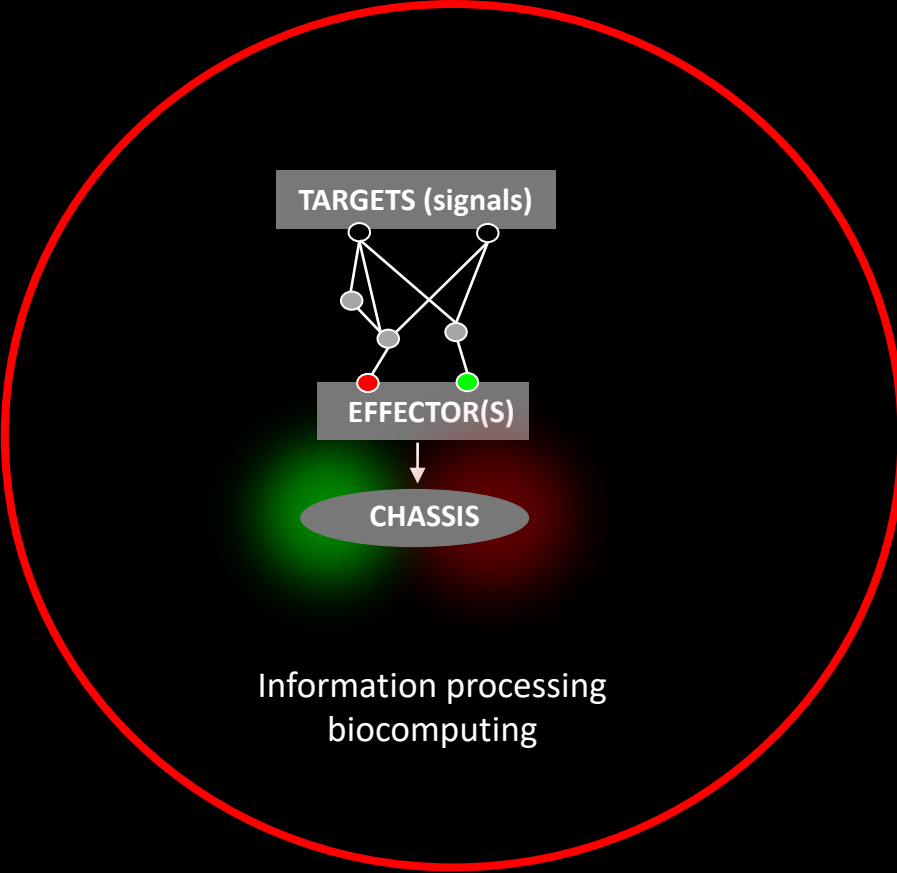
Engineering metabolic and genetic networks *in vitro* (cell-free) and *in vivo* (*E. coli*)



Metabolic engineering



Biosensor engineering



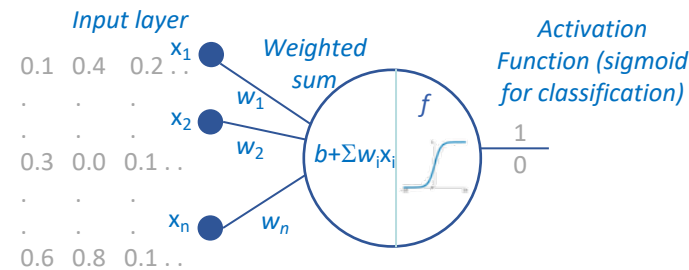
Information processing
biocomputing

Engineering a metabolic perceptron: why?

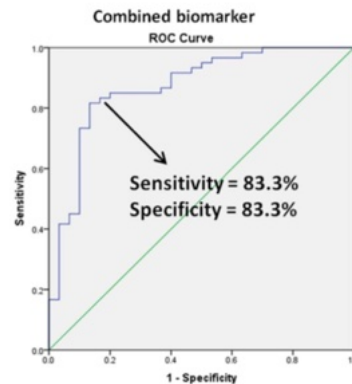
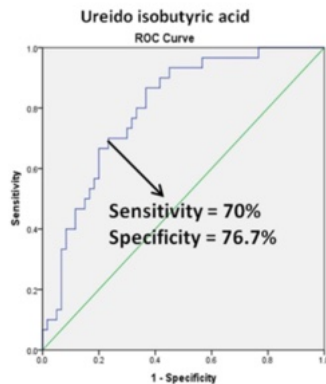


TRAINING THE NETWORK

Perceptron weights (w_i) are learned to increase classifier accuracy



Prostate cancer PCa metabolic score = $b + \sum_{j=1}^j w_j x_{ij}$



- Zang, et al. *PLoS One* 2013 and *J Proteome Res.* 2014
- Shen B, et al. *Cell.* 2020
-



USING THE TRAINED NETWORK

To perform a diagnostic:

- Quantify a panel of biomarkers (metabolites) on clinical samples (using metabolomics)
- Feed measured biomarkers concentrations (x_i) to

$$f(b + \sum w_i x_i)$$

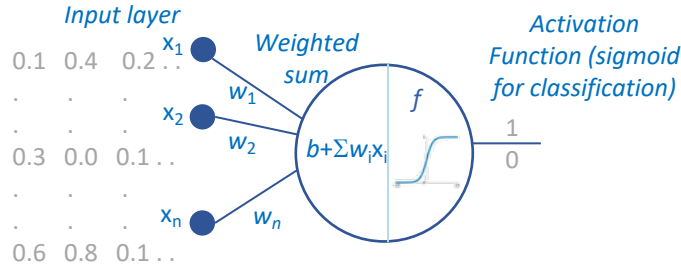
- Is it possible to avoid biomarker concentration measurements?
 - Engineer the trained network *in vitro* or *in vivo* and directly use it on clinical samples

Engineering a metabolic perceptron: the concept

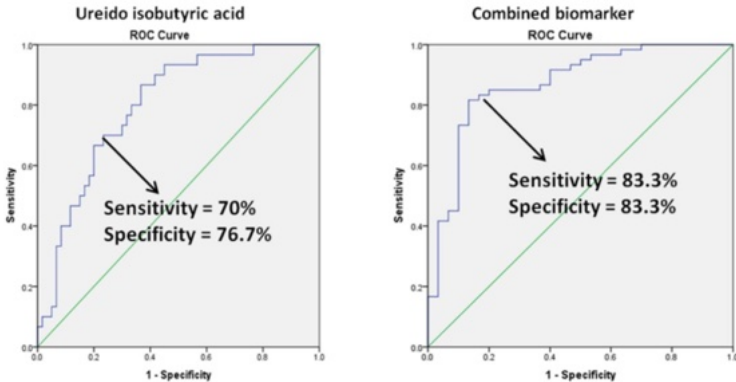


TRAINING THE NETWORK

Perceptron weights (w_i) are learned to increase classifier accuracy

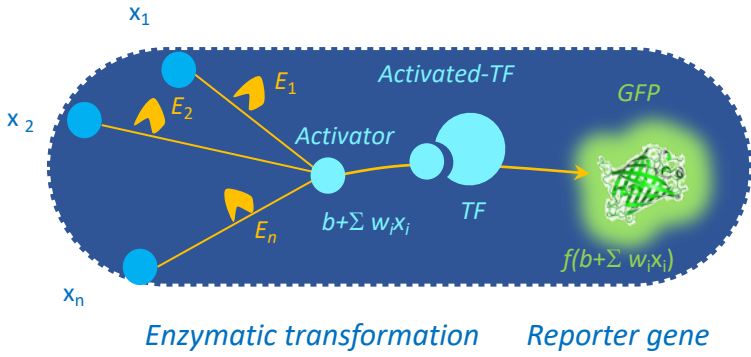


Prostate cancer PCa metabolic score = $b + \sum_{j=1}^j w_j x_{ij}$



ENGINEERING THE TRAINED NETWORK

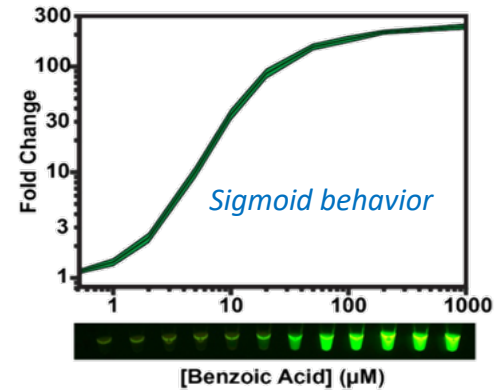
Need to actuate weighted sum and activation function



In theory (Michaelis-Menten) when $x_i \ll [E_i]$:

$$d[p] = \sum k_i [E_i] x_i dt$$

$w_i = k_i [E_i]$
where k_i is a kinetics constant



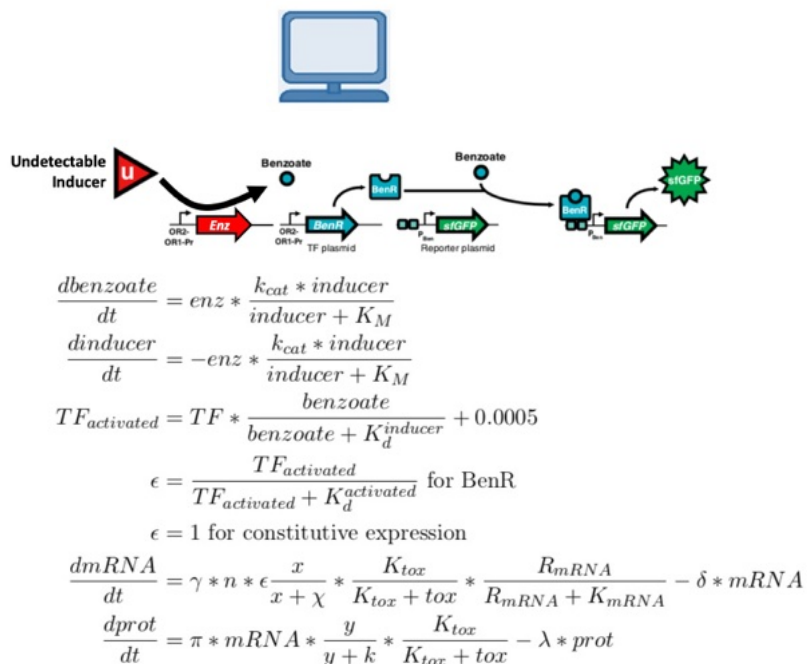
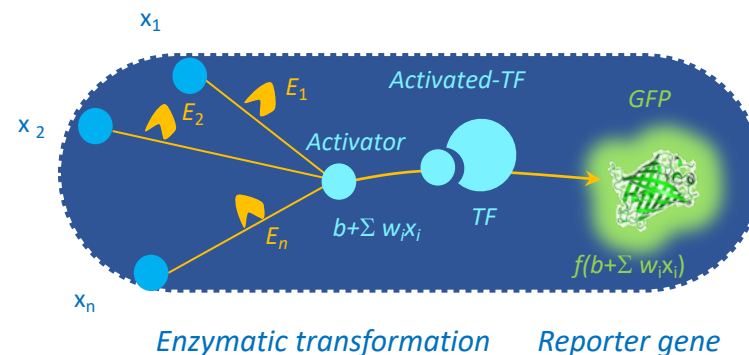
- Zang, et al. *PLoS One* 2013 and *J Proteome Res.* 2014
- Shen B, et al. *Cell.* 2020
-

Engineering a metabolic perceptron: the concept



ENGINEERING THE TRAINED NETWORK

Need to actuate weighted sum and activation function



$$\frac{d\text{benzoate}}{dt} = \text{enz} * \frac{k_{\text{cat}} * \text{inducer}}{\text{inducer} + K_M}$$

$$\frac{d\text{inducer}}{dt} = -\text{enz} * \frac{k_{\text{cat}} * \text{inducer}}{\text{inducer} + K_M}$$

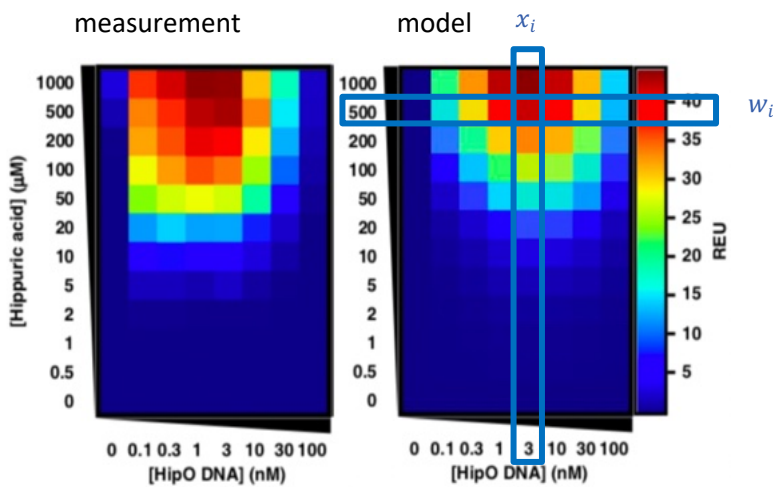
$$TF_{\text{activated}} = TF * \frac{\text{benzoate}}{\text{benzoate} + K_d^{\text{inducer}}} + 0.0005$$

$$\epsilon = \frac{TF_{\text{activated}}}{TF_{\text{activated}} + K_d^{\text{activated}}} \text{ for BenR}$$

$$\epsilon = 1 \text{ for constitutive expression}$$

$$\frac{dmRNA}{dt} = \gamma * n * \epsilon * \frac{x}{x + \chi} * \frac{K_{\text{tox}}}{K_{\text{tox}} + \text{tox}} * \frac{R_{mRNA}}{R_{mRNA} + K_{mRNA}} - \delta * mRNA$$

