

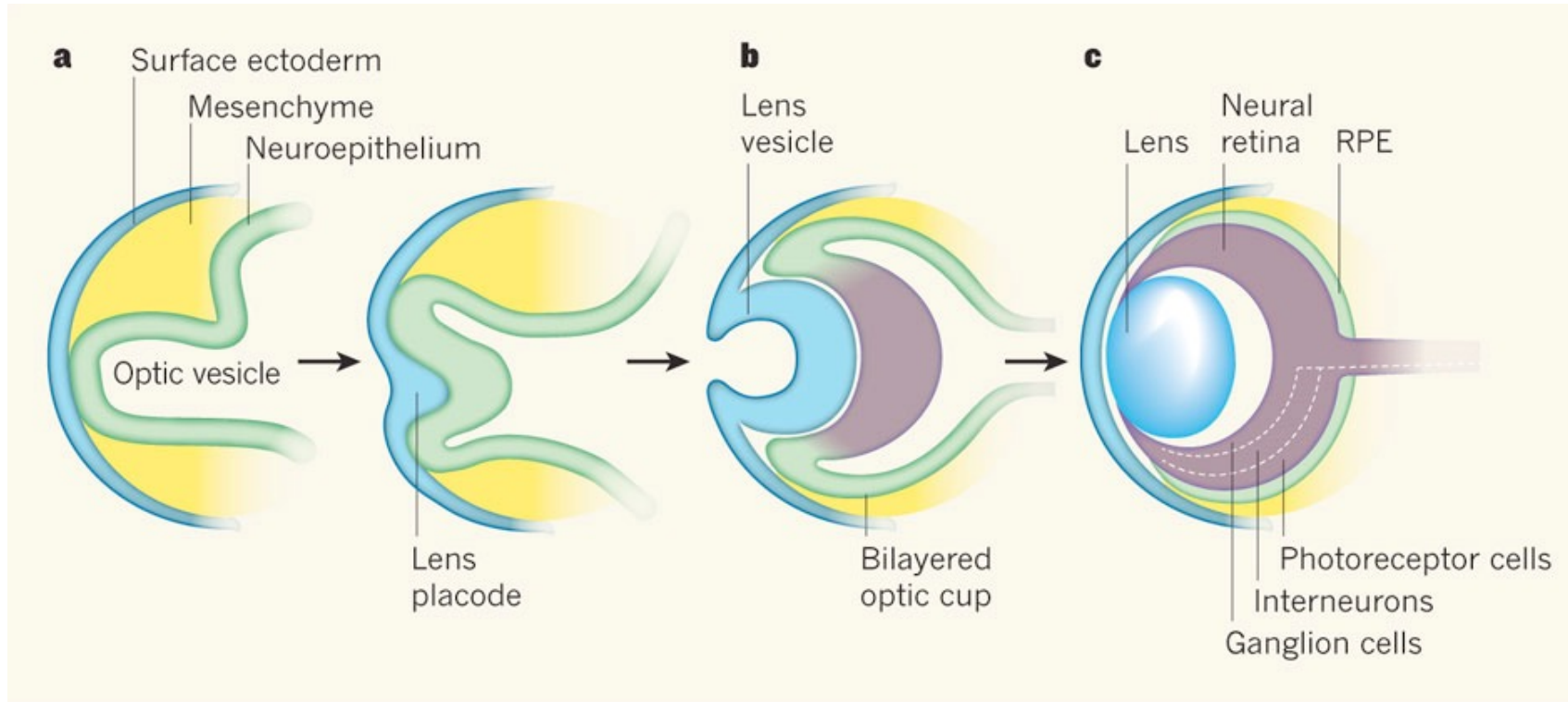
Neural stem cells & Retinal regeneration

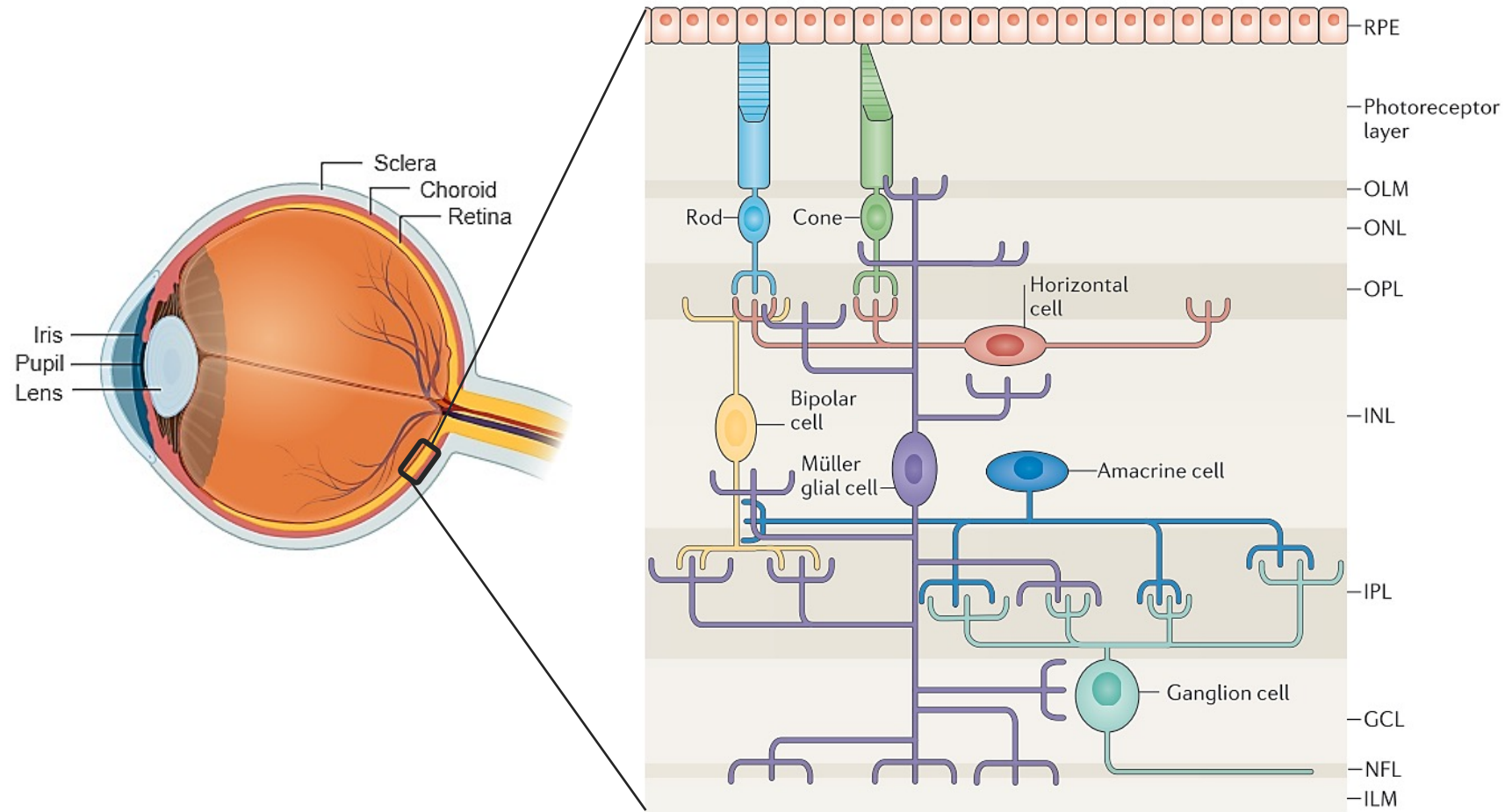


Muriel Perron
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Institut des Neurosciences Paris-Saclay
NeuroPSI
CNRS - Université Paris-Saclay