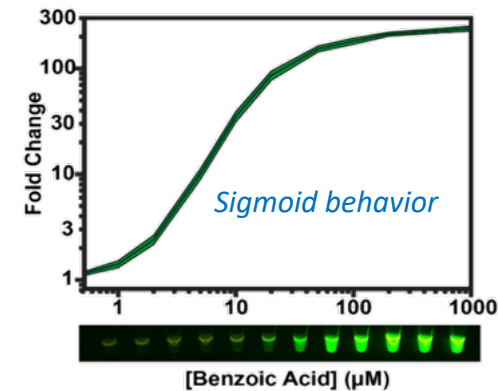
$$\frac{d\text{prot}}{dt} = \pi * mRNA * \frac{y}{y + k} * \frac{K_{\text{tox}}}{K_{\text{tox}} + \text{tox}} - \lambda * \text{prot}$$



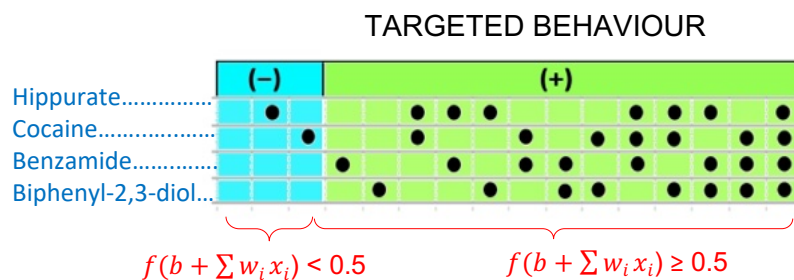
In theory (Michaelis-Menten)
when $x_i \ll [E_i]$:

$$d[p] = \sum k_i [E_i] x_i dt$$

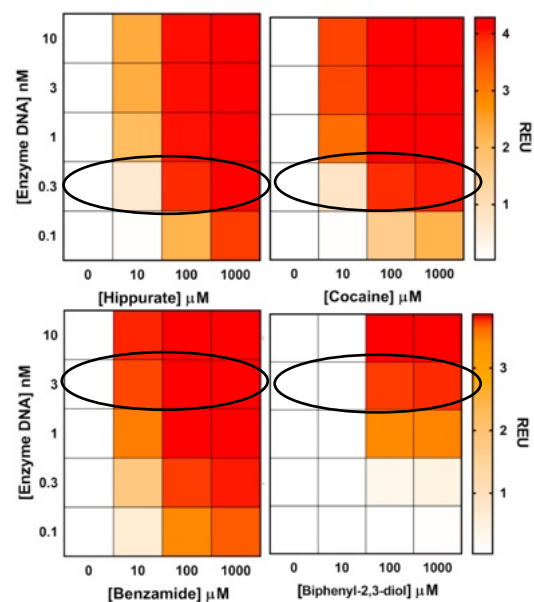
$w_i = k_i [E_i]$
where k_i is a kinetics
constant



Engineering a metabolic perceptron



- Kinetics model is used to compute the enzyme concentration for each weight



Logistic regression

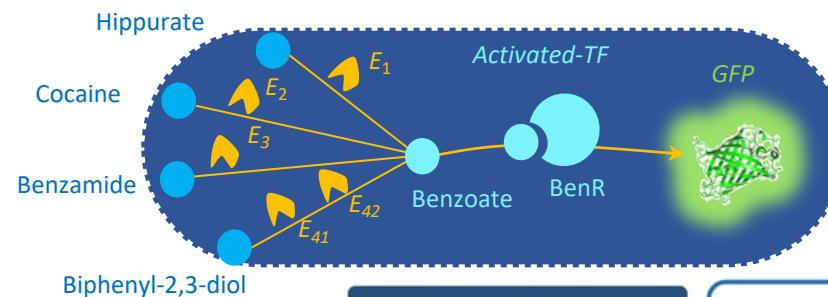
$$f(b + \sum w_i x_i)$$

$$b = -0.50$$

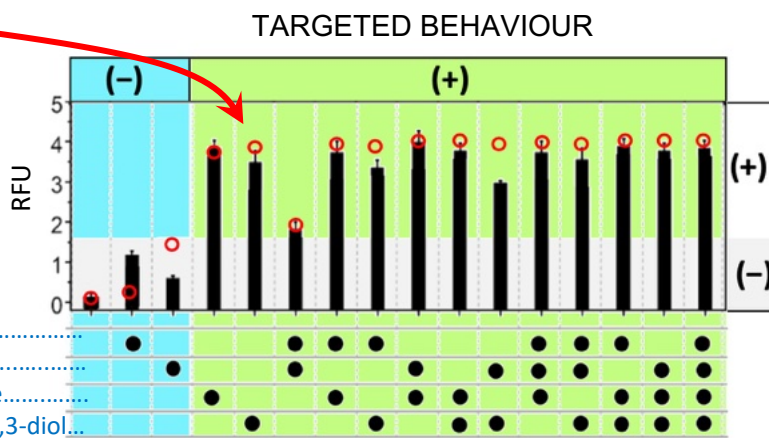
$$w_1 = 0.25$$

$$w_2 = 0.25$$

$$w_3 = 0.50$$

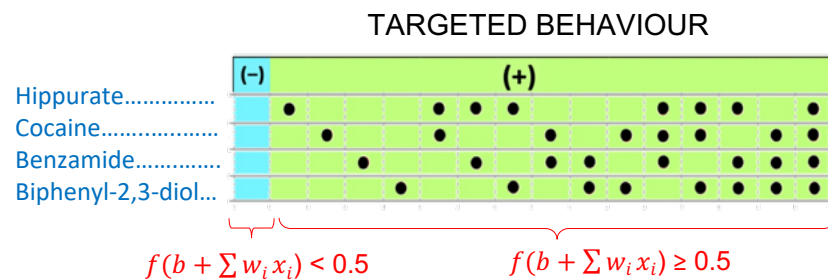
$$w_4 = 0.50$$


Retrosynthesis Workflow »

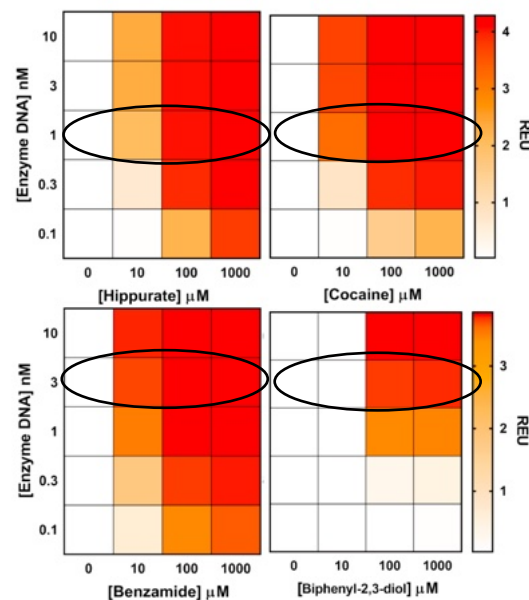


OBSERVED BEHAVIOUR

Engineering a metabolic perceptron

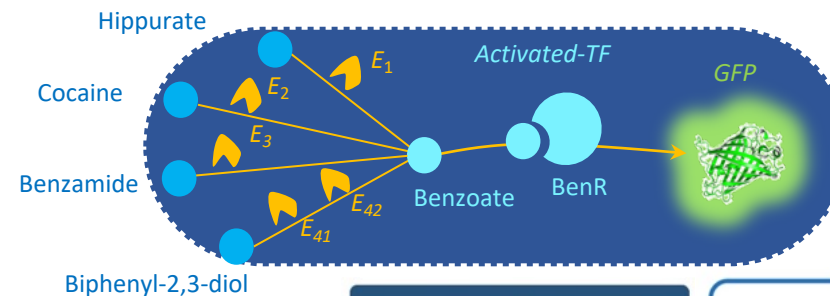


- Kinetics model is used to compute the enzyme concentration for each weight

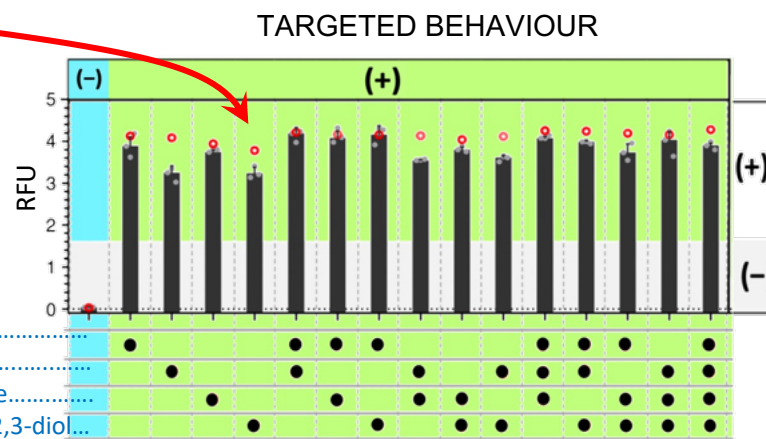


Logistic regression

$f(b + \sum w_i x_i)$
 $b = -0.50$
 $w_1 = 0.25$
 $w_2 = 0.25$
 $w_3 = 0.50$
 $w_4 = 0.50$



Retrosynthesis Workflow » Galaxy



OBSERVED BEHAVIOUR

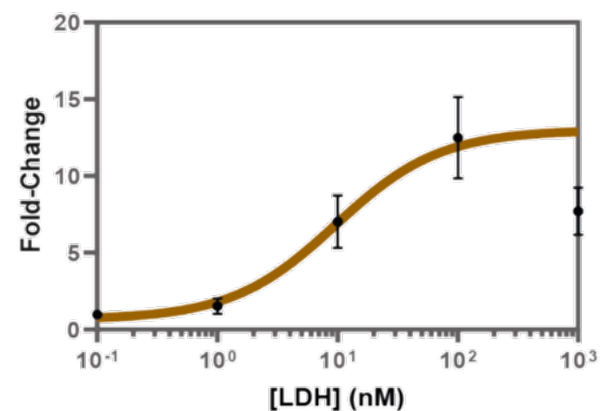
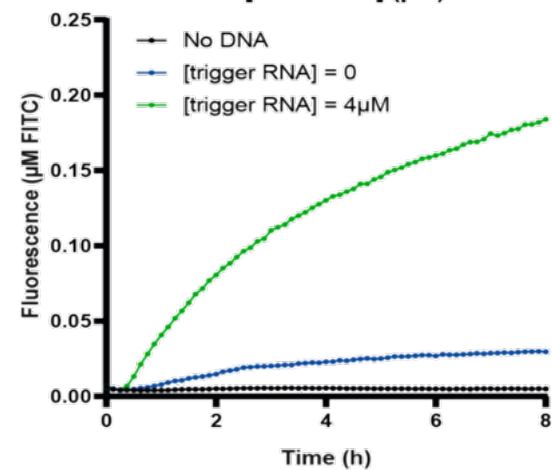
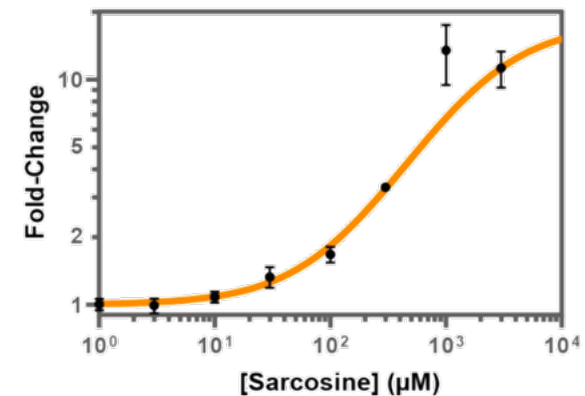
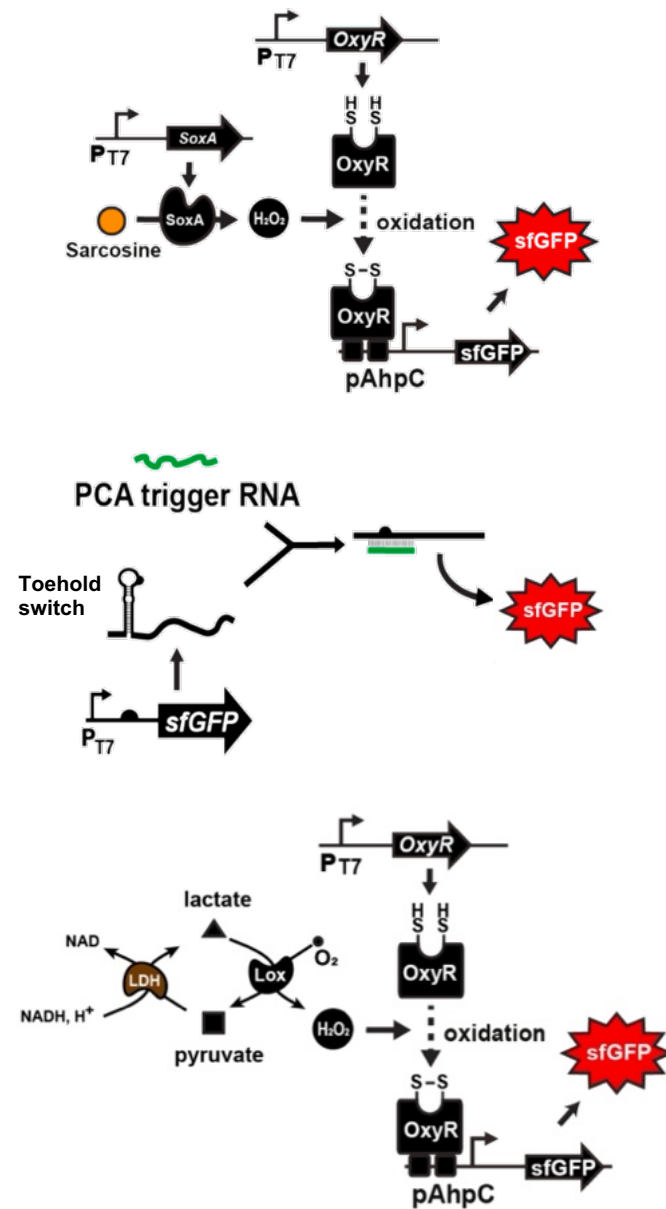
Toward engineering a multimodal perceptron