Retinal development in vertebrates

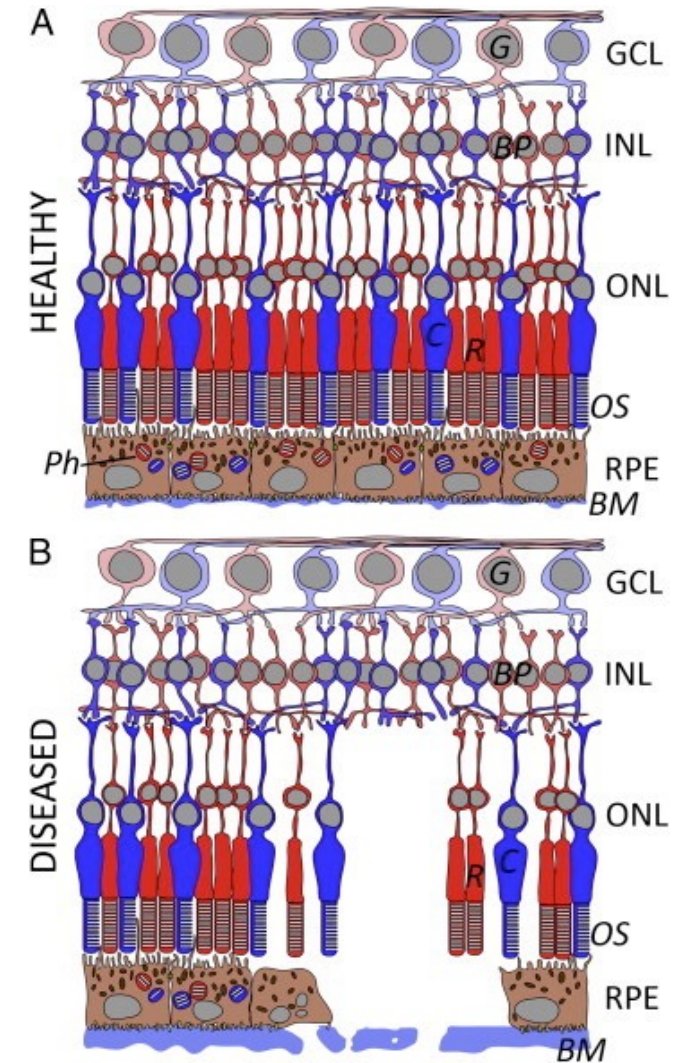


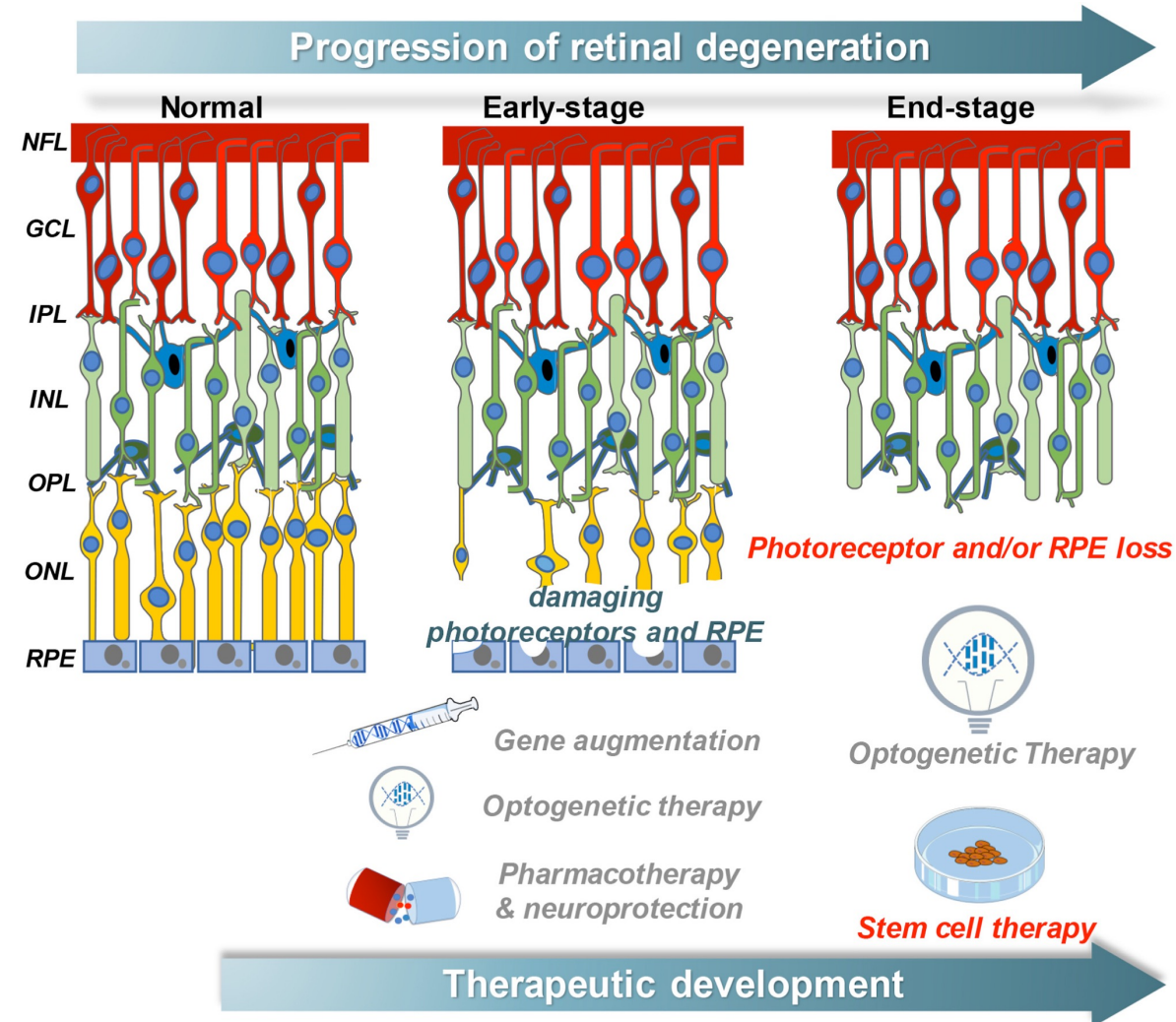


Goldman & al., 2014

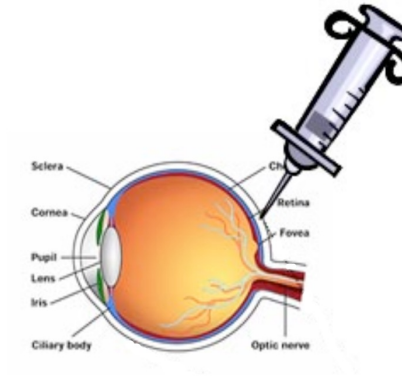
Patients with retinal degenerative diseases in Europe

- AMD: > 10 millions (25% at 75 years-old; 60% at 90 years-old)
- Retinitis pigmentosa: 400 000
- Glaucoma: > 9 millions

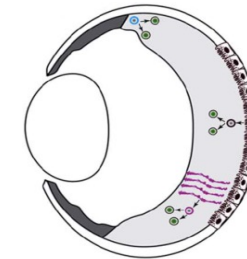




- Transplantation from stem cell-derived retinal cells



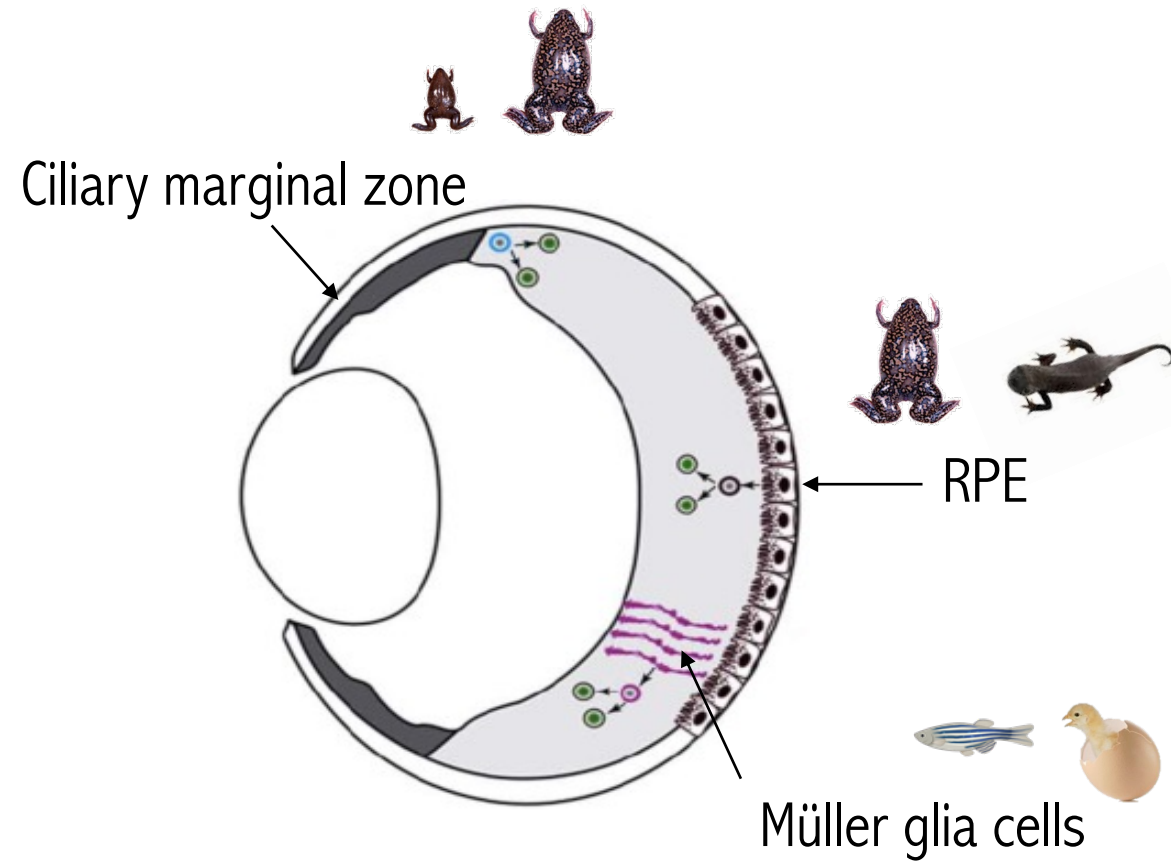
- Regeneration from endogenous stem cells