Metabolite
(sarcosine)

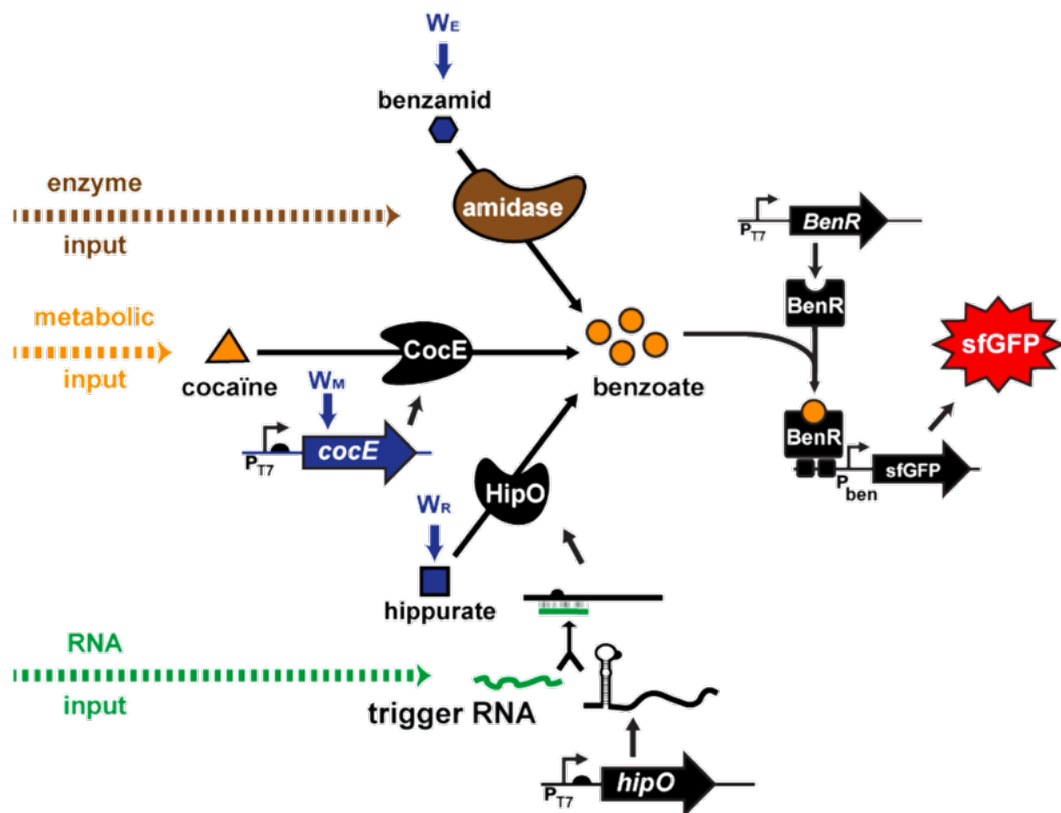
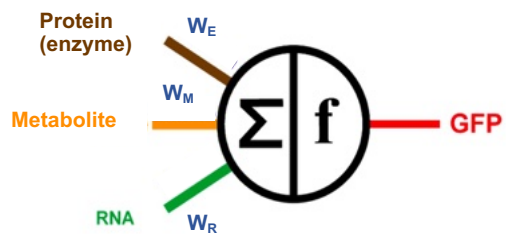
RNA
(PCA3)

Protein
(LDH)

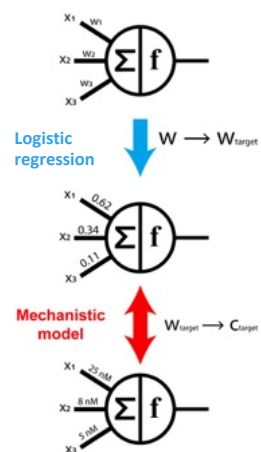
prostate
cancer
biomarkers



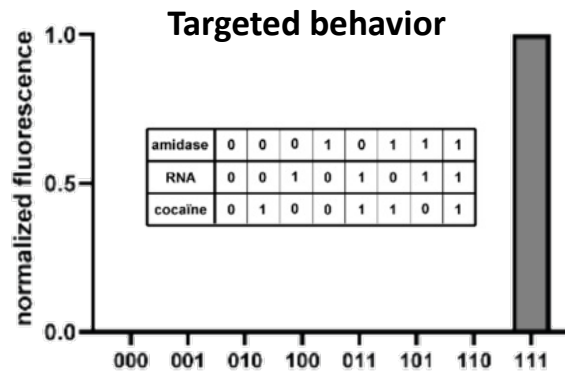
Engineering a multimodal perceptron



MODEL BASED APPROACH



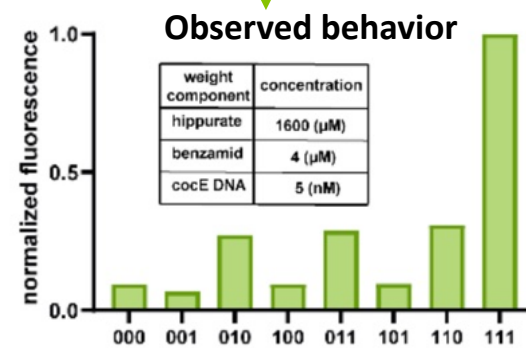
- Pandi A. *et al. Nature Comm.* 2019



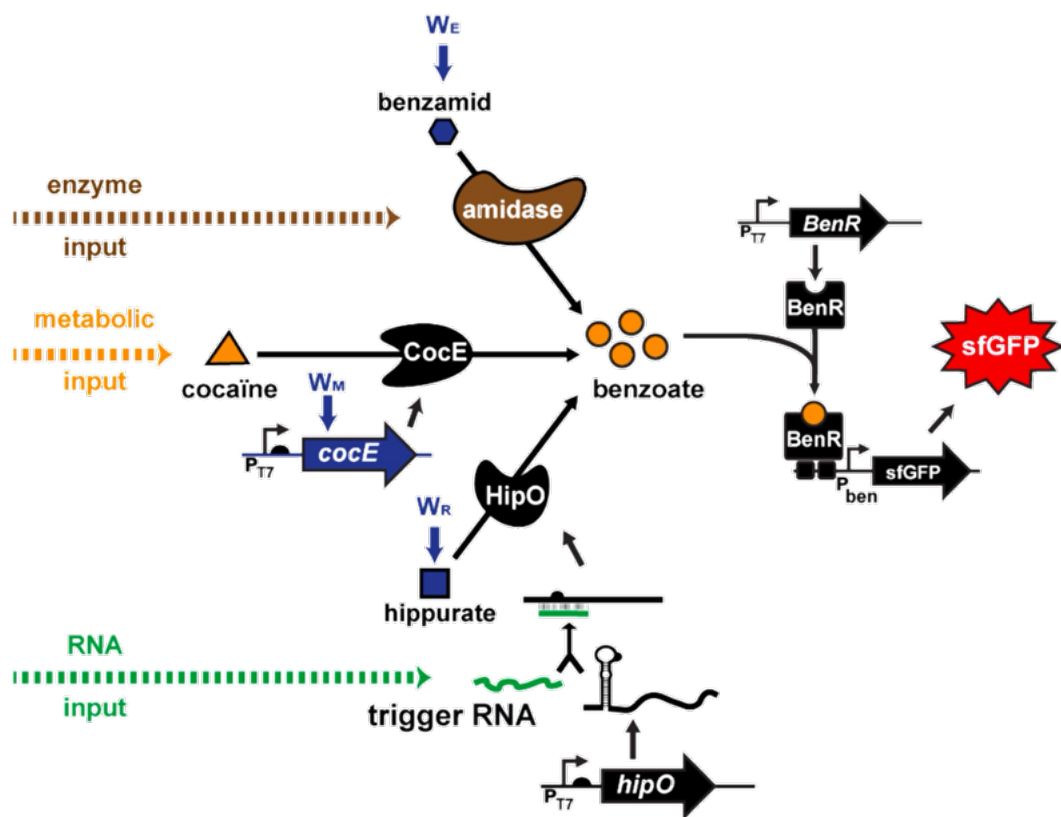
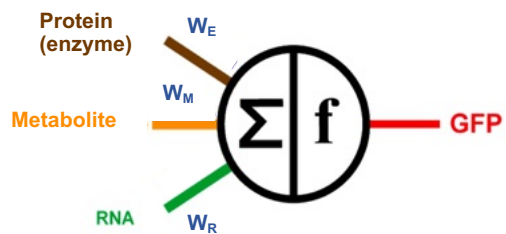
ACTIVE LEARNING APPROACH



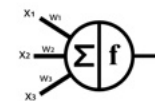
- Borkowski O. *et al. Nature Comm.* 2020
- & Pandi A. *et al. Nature Comm.* 2022



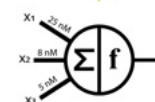
Engineering a multimodal perceptron



ACTIVE LEARNING APPROACH

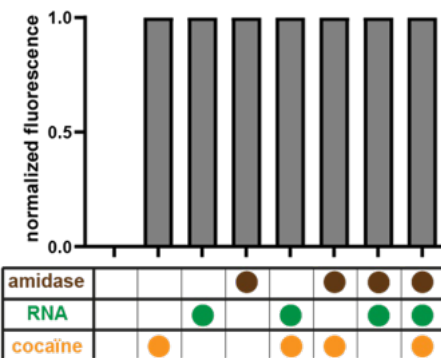


active learning loop

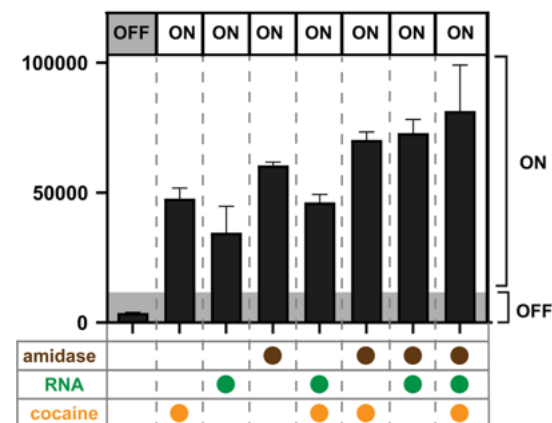


Targeted behavior

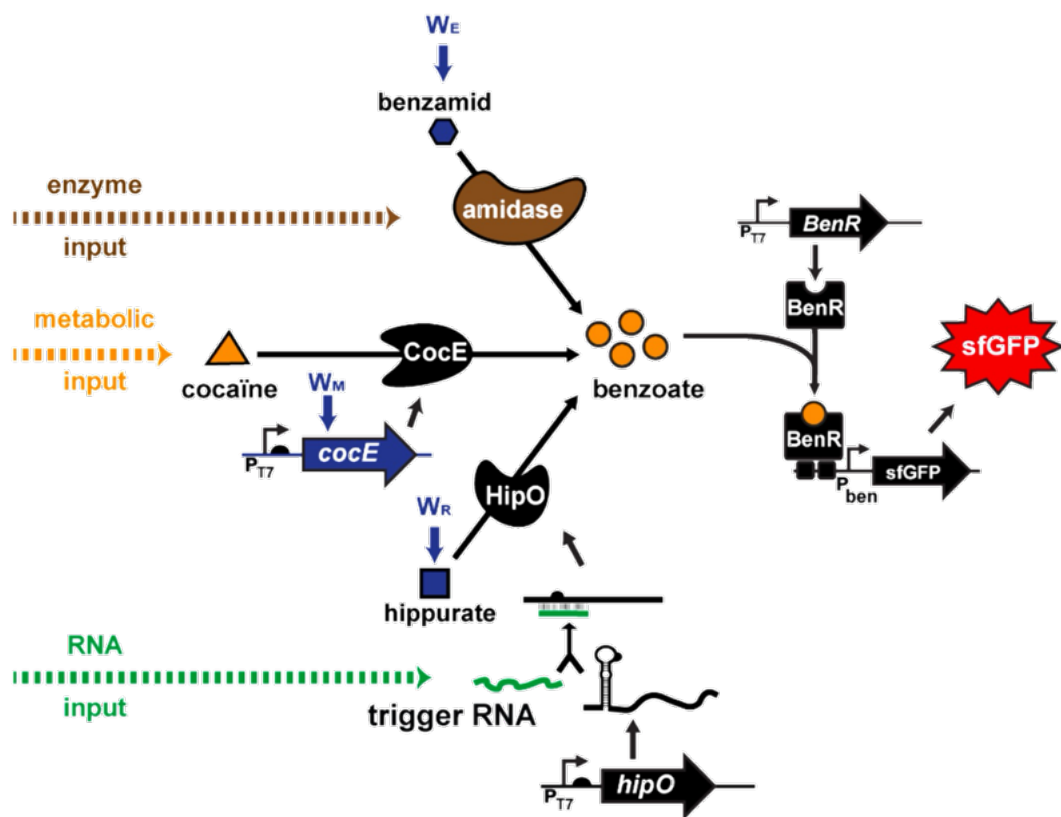
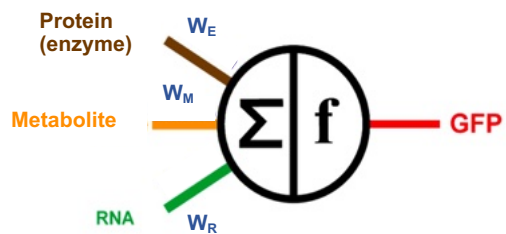
Observed behavior