Cellular sources for retina regeneration



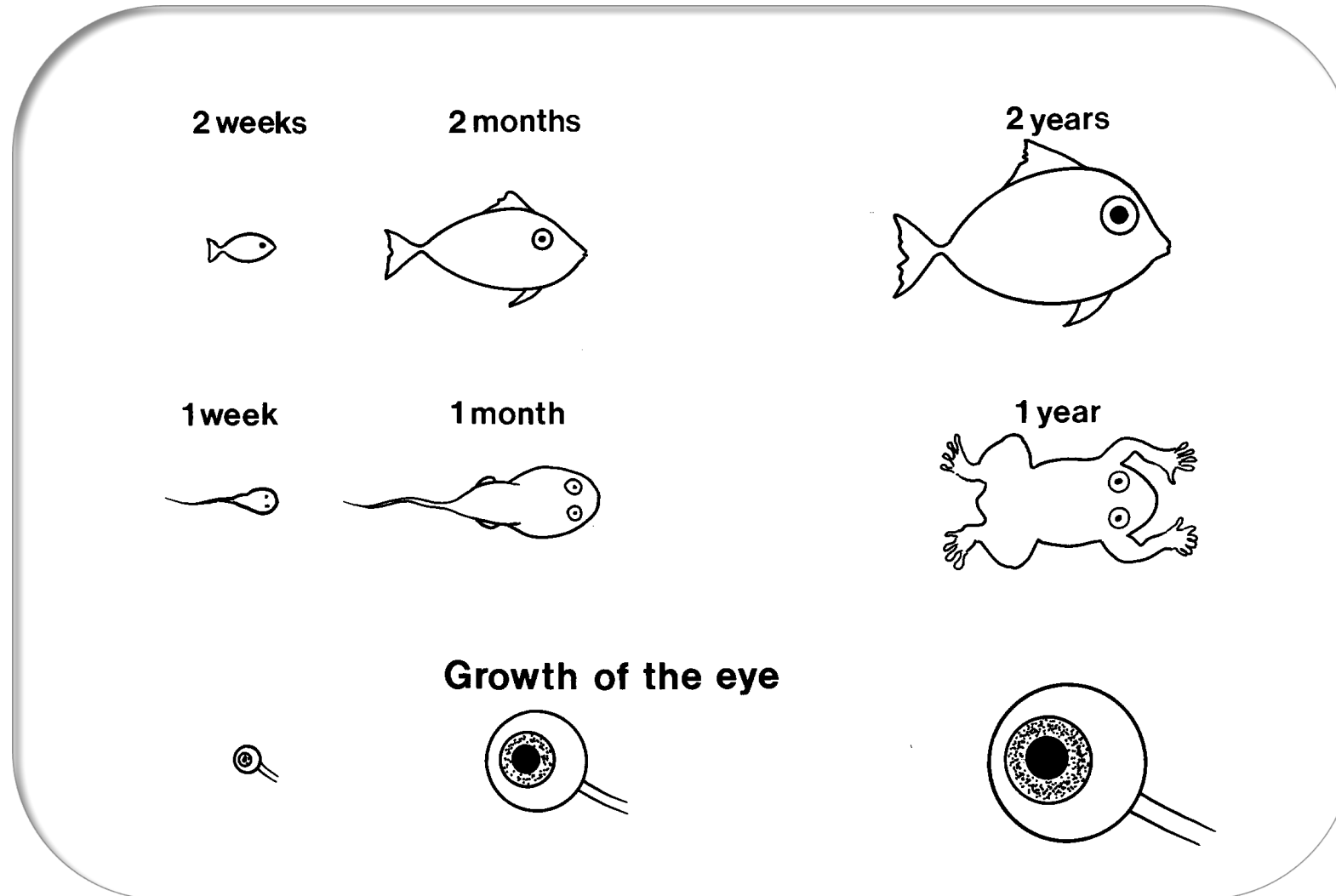
No regeneration



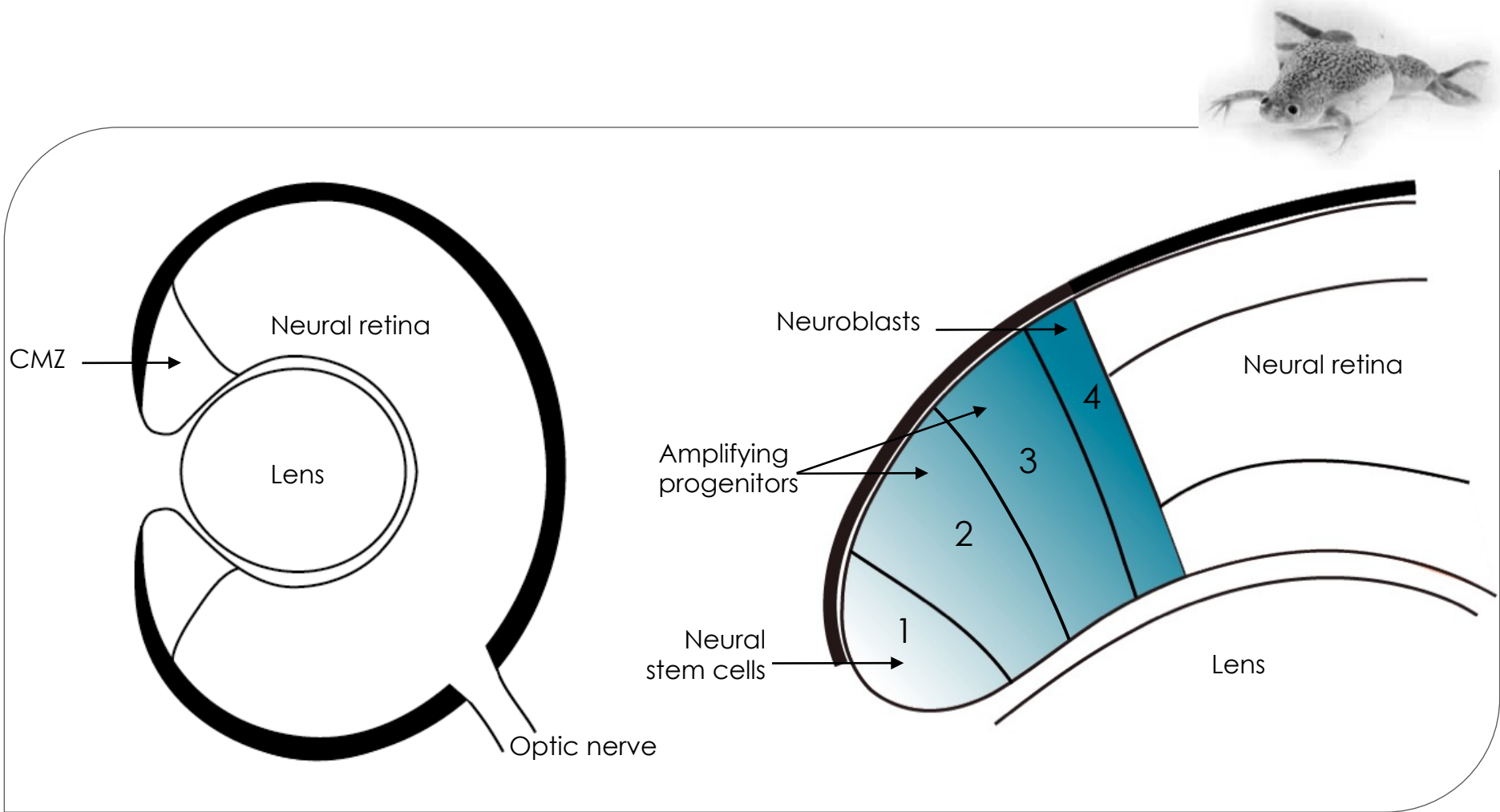
Adapted from Karl & Reh 2010

Ciliary marginal zone stem cells

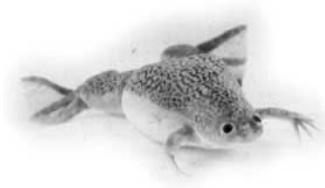
Continuous growth in fish and amphibians



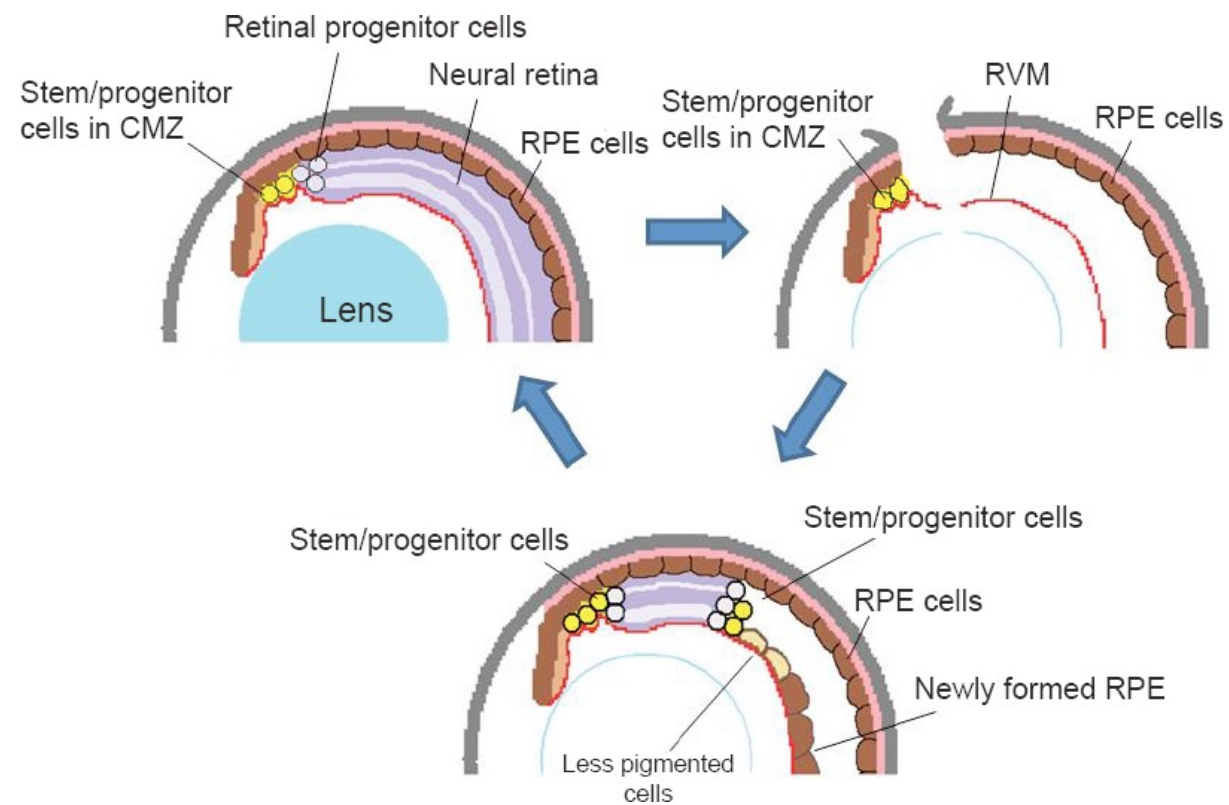
Retinal stem cells in the ciliary marginal zone



Regeneration in *Xenopus tropicalis*



Xenopus tropicalis



Cell Identification of a Proliferating Marginal Zone of Retinal Progenitors in Postnatal Chickens

Developmental Biology 220, 197–210 (2000)

doi:10.1006/dbio.2000.9640, available online at <http://www.idealibrary.com> on IDEAL®



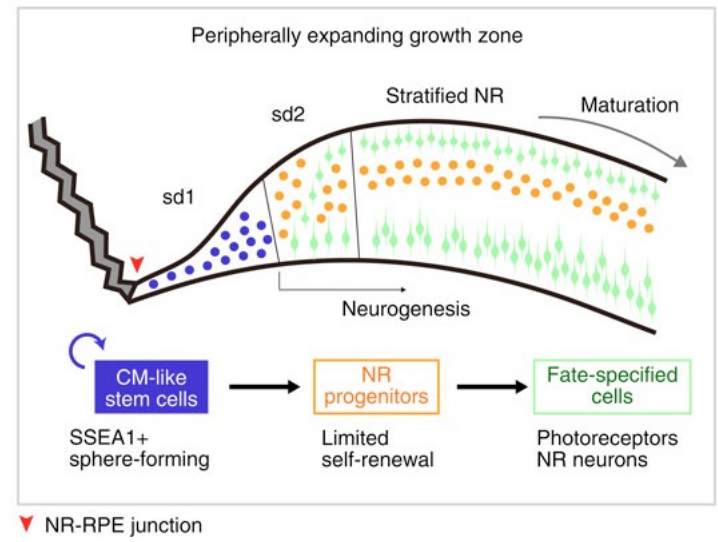
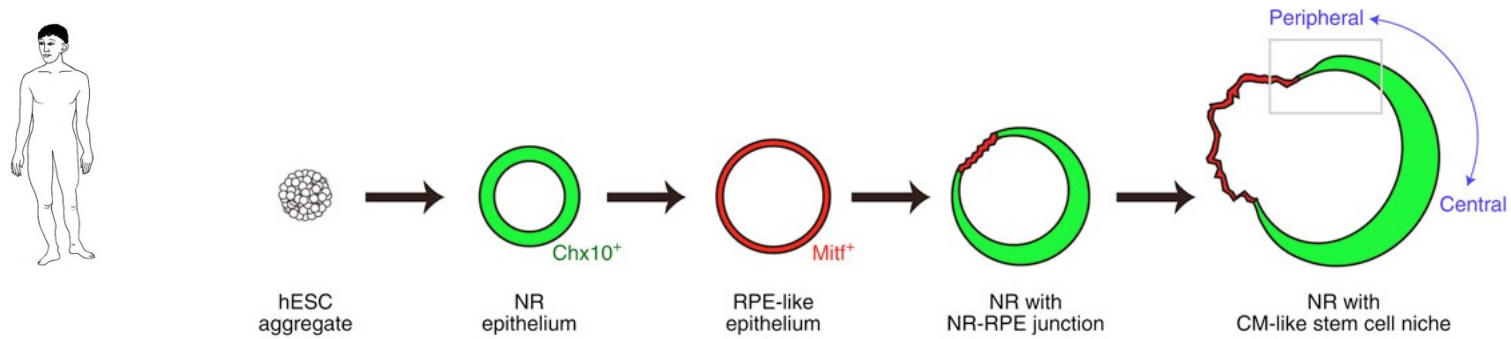
Identification of a Proliferating Marginal Zone of Retinal Progenitors in Postnatal Chickens

Andy J. Fischer and Thomas A. Reh¹

Department of Biological Structure, University of Washington, Seattle, Washington 98195

Only amacrine and bipolar cells are produced by progenitors at the retinal margin of postnatal chicks

Generation of a ciliary margin-like stem cell niche from self-organizing human retinal tissue



nature communications

Article

Single-cell analyses reveal transient retinal progenitor cells in the ciliary margin of developing human retina

<https://doi.org/10.1038/s41467-024-47933-x>

Birthe Dorgau^{1,5}, Joseph Collin^{1,5}, Agata Rozanska¹, Darin Zert^{1,2},
Adrienne Unsworth¹, Moira Crosier¹, Rafiqul Hussain¹, Jonathan Coxhead¹,
Rachel Queen¹ ✉ & Jane C. Sowden³, David R. FitzPatrick^{1,4}

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Cell Reports

Volume 44, Issue 4, 22 April 2025, 115460

Article

Ciliary marginal zone of the developing human retina maintains retinal progenitor cells until late gestational stages

Kiara C. Eldred¹, Sierra J. Edgerton¹, Isabel Ortuño-Lizarán^{1,2},
Juliette Wohlschlegel¹, Stephanie M. Sherman¹, Sidnee Petter¹,
Gracious Wyatt-Draher¹, Dawn Hoffer¹, Ian Glass^{3,4}, Anna La Torre⁵, Thomas A.
Reh^{1,6} ✉



Science Translational Medicine

HOME > SCIENCE TRANSLATIONAL MEDICINE > VOL. 17, NO. 791 > IDENTIFICATION AND CHARACTERIZATION OF HUMAN RETINAL STEM CELLS CAPABLE OF RETINAL...

Identification and characterization of human retinal stem cells capable of retinal regeneration

HUI LIU¹, YUNLONG MA¹, NA GAO¹, YUJUN ZHOU^{1,2,3}, AND...

SCIENCE TRANSLATIONAL MEDICINE • 26 Mar 2025 •

+14 authors

[Authors Info & Affiliations](#)

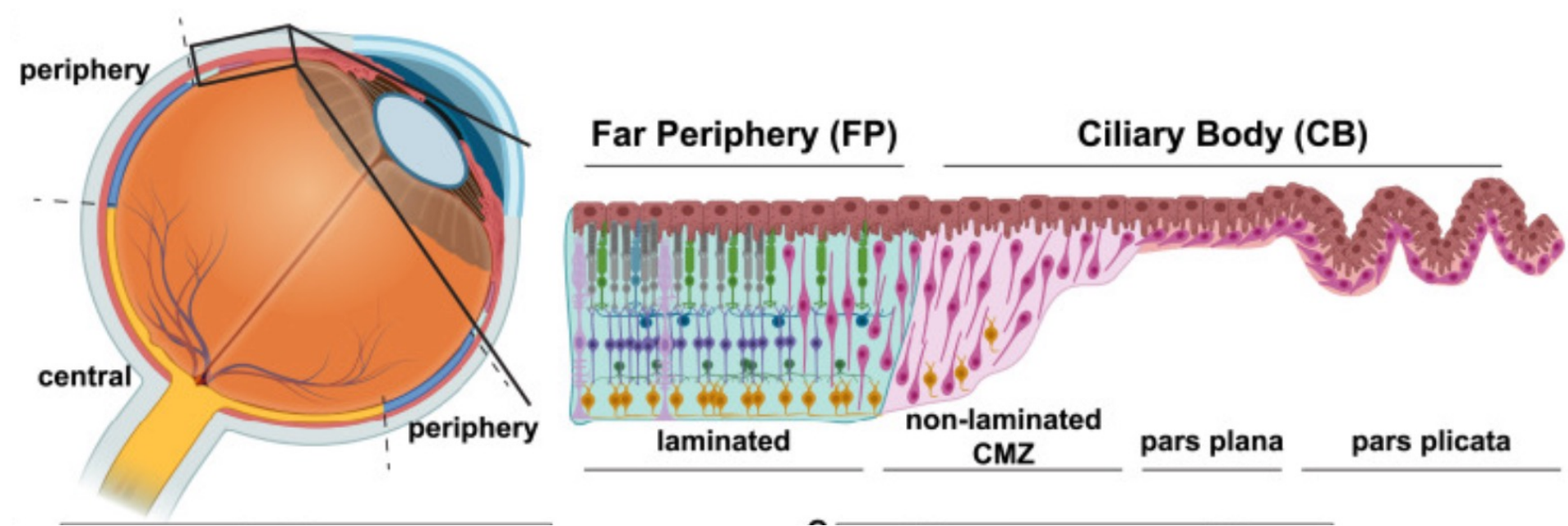
translmed.adp6864



About

More

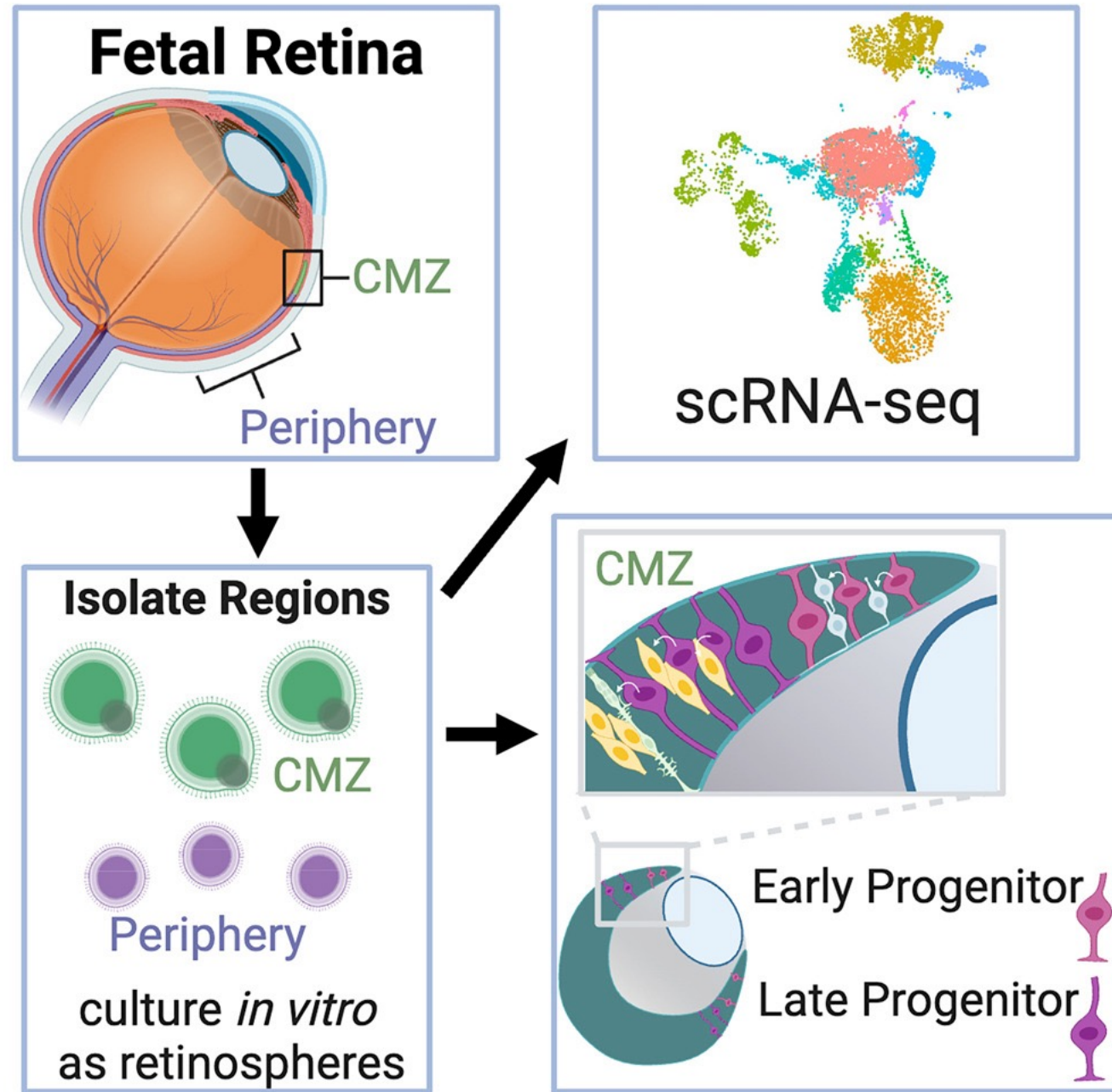
Developing human retina maintains a zone of proliferating cells at the ciliary margin



Cells in the CMZ remain mitotically active after the rest of the retina is no longer proliferating

Early progenitors, located at the tip of the CMZ, give rise to early-born cell types such as ganglion cells.

Late progenitors, located further out in the far periphery of the retinosphere, give rise to late-born cell types such as rods.



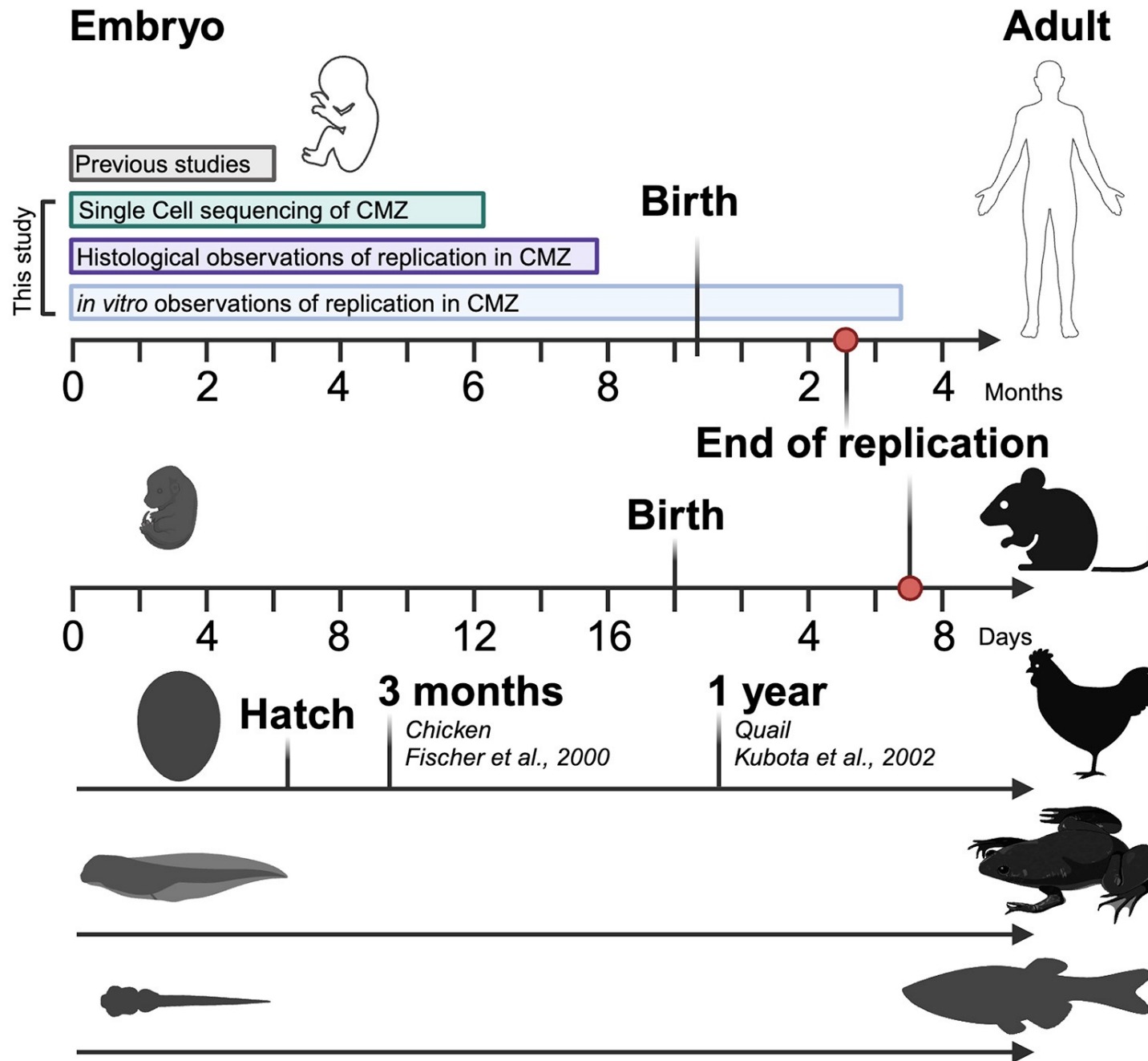
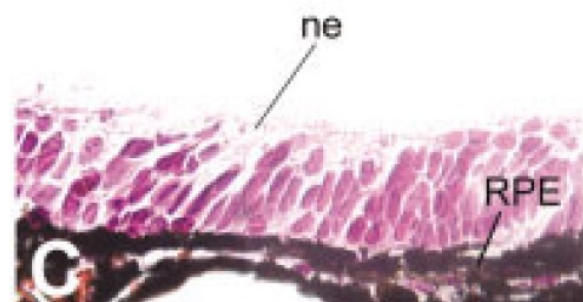
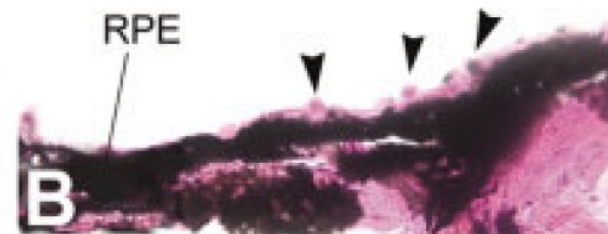
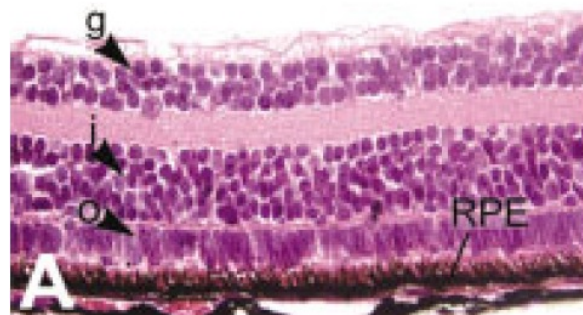