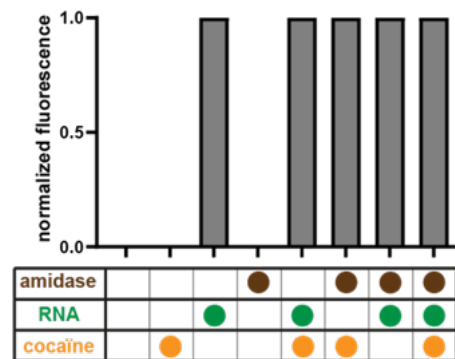
enzyme OR RNA OR metabolite Gate



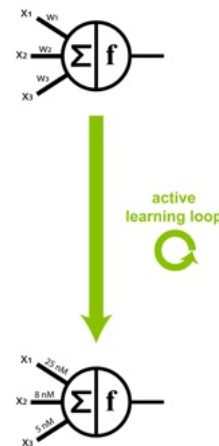
Engineering a multimodal perceptron



Targeted behavior

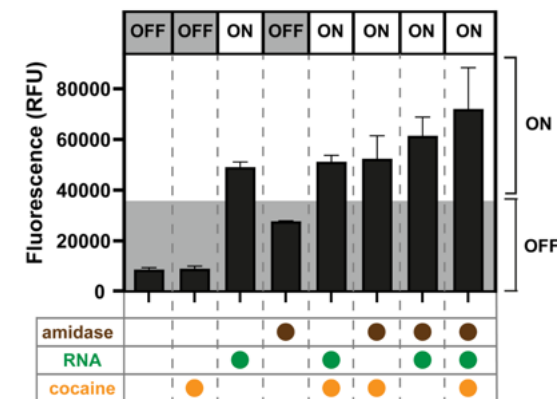


ACTIVE LEARNING APPROACH

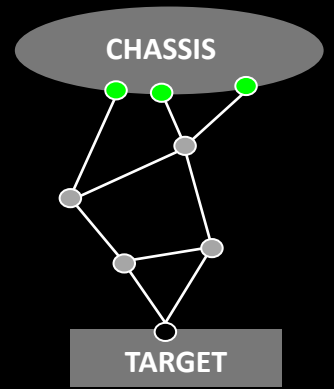


Observed behavior

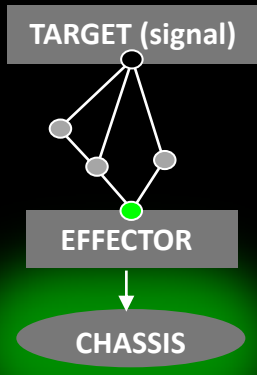
(metabolite AND enzyme) OR RNA Gate



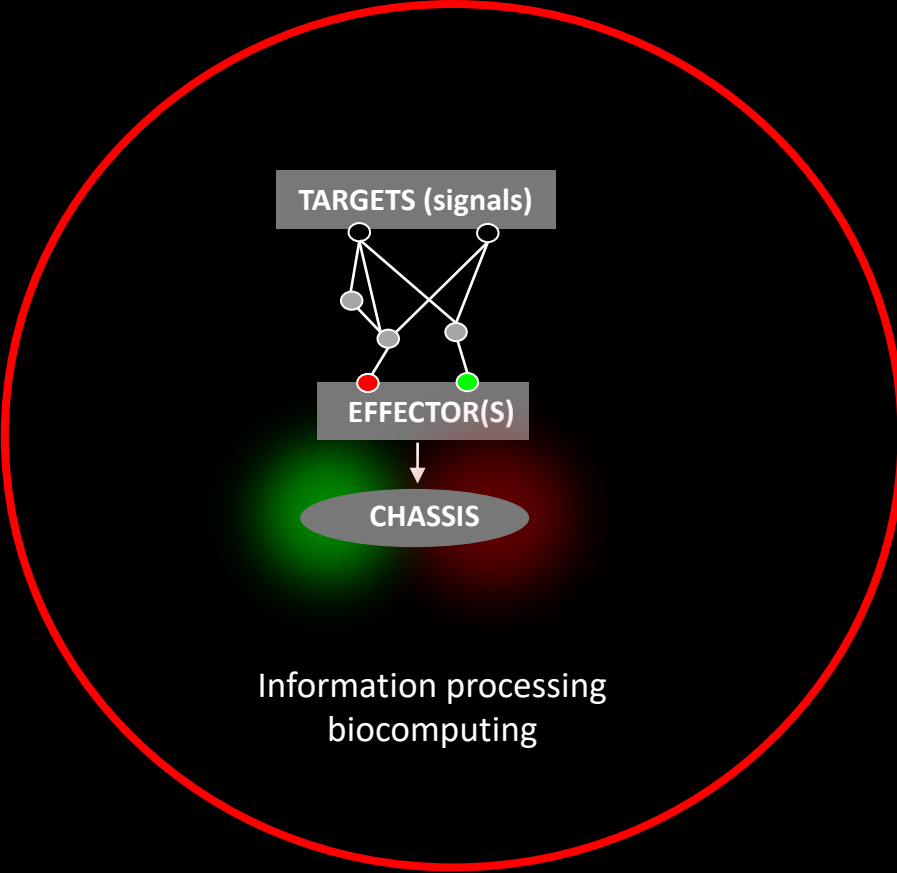
Engineering metabolic and genetic networks in vitro (cell-free) and in vivo (E. coli)



Metabolic engineering



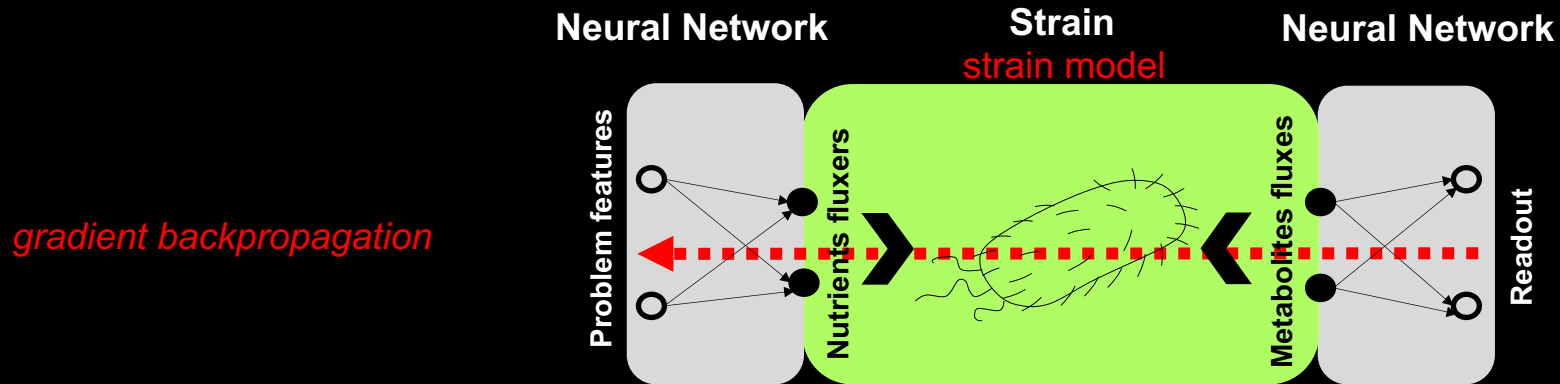
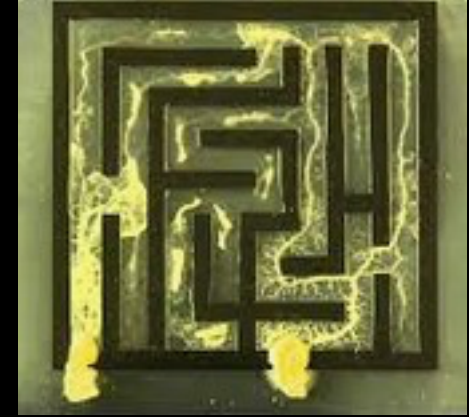
Biosensor engineering



Information processing
biocomputing

Engineering complex metabolic devices in vivo?

- Can we divert native metabolism to handle problems that are usually solved *in silico*?



A model allowing gradient backpropagation accurately reproducing phenotype for different media composition

Engineering complex metabolic devices in vivo?

Classical mechanistic model (FBA):

$$\text{Max } (v_{biomass})$$

Subjected to:

$$S V = 0$$

$$0 \leq V \leq V_{in}$$

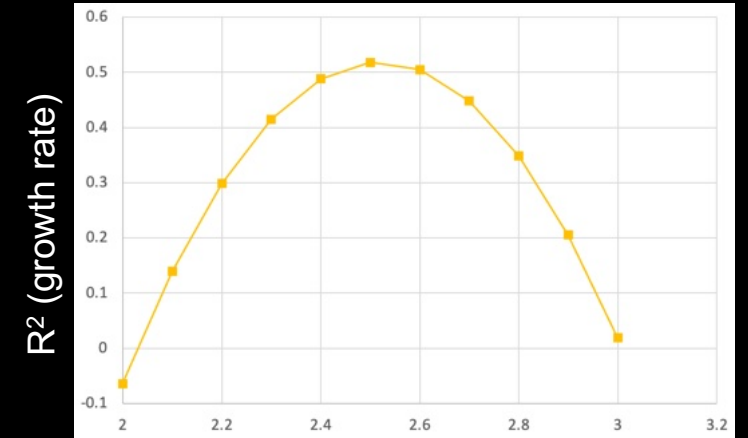
where

– V = set of all reaction fluxes

– S = stoichiometric matrix

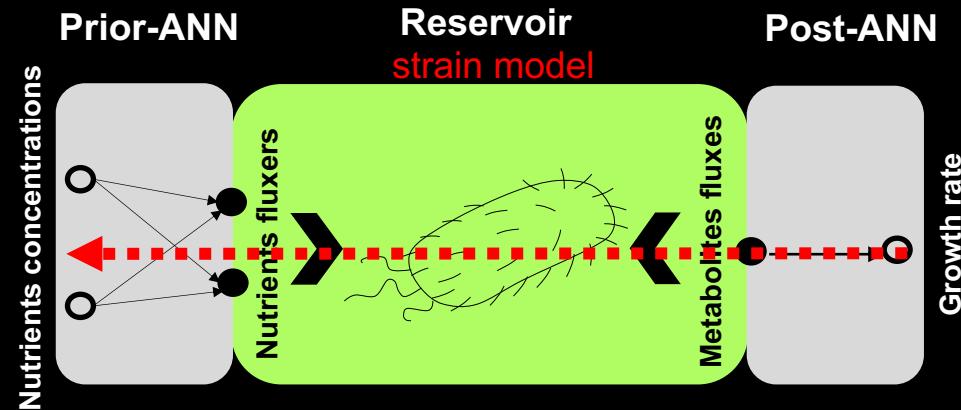
– V_{in} = uptake medium fluxes upper bounds

FBA (Cobrapy) growth rates vs. measured growth rate in E. coli DH5-alpha for 1 to 4 nutrients added to M9



Concentration to flux scaler

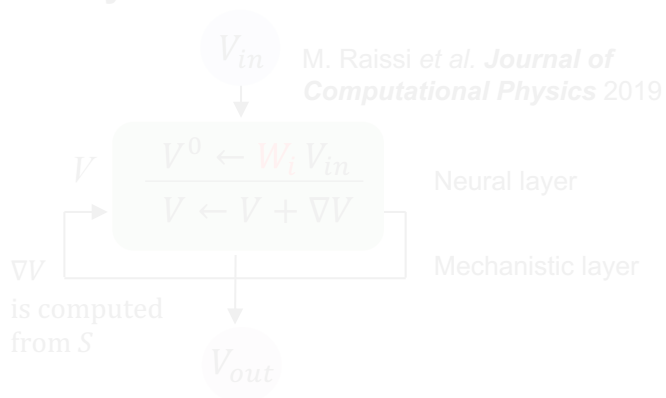
gradient backpropagation



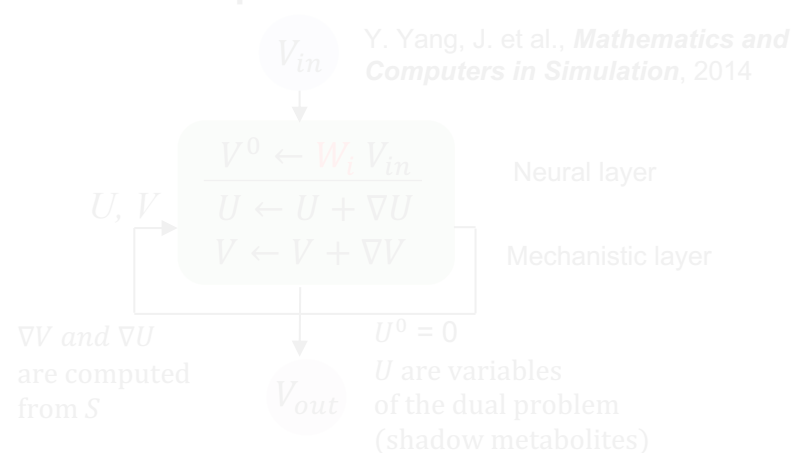
a model allowing gradient backpropagation that accurately reproduce phenotype for different media composition

Hybrid models: gradient backpropagation compatible solutions surrogating classical mechanistic models