Diagram of timing of CMZ development across organisms

Retinal Pigment Epithelium

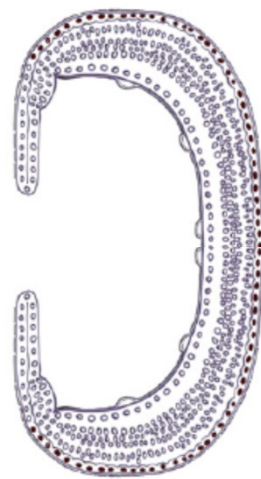
Regeneration in the Newt from the RPE



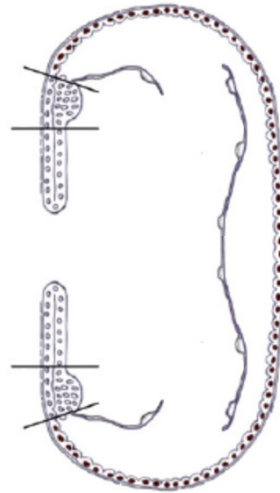
Regeneration in post-metamorphic *Xenopus*



Xenopus laevis



retinectomy



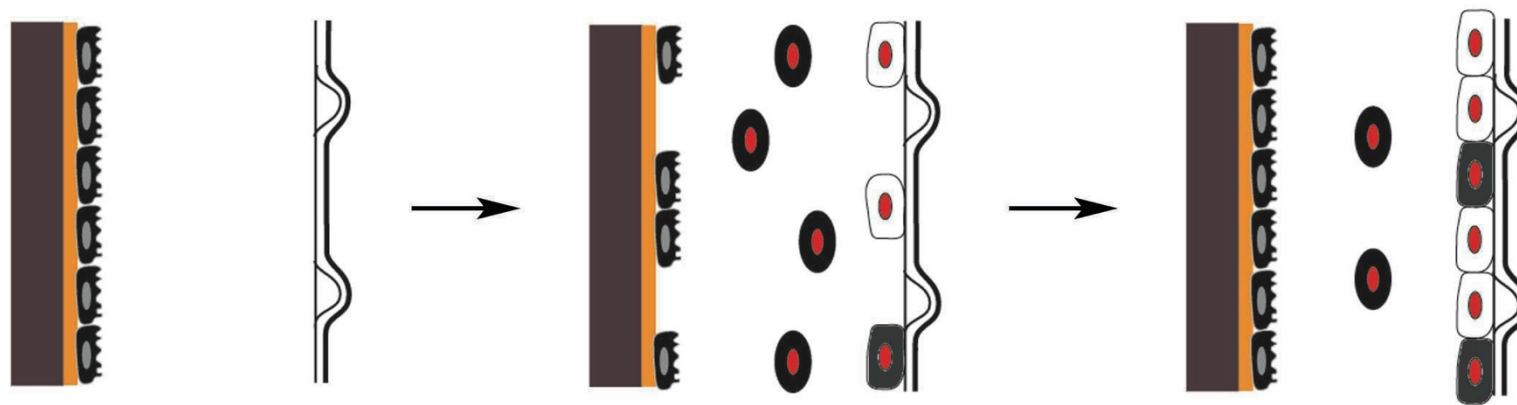
regeneration



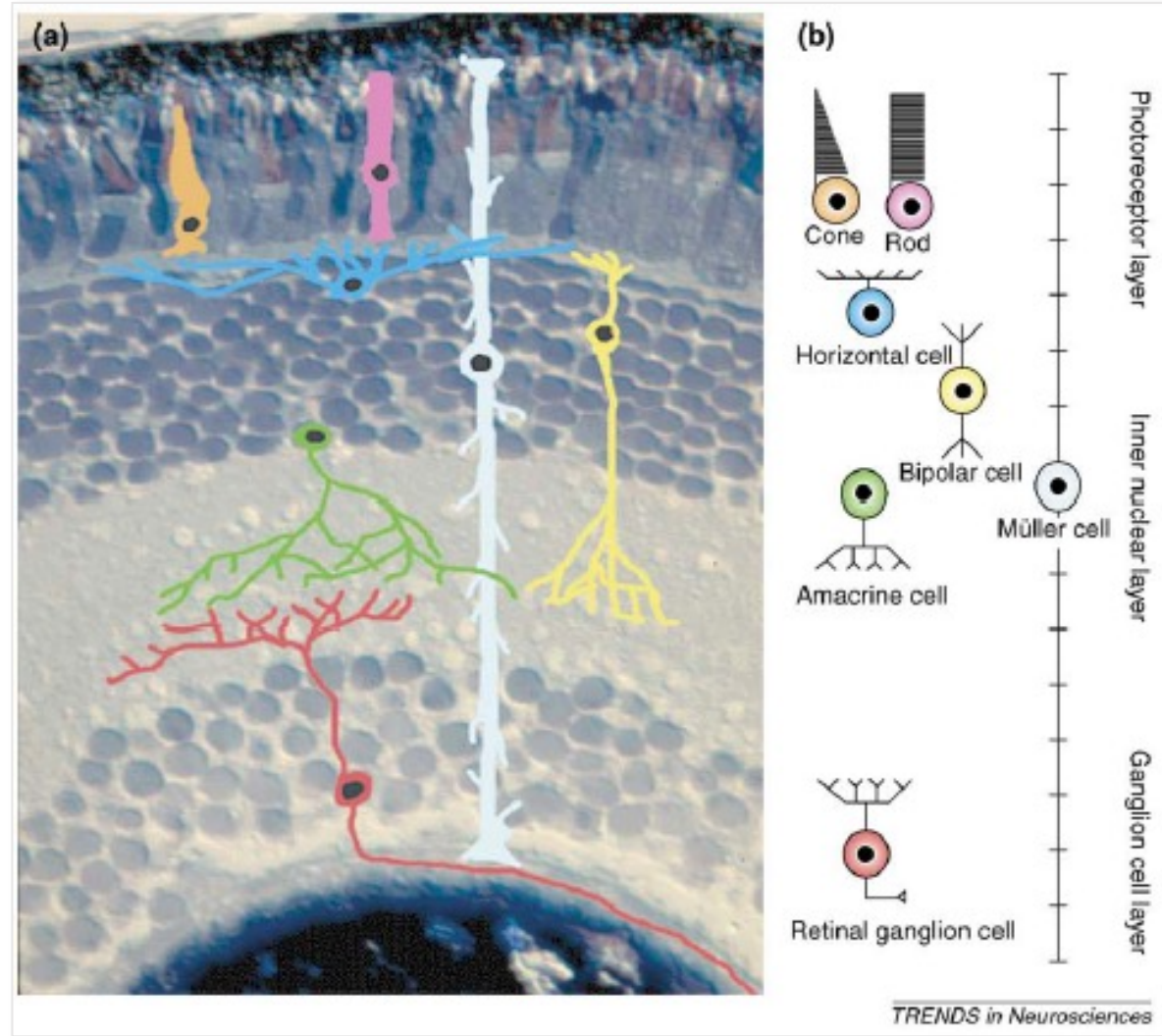
Regeneration in post-metamorphic *Xenopus*



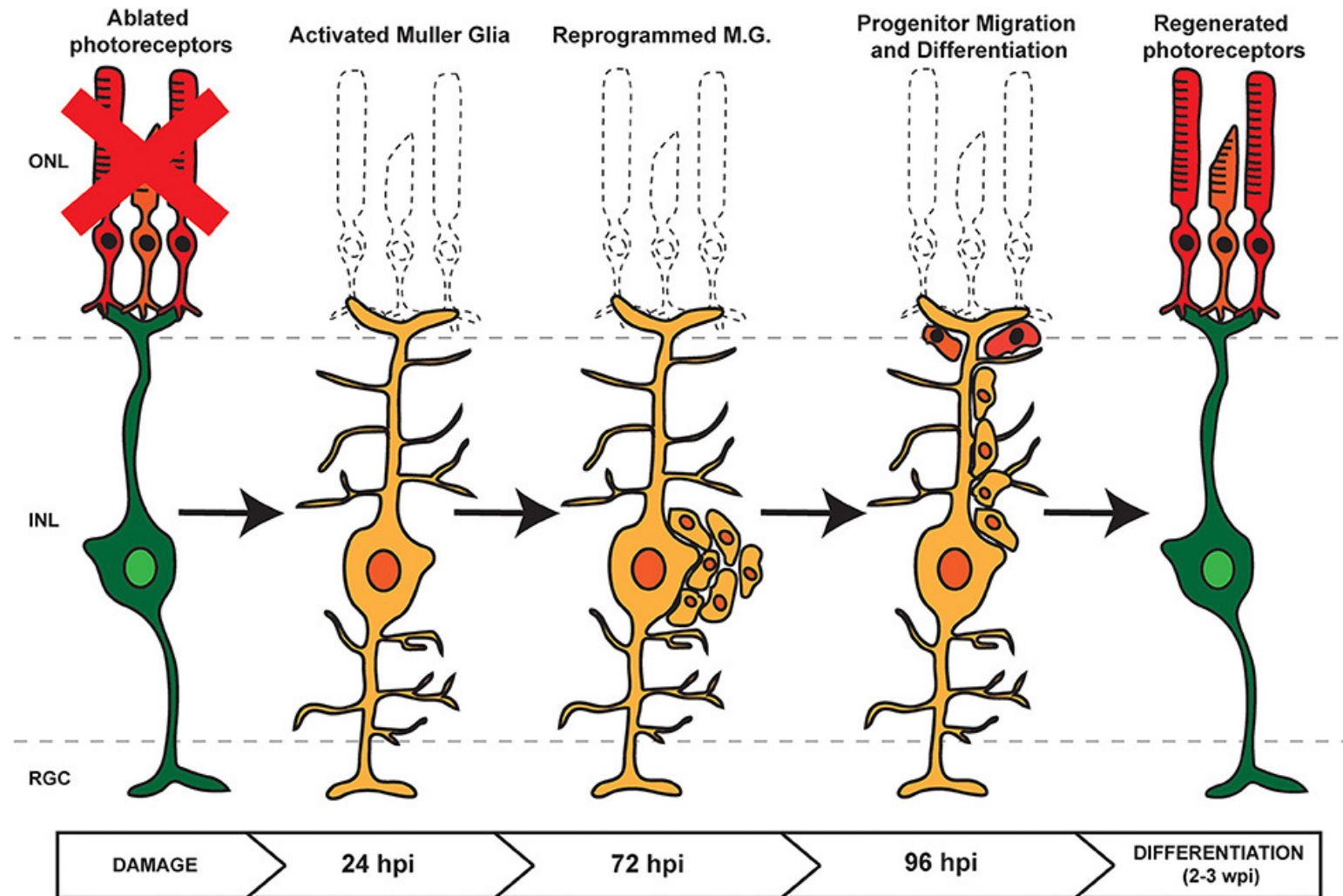
Xenopus laevis



Müller cells

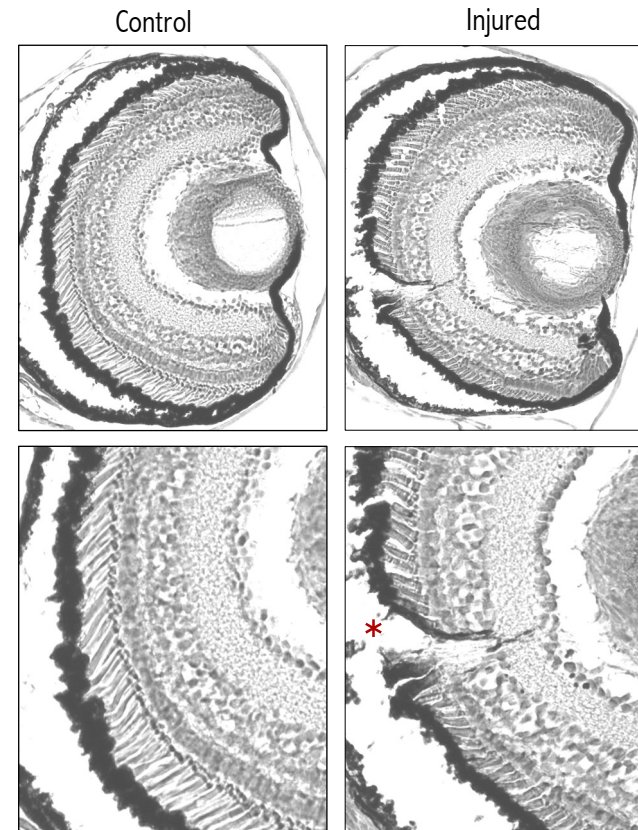
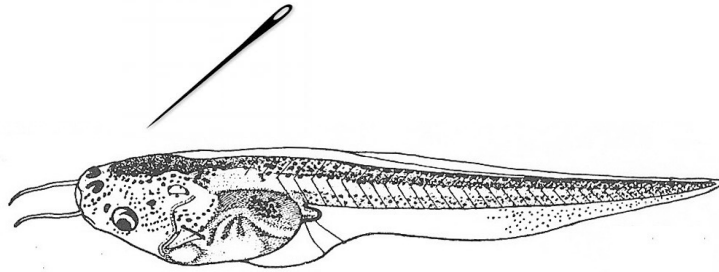


Müller glia regenerative potential in the fish retina

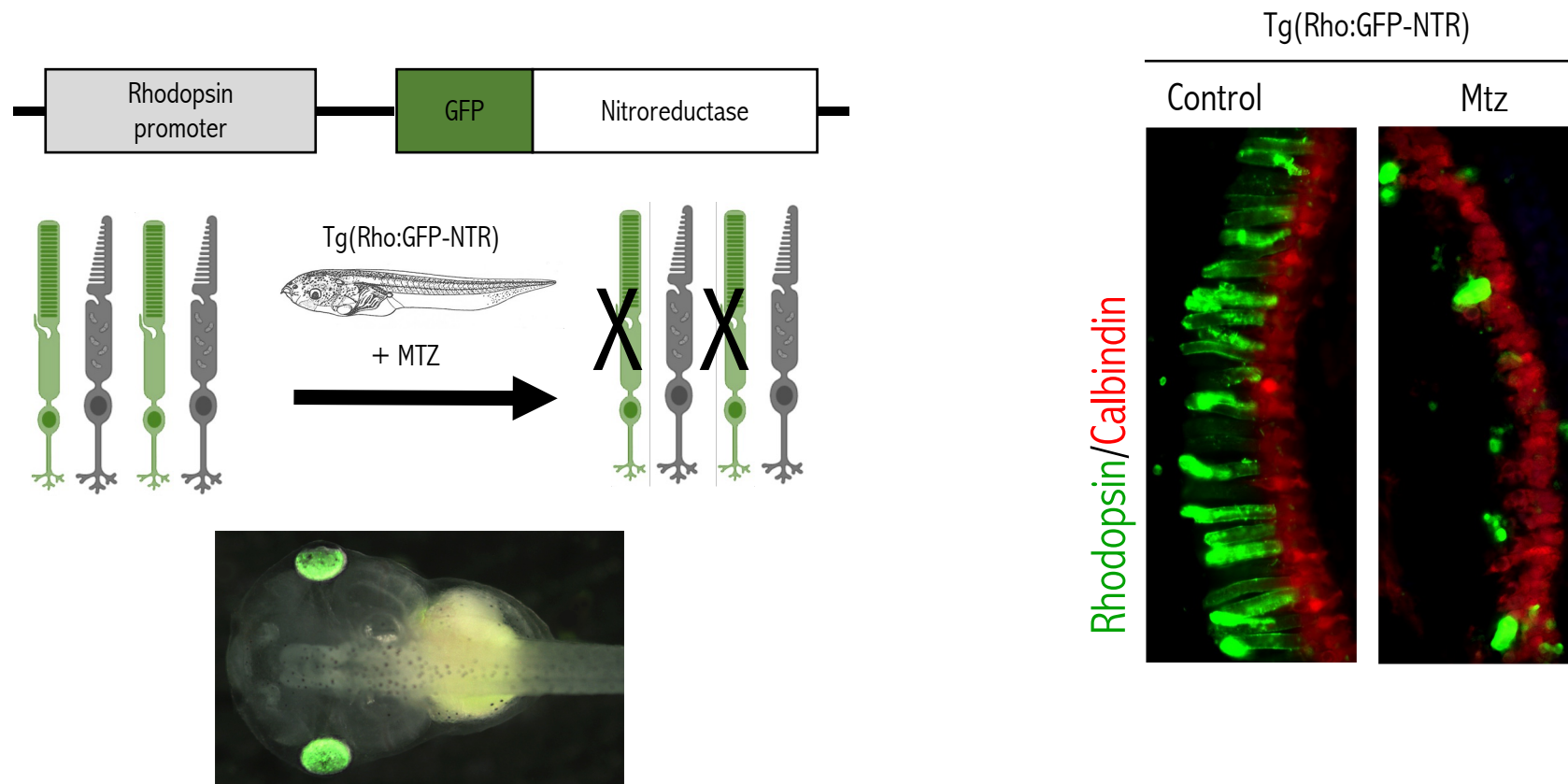


- Can amphibian also recruit their Müller cells in case of injury?
- What are the mechanisms that sustain or constrain Müller cell response to injury?
- Are intrinsic and/or extrinsic factors key regulators of regeneration?
- Can we awake mammalian dormant Müller cells for therapeutic purposes?

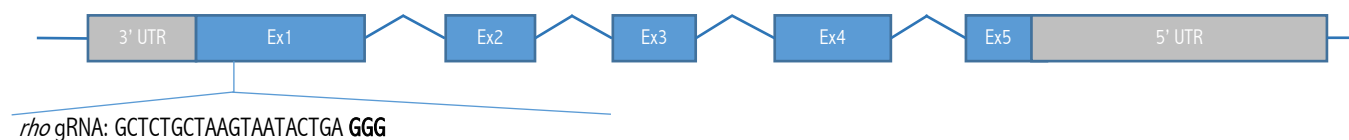
Mechanical injury



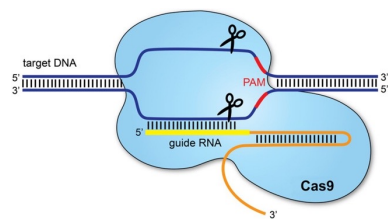
NTR-MTZ Conditional photoreceptor ablation



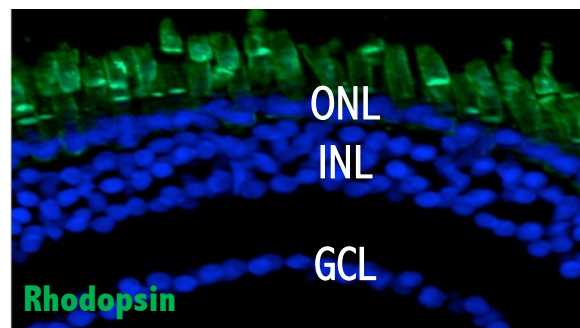
CRISPR-dependent photoreceptor degeneration as a model of retinitis pigmentosa