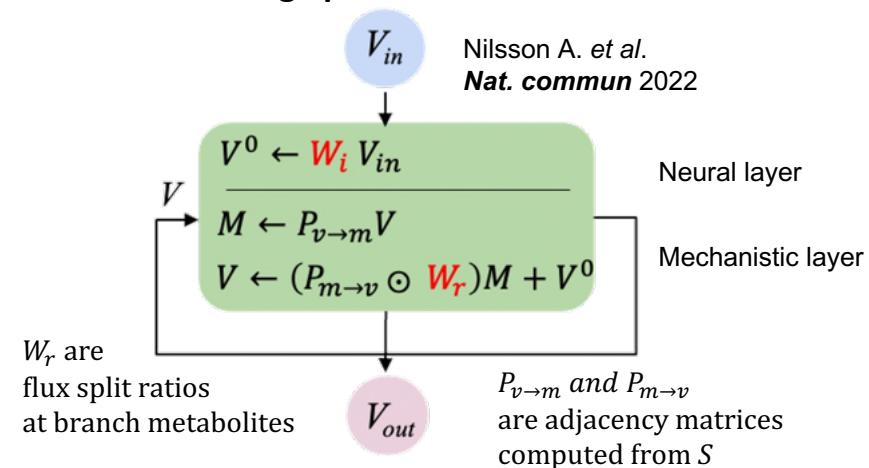
Physics informed neural network



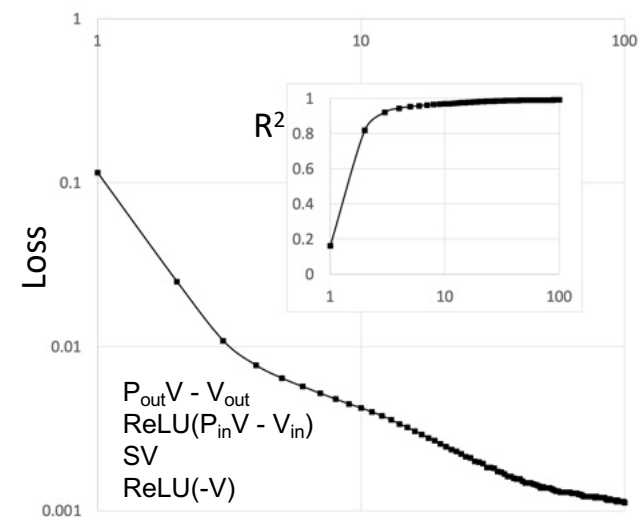
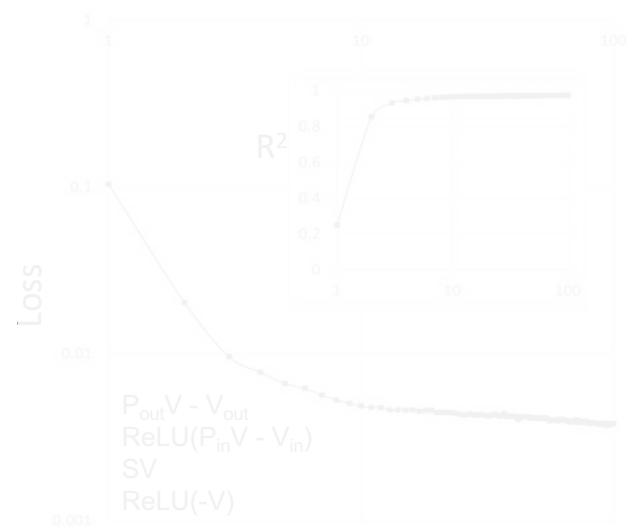
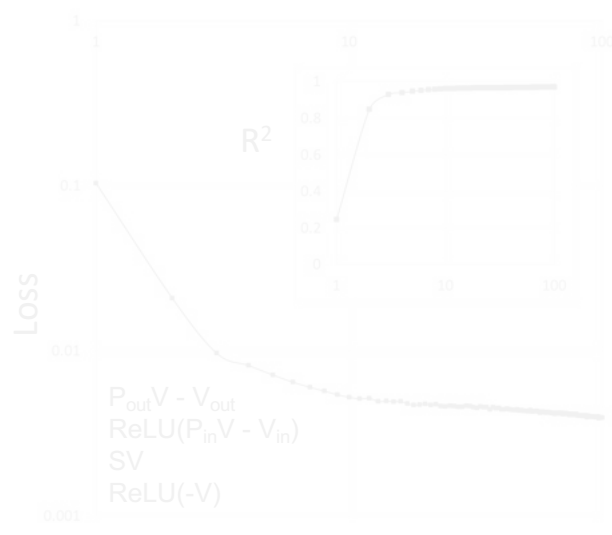
Hopfield's network



Knowledge primed neural network

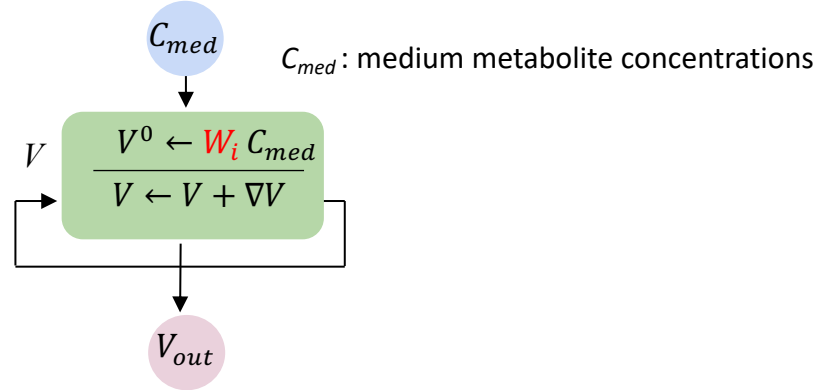


Trained on FBA simulated growth rates with *E. coli core* model for 1000 different media (media = minimal medium + 1 to 6 metabolites chosen at random among 13)

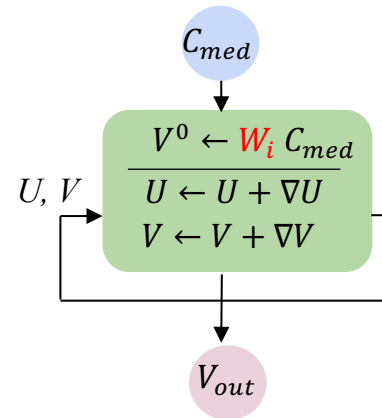


Hybrid models: gradient backpropagation compatible solutions surrogating classical mechanistic models

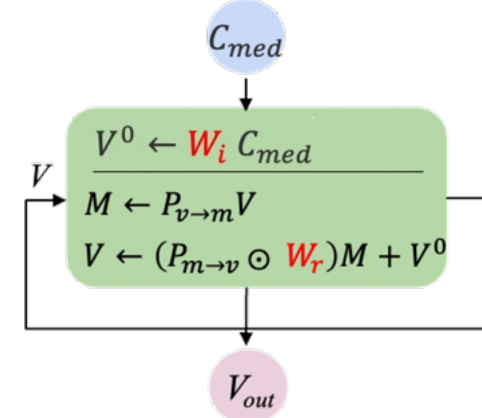
Physics informed neural network



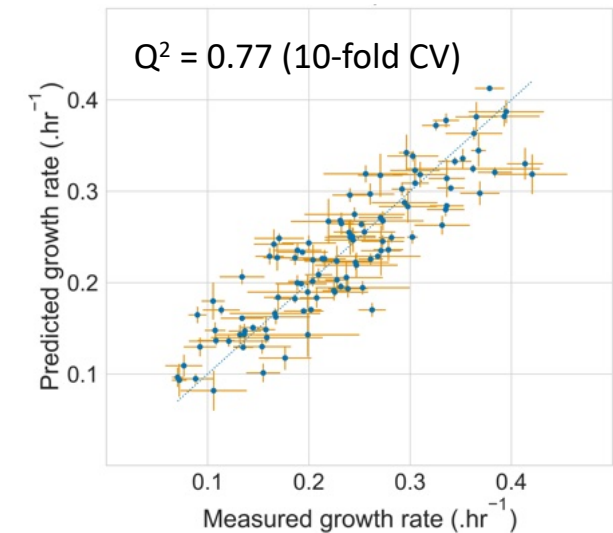
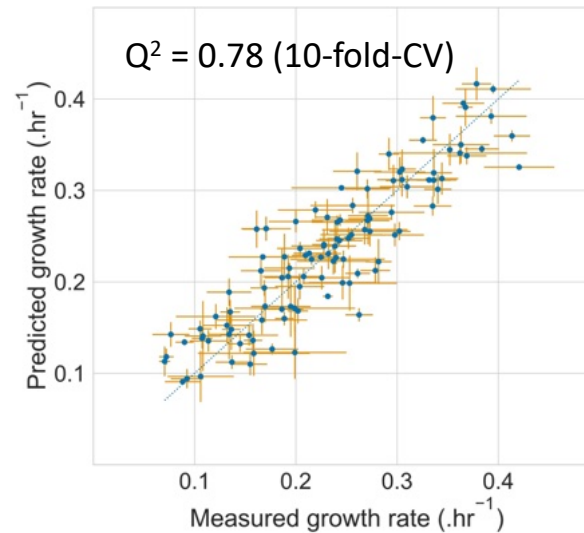
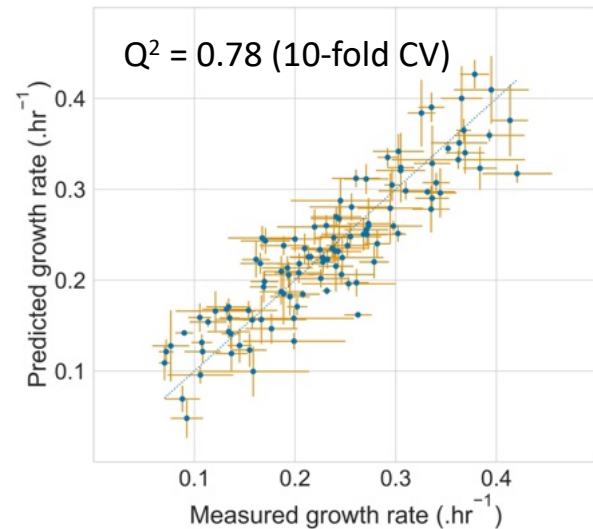
Hopfield's network



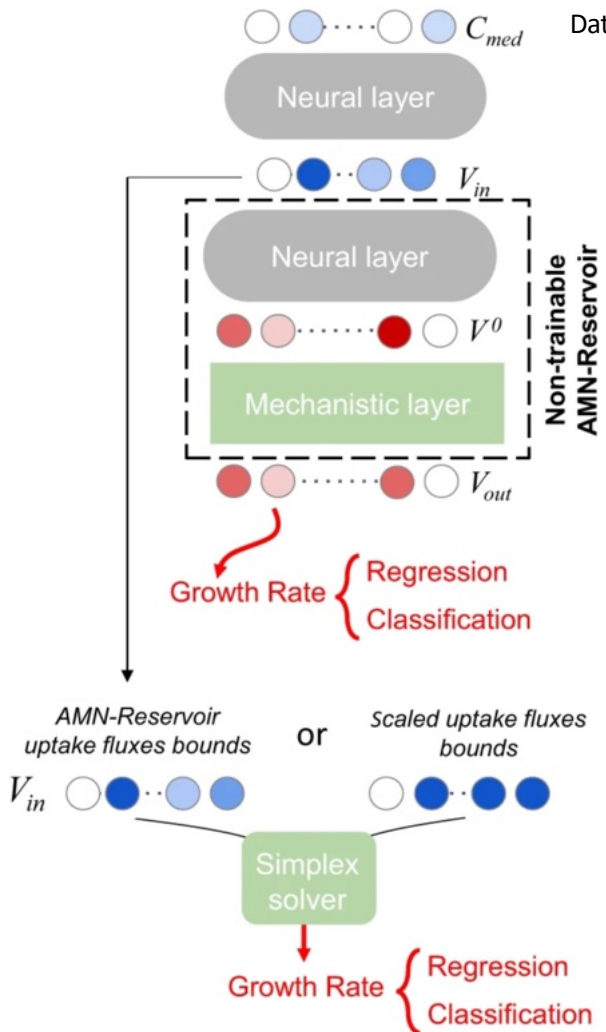
Knowledge primed neural network



Prediction after being trained on an experimental data set where growth rates were measured for 110 different media compositions for *E. coli* DH5-alpha strain (media = M9 + 4 nutrients chosen at random among 10)

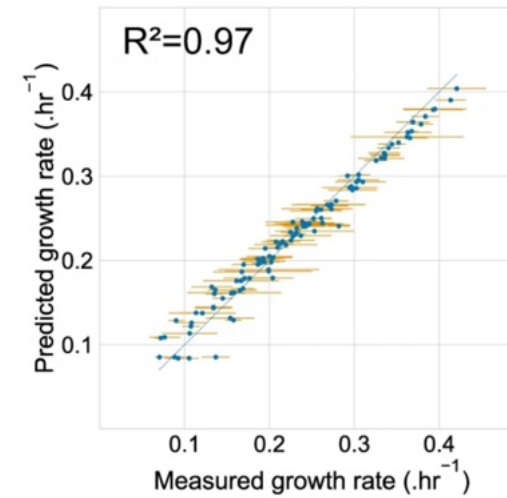


Hybrid models: can be used to parameterize mechanistic models

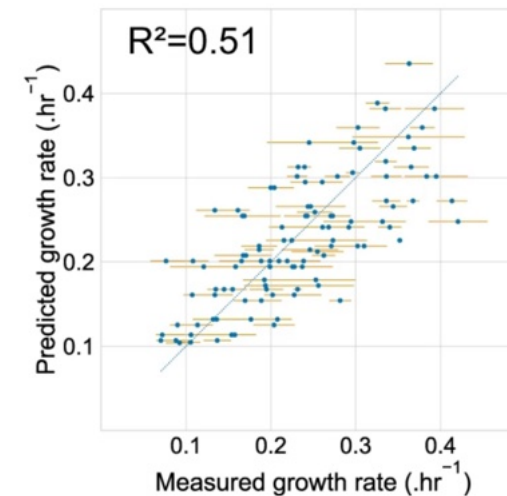


Data set 110 different media compositions for *E. coli* DH5-alpha for which growth rates have been measured

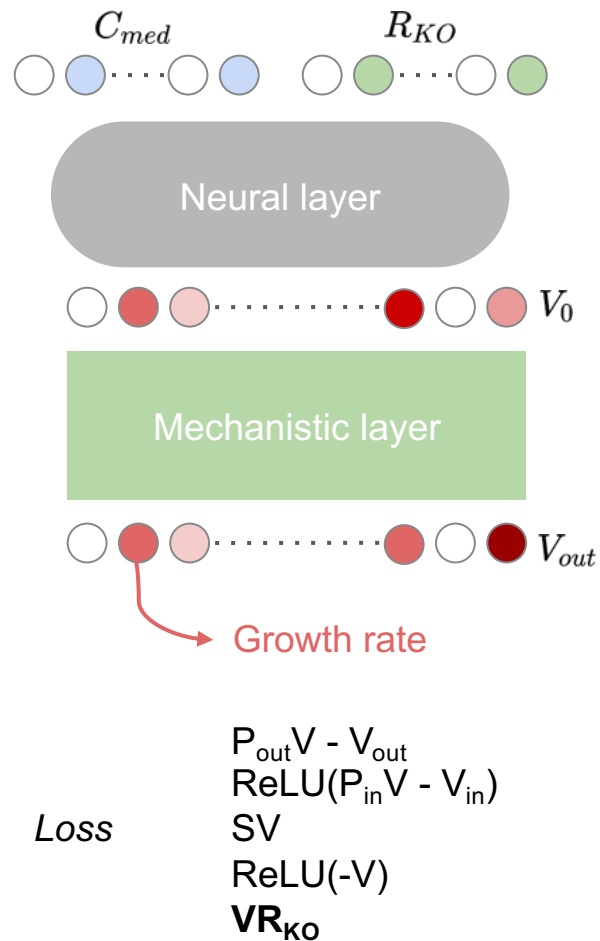
Hybrid model results



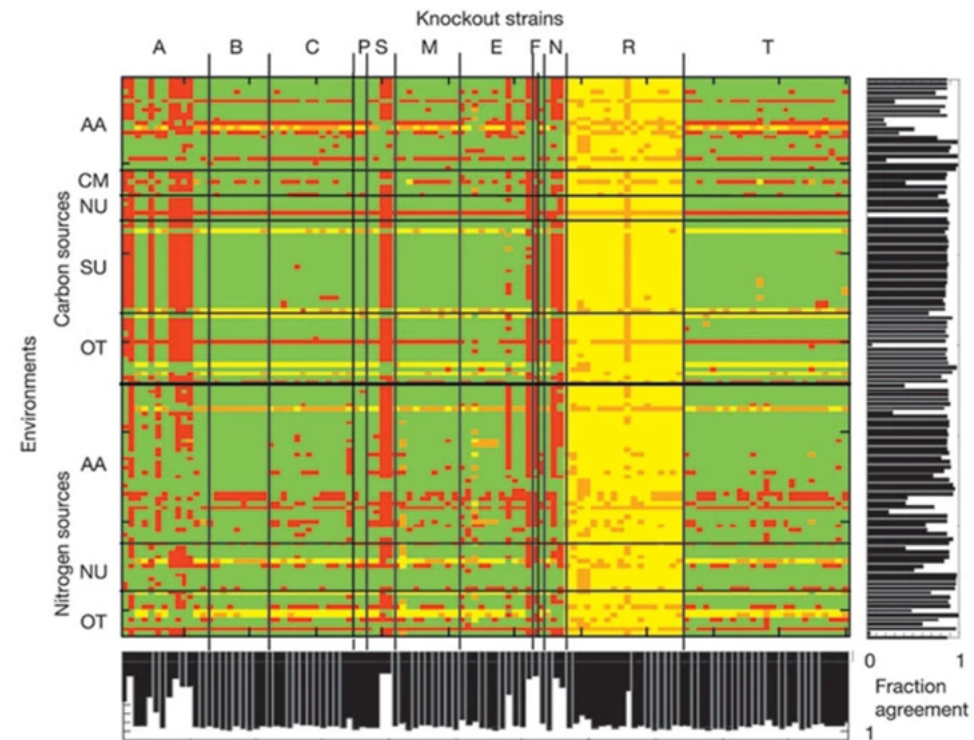
Mechanistic model results



Hybrid models: can integrate gene regulation

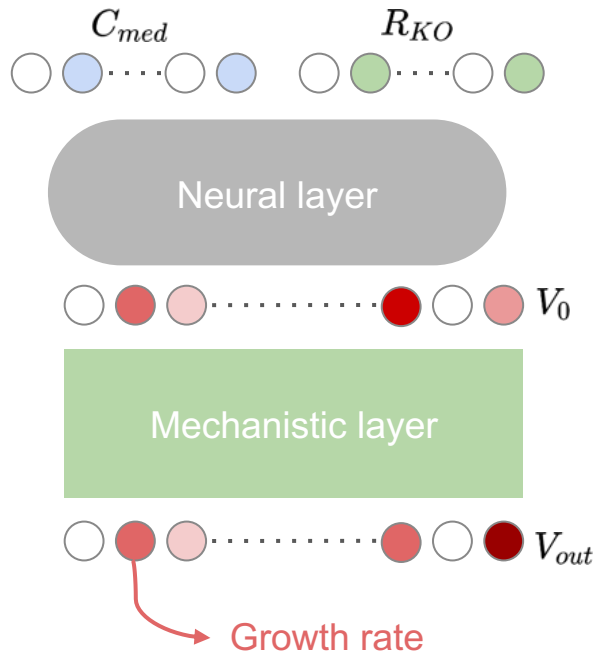


Dataset from ASAP database (Glasner D. *et al.* NAR 2003)



- 120 metabolic genes targeted (KOs), targeting 127 reactions
- 145 conditions (1 or 2 substrates, some with added succinate to enable growth)
- Each KO screened in all conditions (145*120=17400)

Hybrid models: can integrate gene regulation



Loss

$$P_{out}V - V_{out}$$

$$\text{ReLU}(P_{in}V - V_{in})$$

$$SV$$

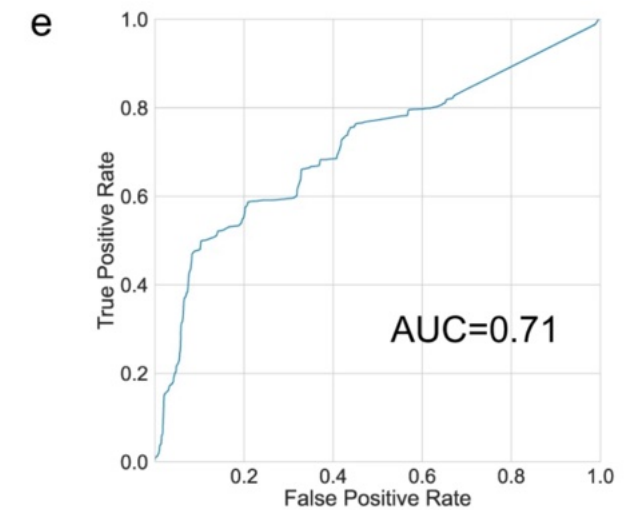
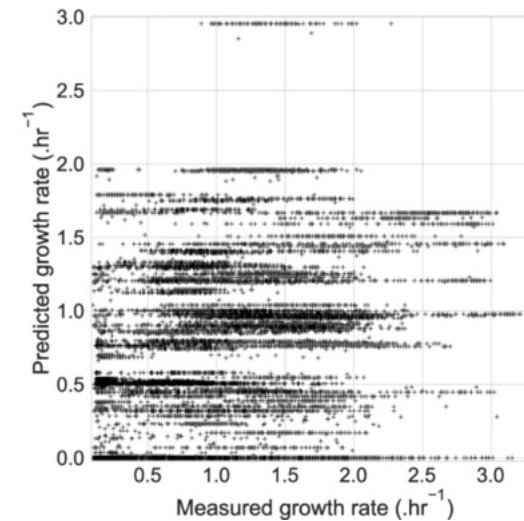
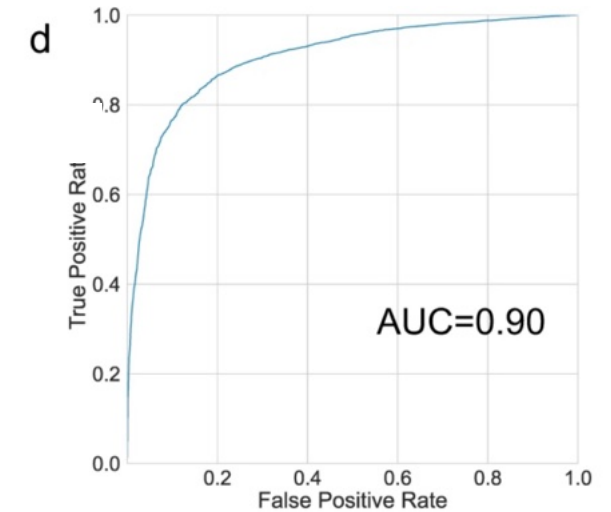
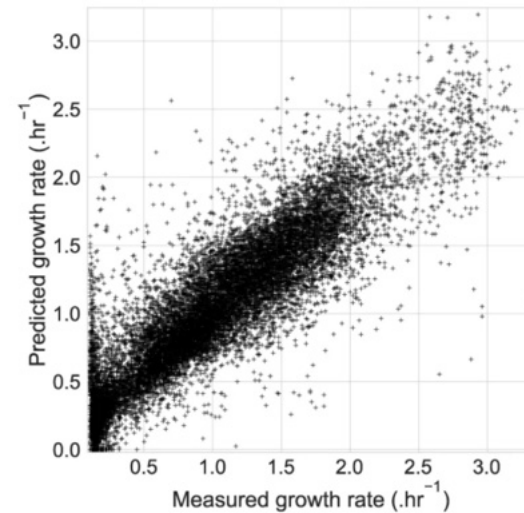
$$\text{ReLU}(-V)$$

$$VR_{KO}$$

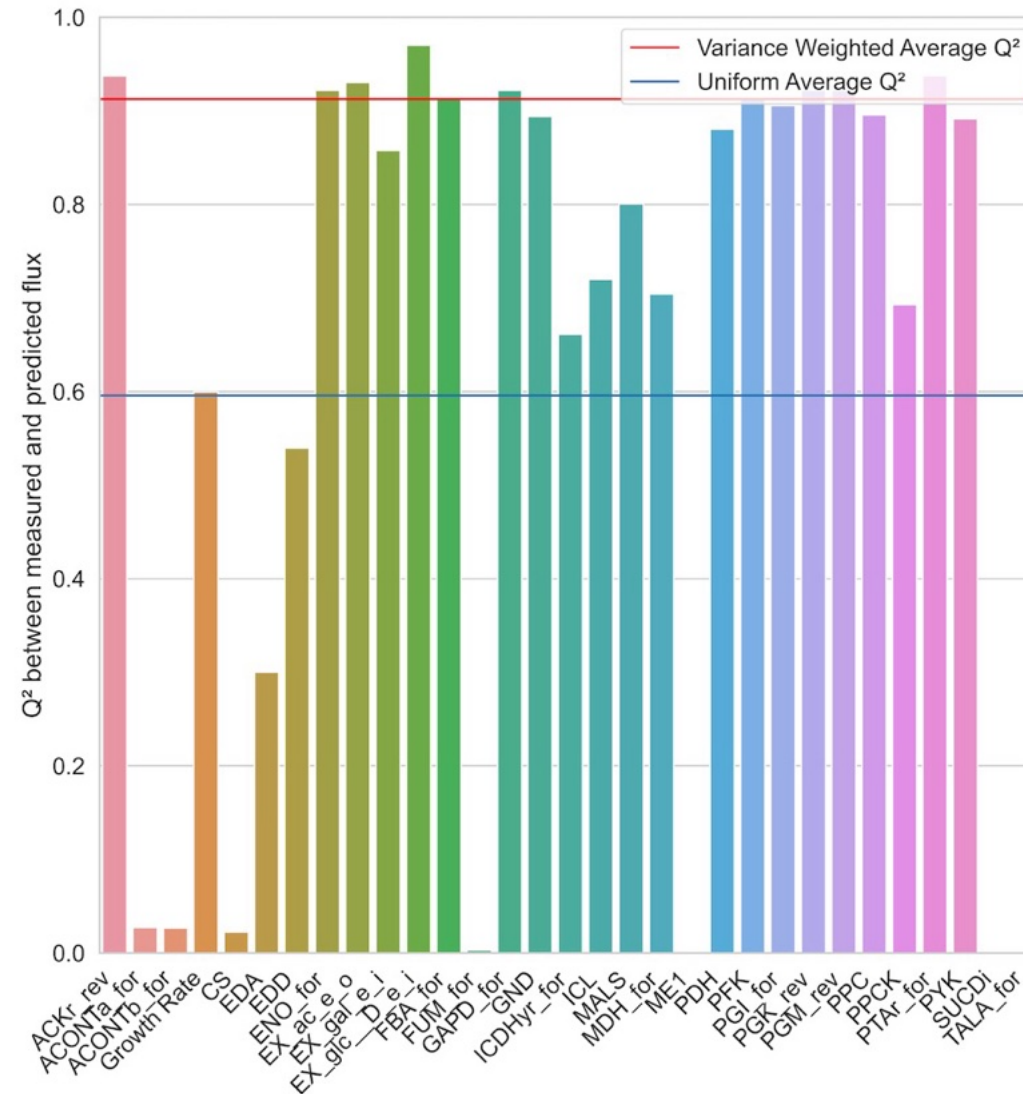
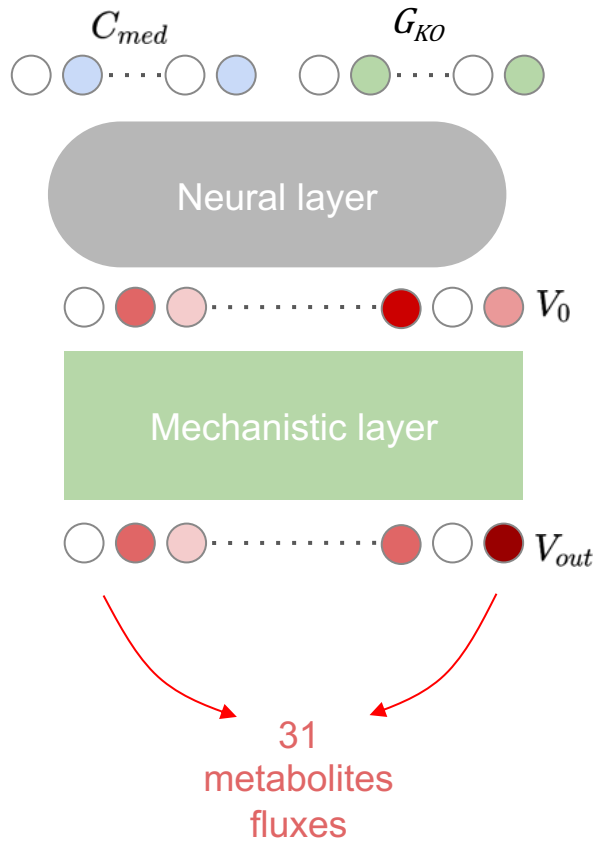
Hybrid model results