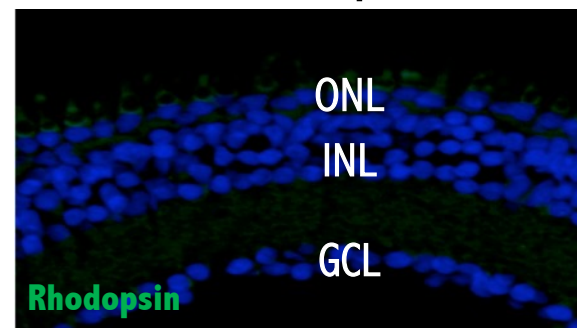
rhodopsin gene



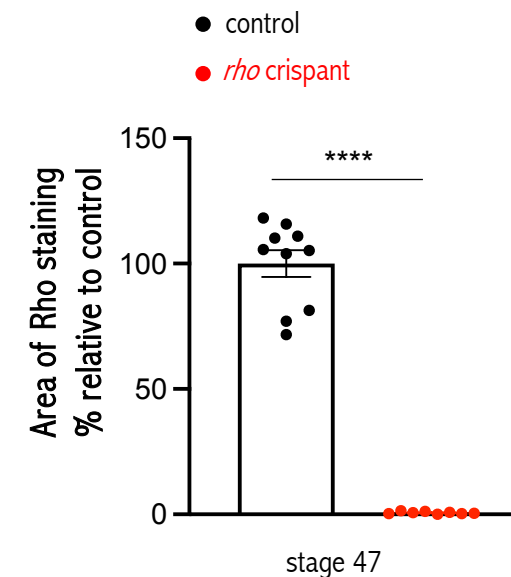
control



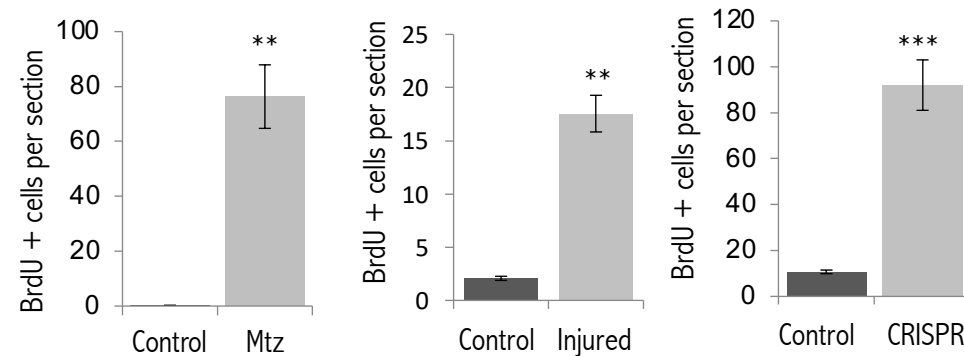
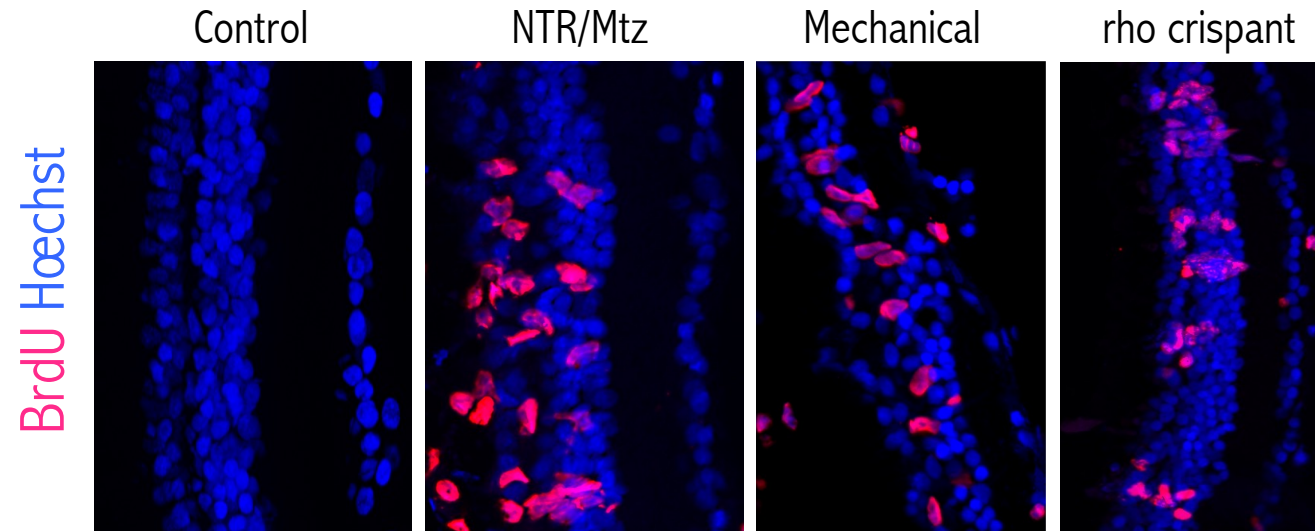
rho crispant



unpublished



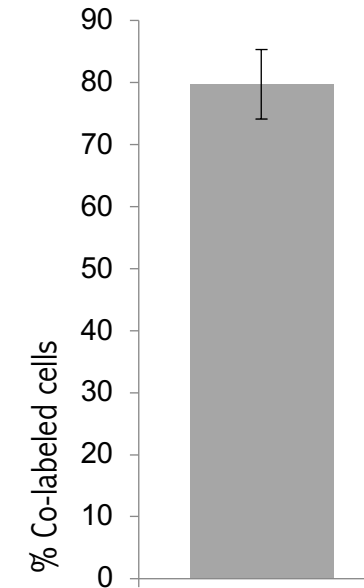
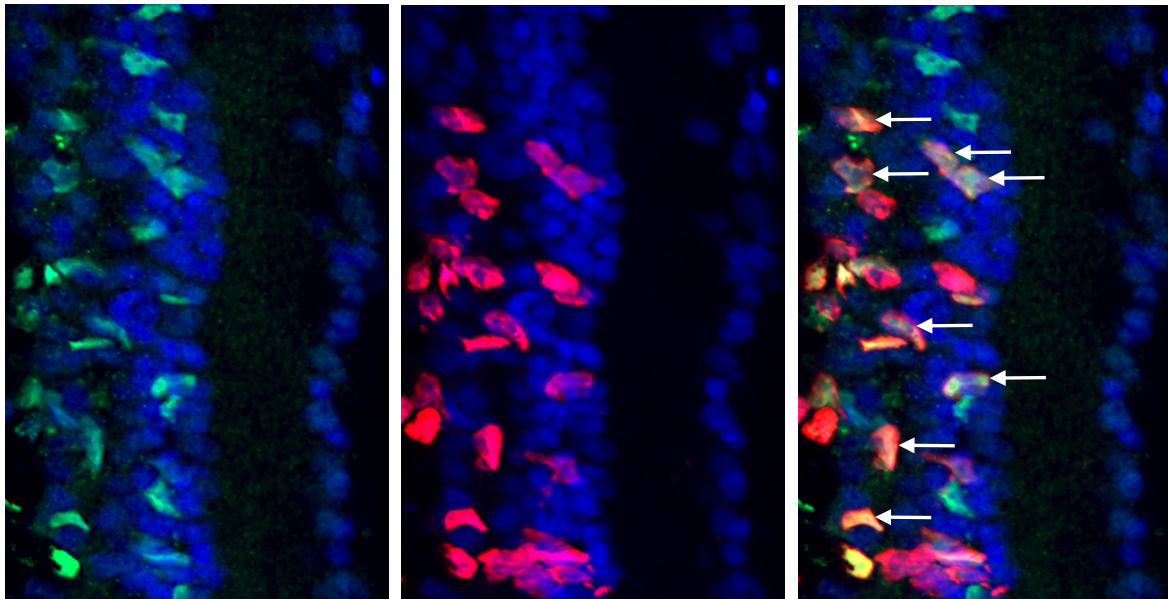
Proliferative response after retinal injury



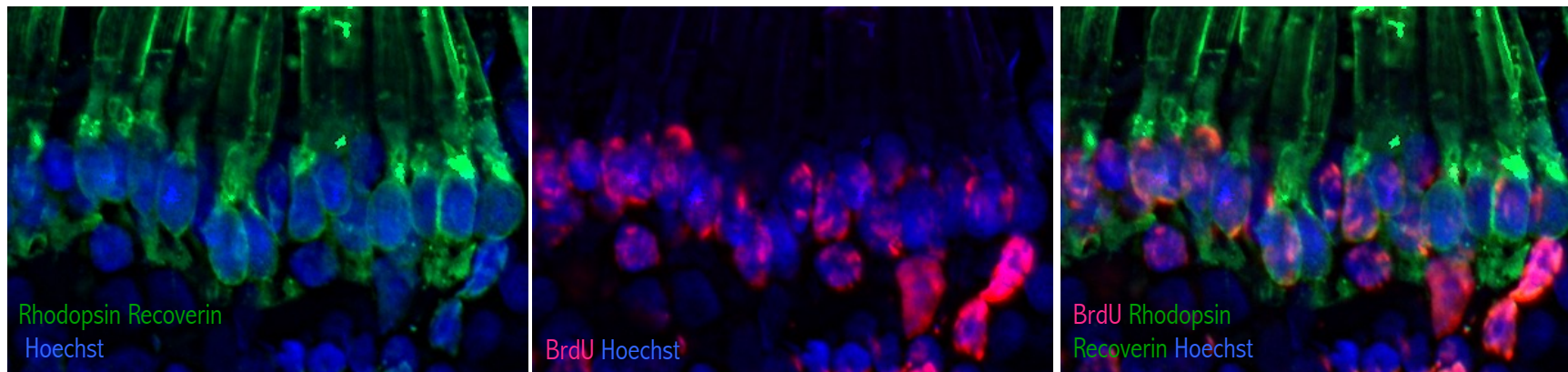
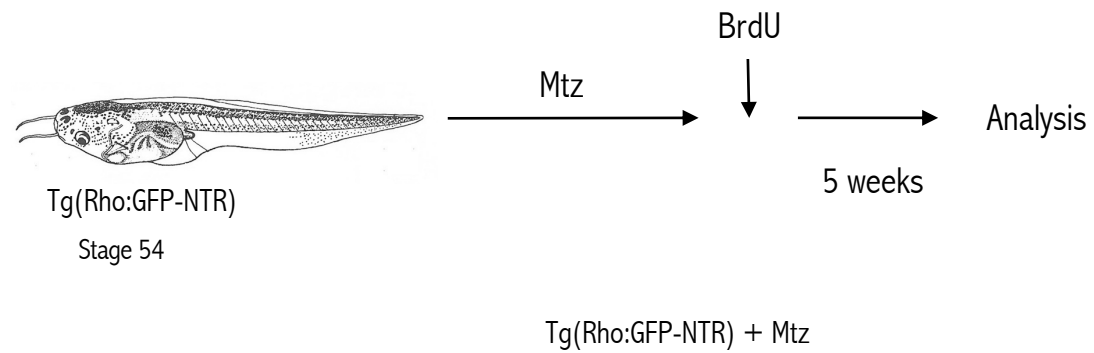
Müller cells proliferate following retinal injury