Mechanistic model results

Dataset from ASAP database (Glasner D. *et al.* NAR 2003)



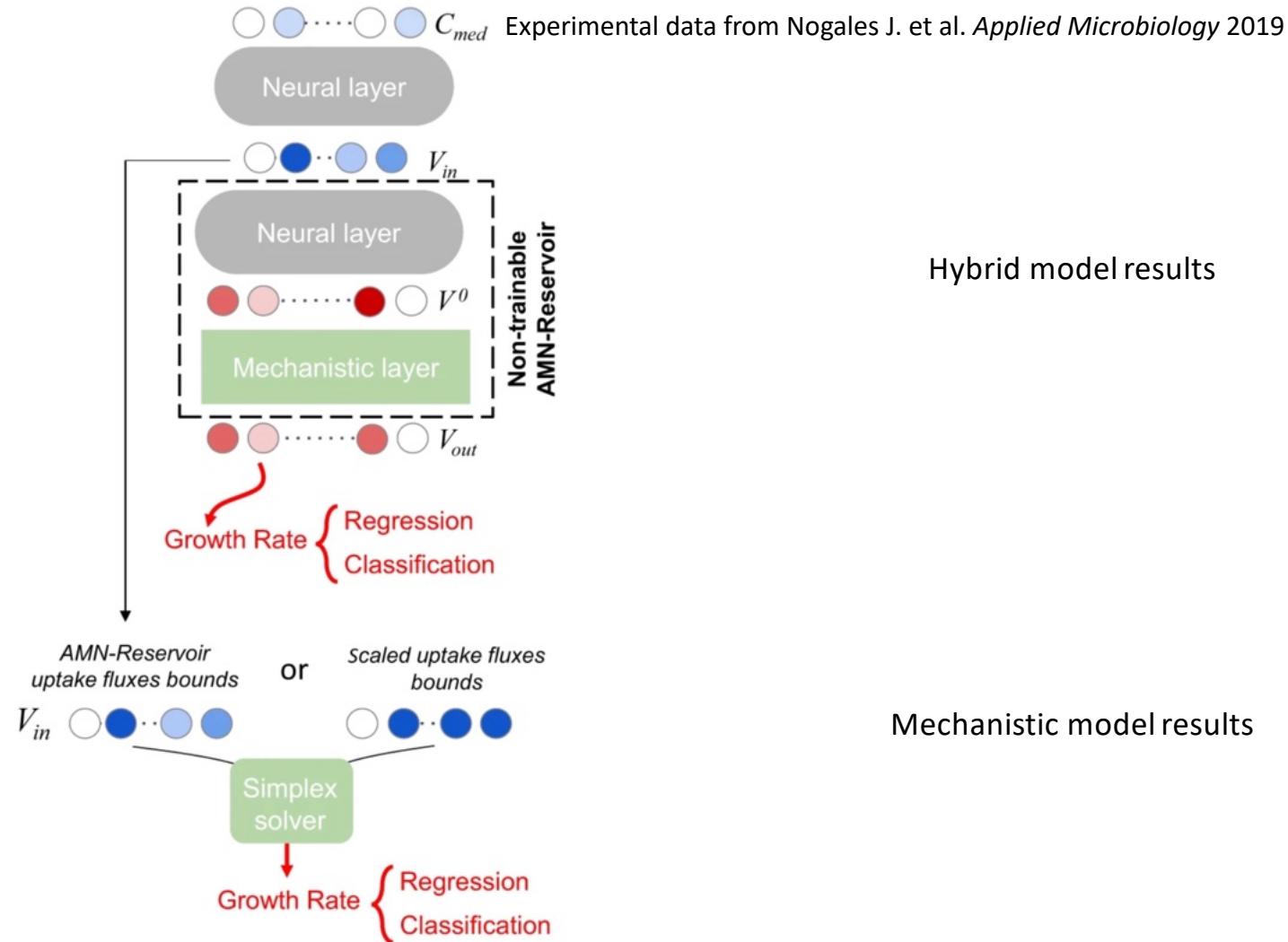
Hybrid models: can learn more than the growth rate



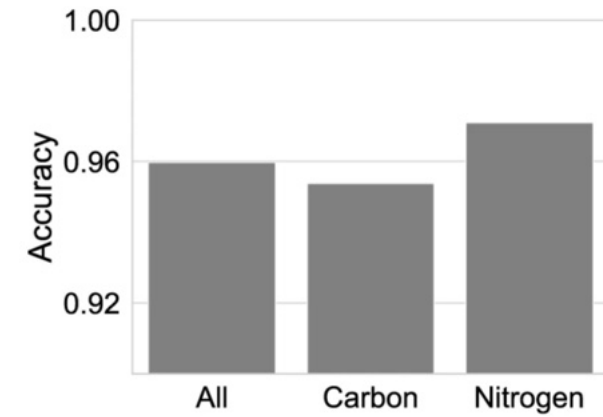
E. coli dataset from Rijsewijk *et al. Mol. Syst. Biol.* **7**, 477, 2011

- 128 experiments, each containing 31 measured fluxes.
- 2 media compositions (glucose or galactose as carbon source)
- 64 regulator gene KOs mutants (GKO)

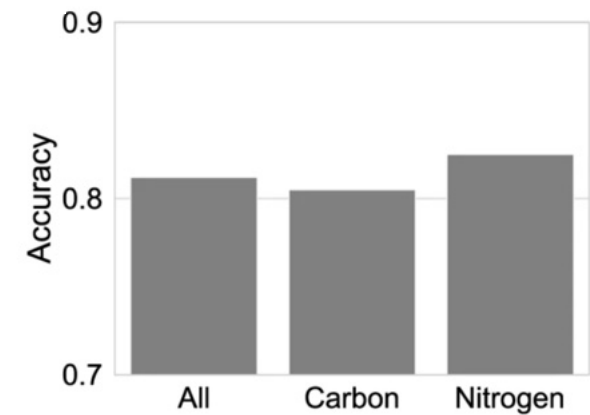
Hybrid models: Application to *P. putida*



Hybrid model results



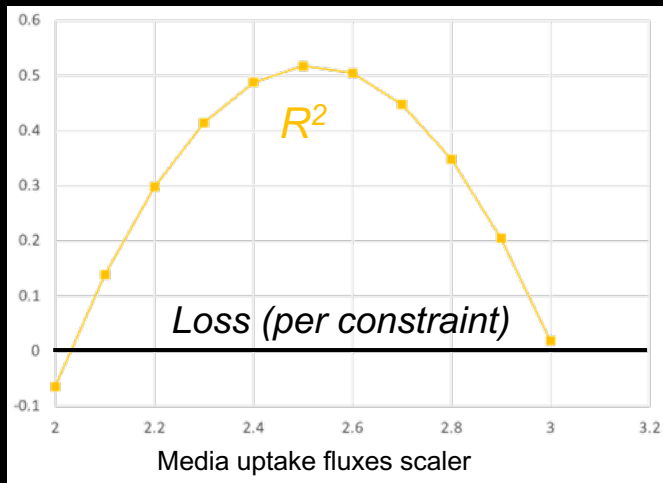
Mechanistic model results



Hybrid model: a new modelling paradigm

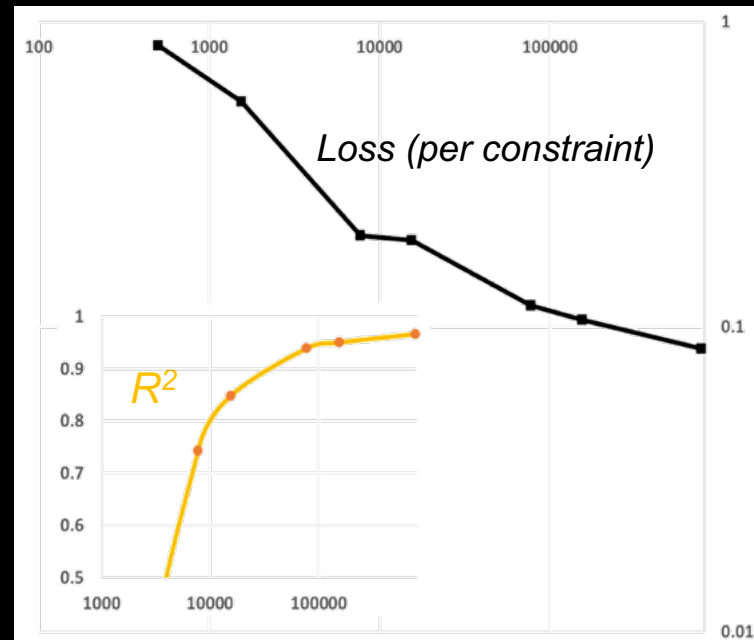
Training set : reaction flux for different media compositions (strain *E coli* MG1655, model = iML1515, media = M9 + 4 nutrients chosen at random among 10, measured reaction flux is chosen at random among all reactions).

Mechanistic model (FBA Cobrapy)
Training set not needed



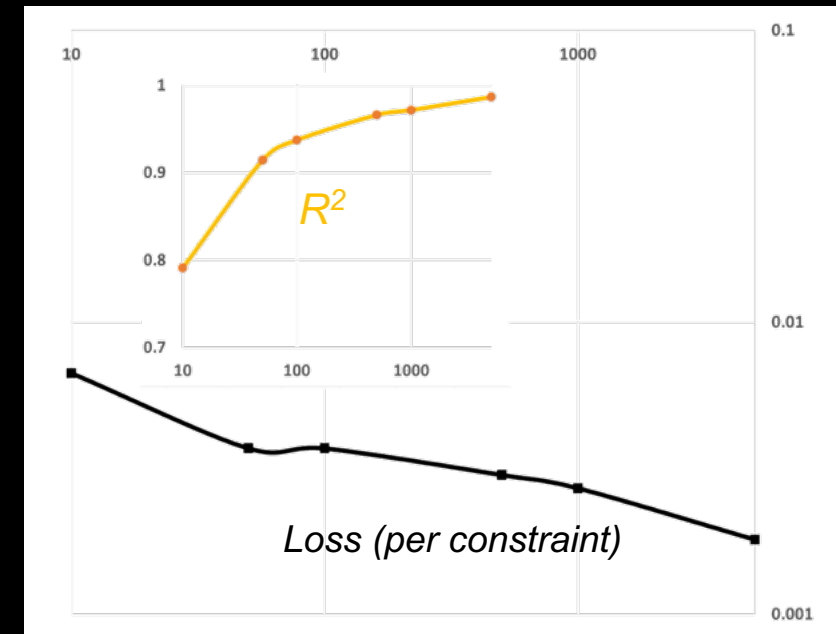
Mechanistic constraints are respected but poor match with measured (training) data

Black –box neural model (dense-ANN)
> 500k entries needed in training

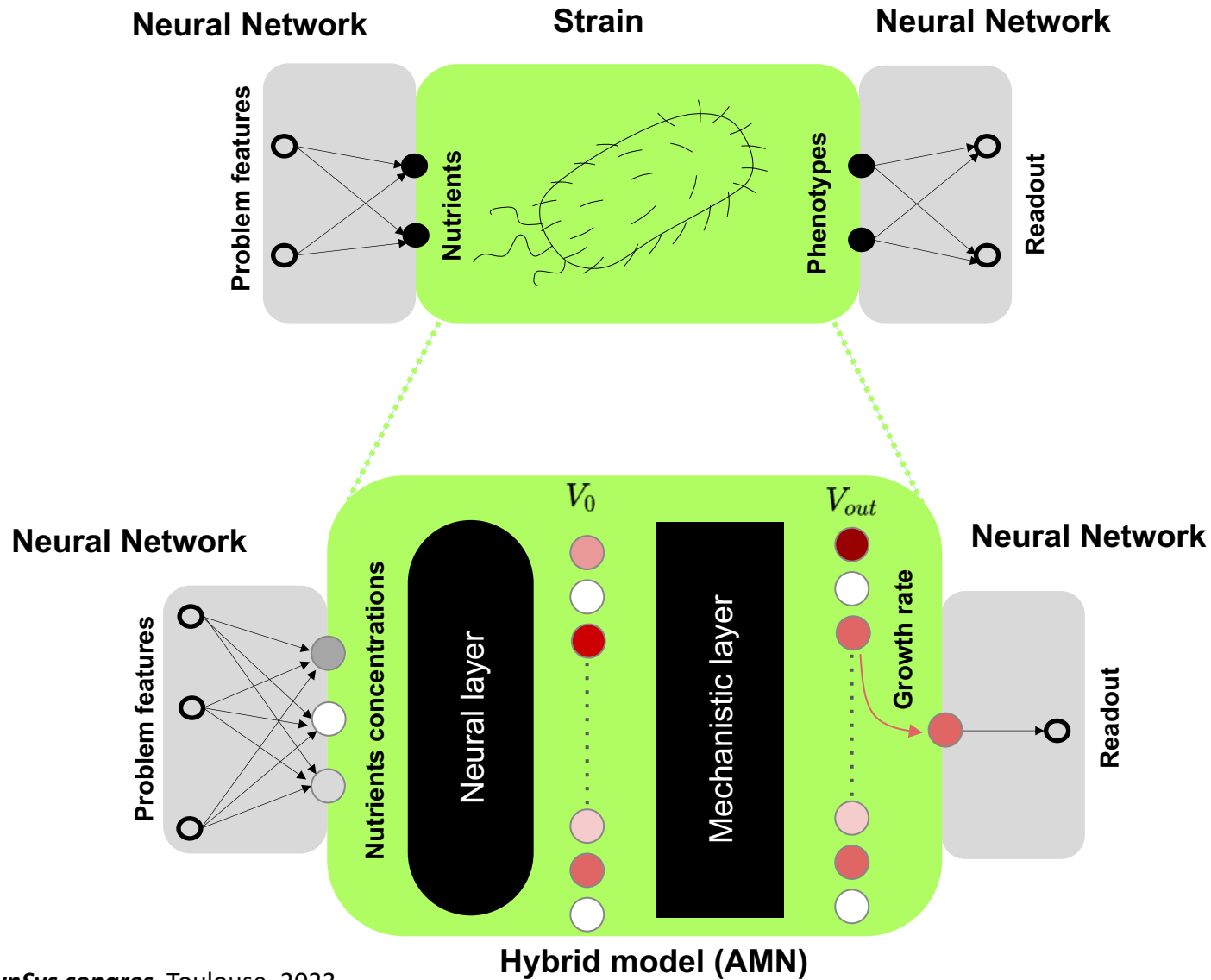


Predictions match measured training data but do not fit mechanistic constraints

Hybrid-model
Less than 1000 entries in training are enough

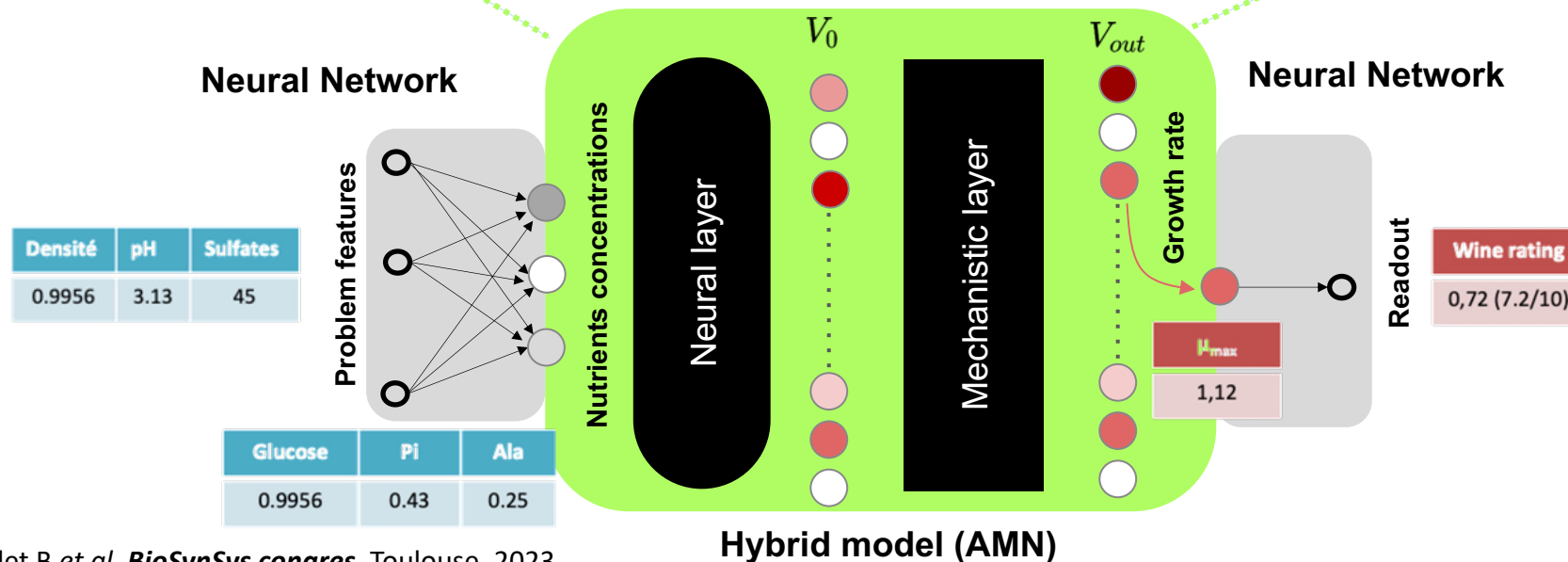
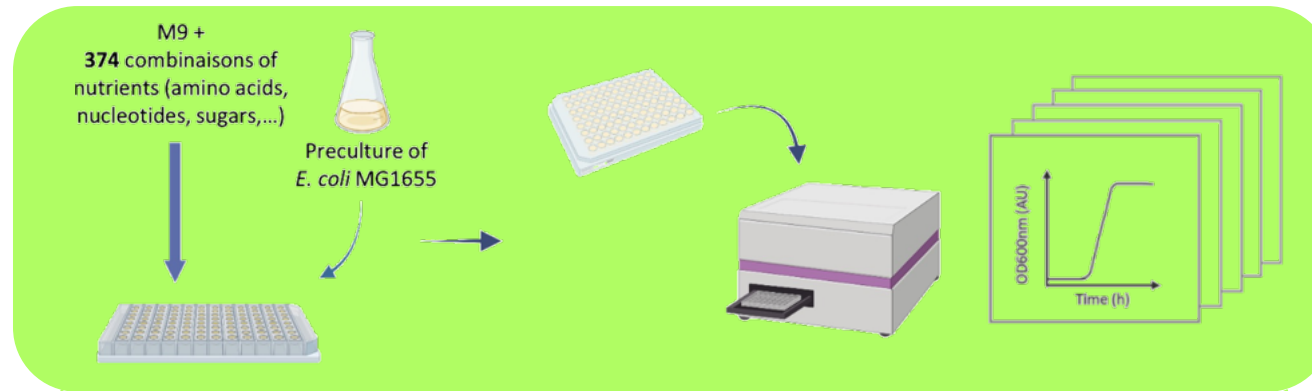


Can microorganism metabolism be diverted to solve classical machine learning problems?

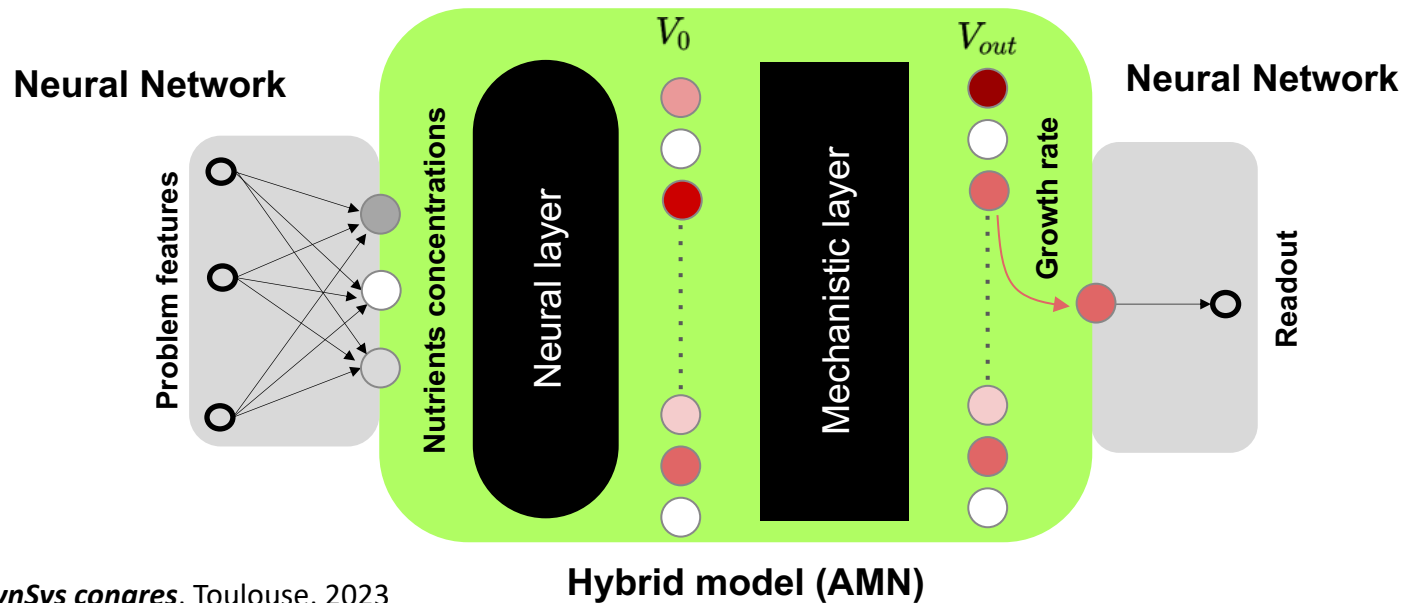
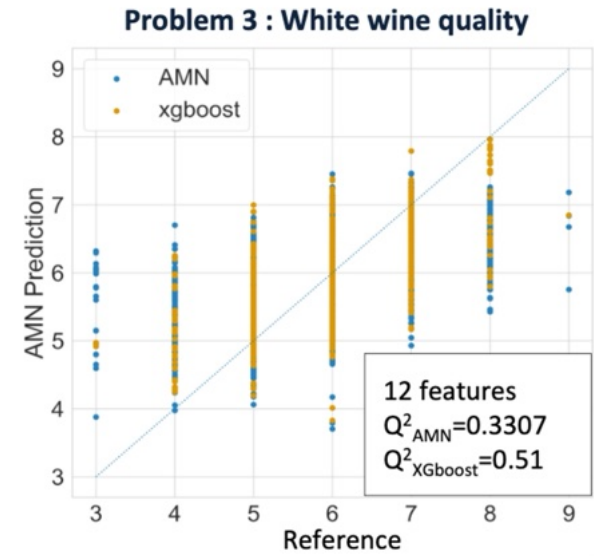
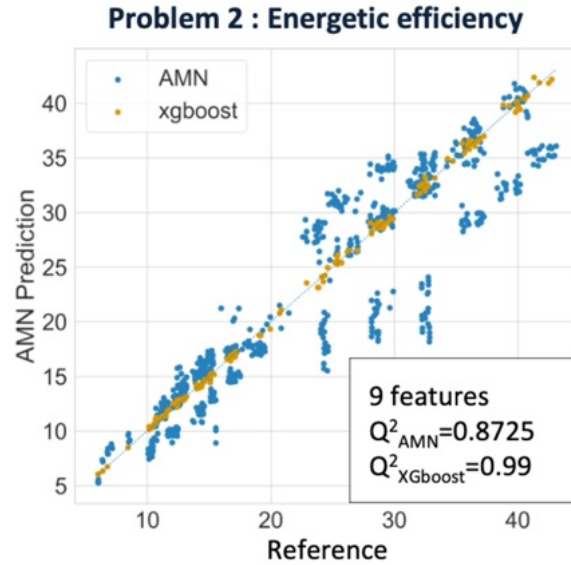
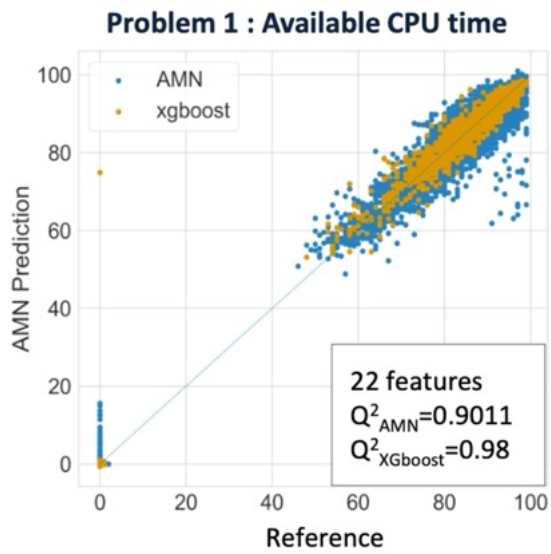


Can microorganism metabolism be diverted to solve classical machine learning problems?

Example : White wine quality dataset (score from 1 to 10)



Can microorganisms solve classical machine learning problems?



More pragmatic (and funded) applications for hybrid modelling

Bioproduction

- Find the best gene deletion in *P. putida* to produce terpenes, biodegradable polyesters (polylactic acid, polyethylene furanoate), and methylacrylate (a building block for plexiglass)



Biodegradation for biotherapy

- Find the best gene deletion in *E. coli* Nissle 1917 engineered strain to degrade p-cresol and 4-ethylphenol (two metabolites involved in Autism)



Diagnostic

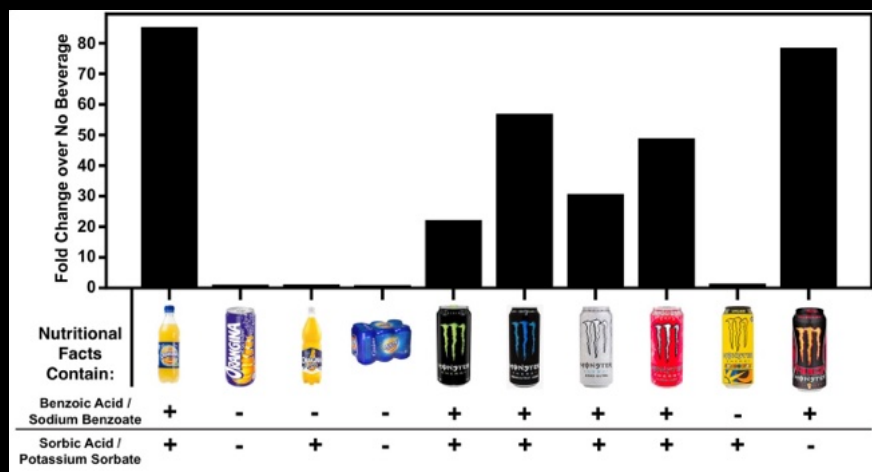
- Find the best gene deletion in *E. coli* DH5 alpha to classify (benign from severe) Covid-19 samples
- Find the best gene deletion in *E. coli* pilot strain (an engineered strain with CTCs receptors breast cancer) which upon binding produces violacein derivatives (molecules detectable by SERS)





★ Molecular Biology ★ Computational Biology

Benzoic acid (E210) is used as a food preservative
 Biosensor detection in commercial beverages



*We have internships and PhD
 scholarships to propose*

Jean-Loup.Faulon@inrae.fr

<https://jfaulon.com>