Tg(Rho:GFP-NTR) + Mtz

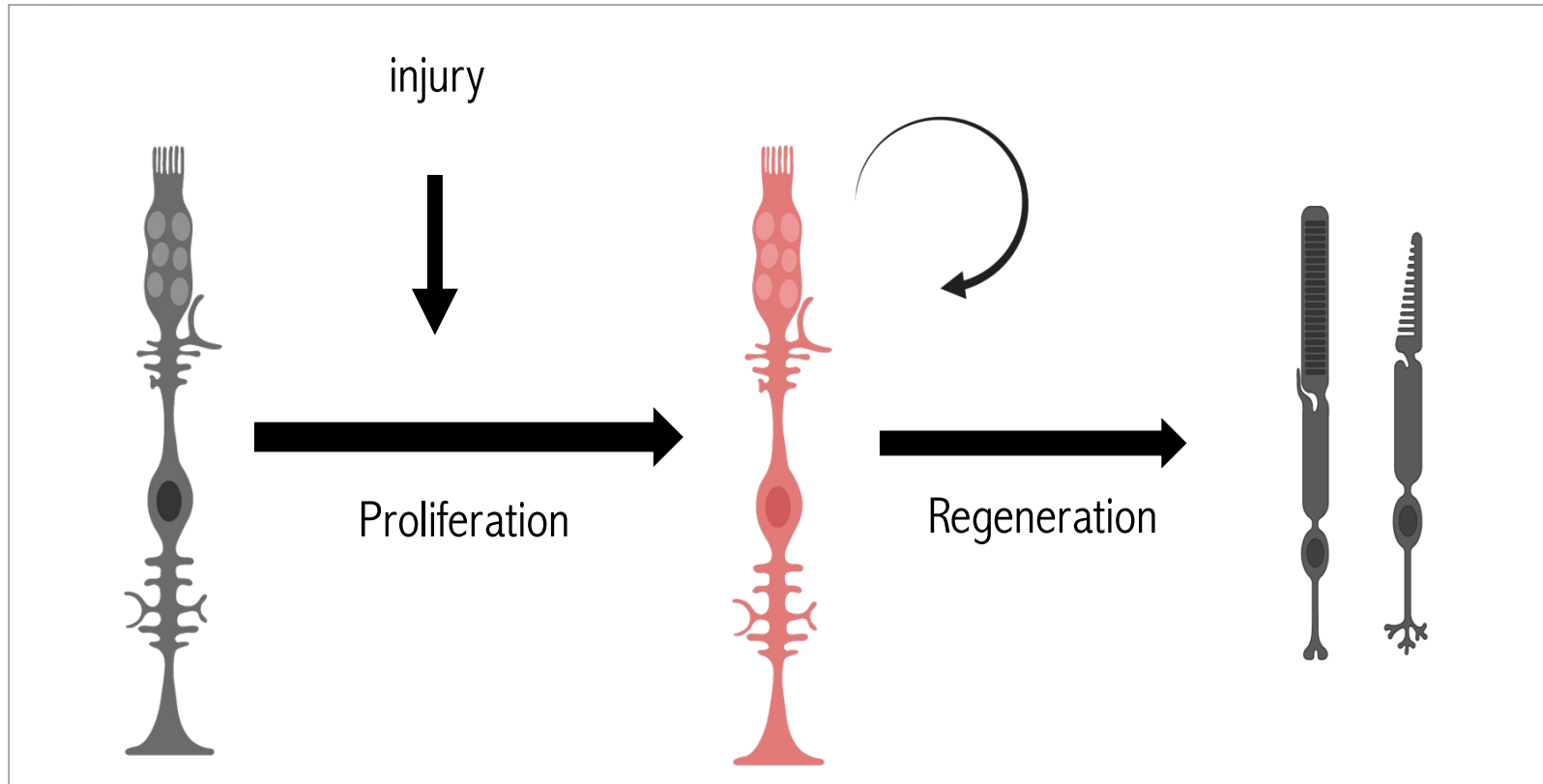
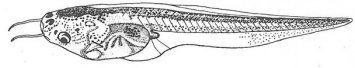
Sox9 BrdU Hoechst



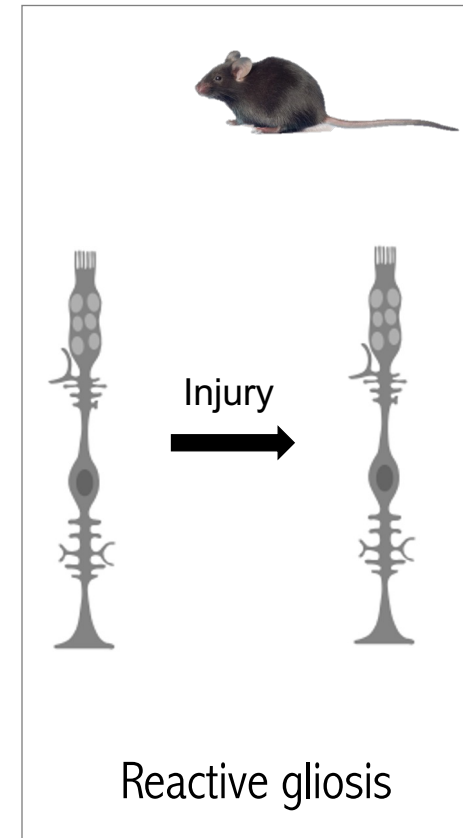
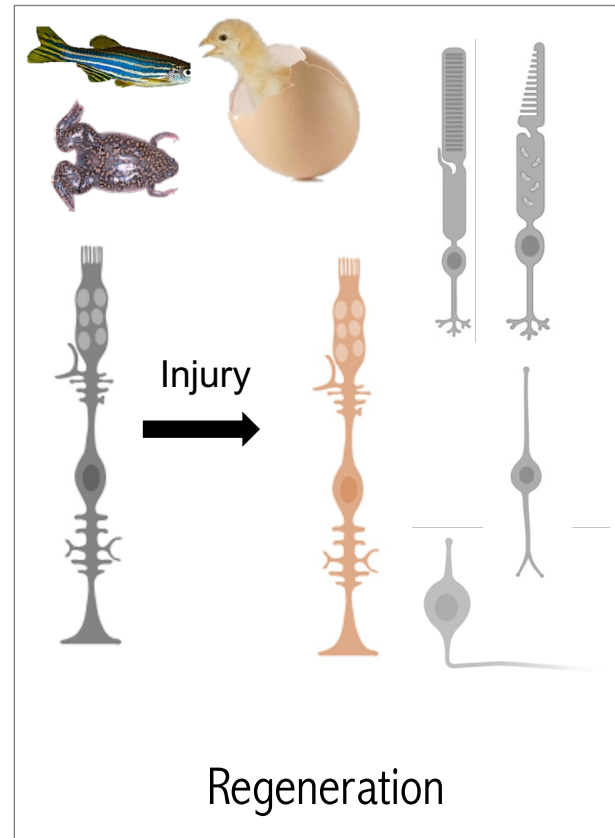
Müller cell-dependent photoreceptor regeneration



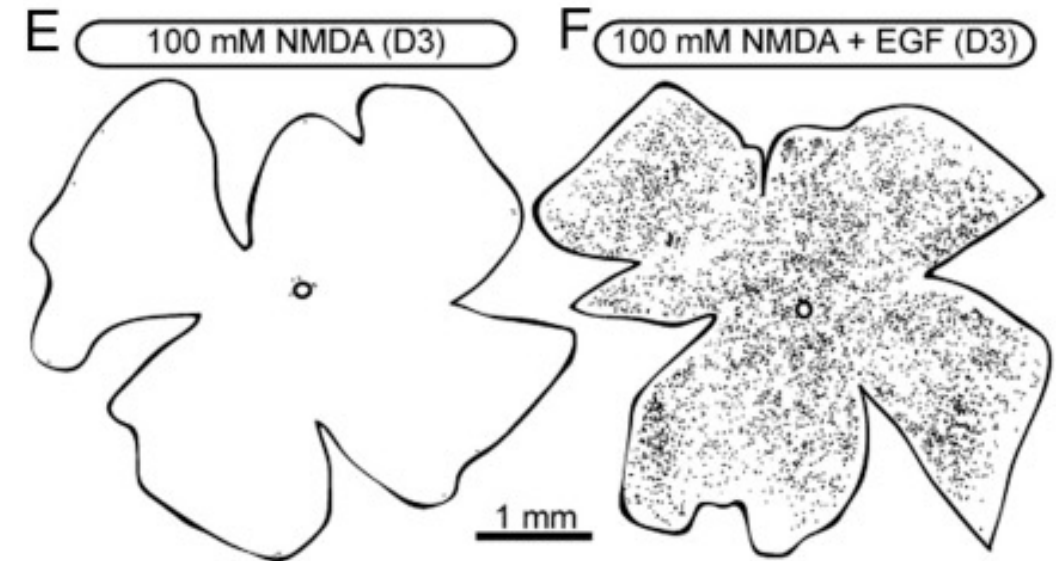
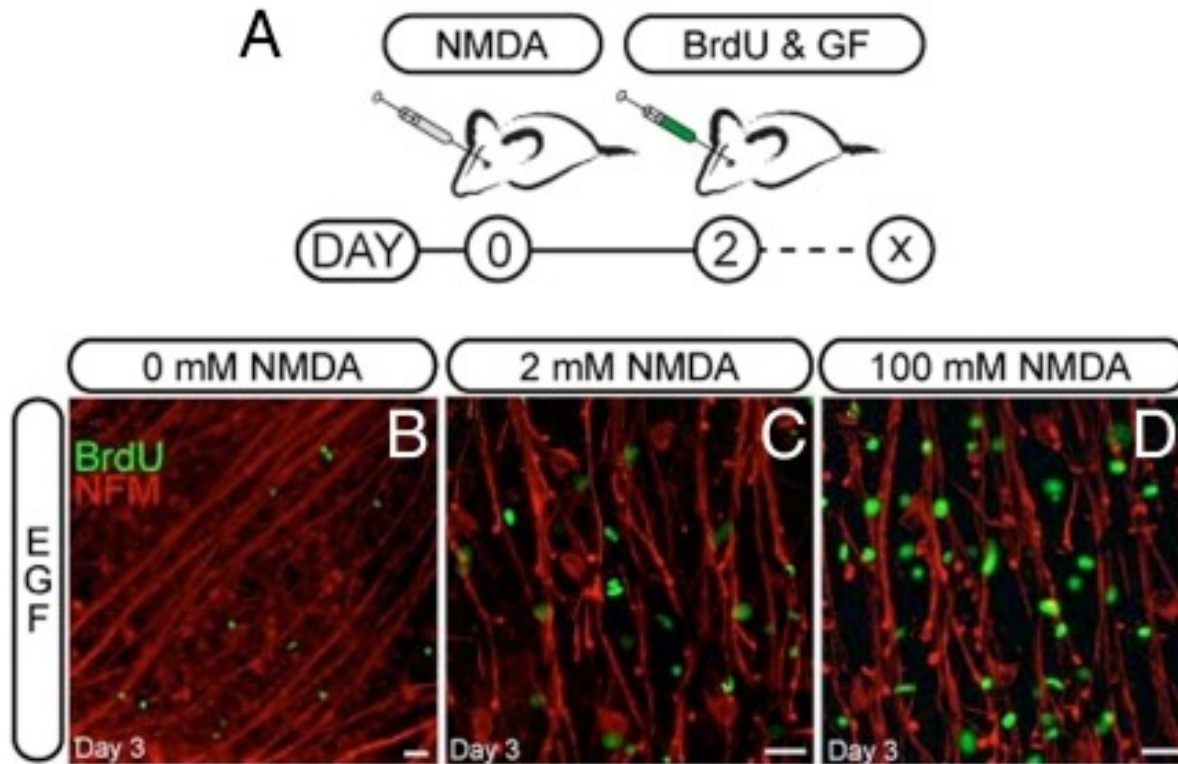
Müller glial cell potential for retinal tissue repair in *Xenopus*



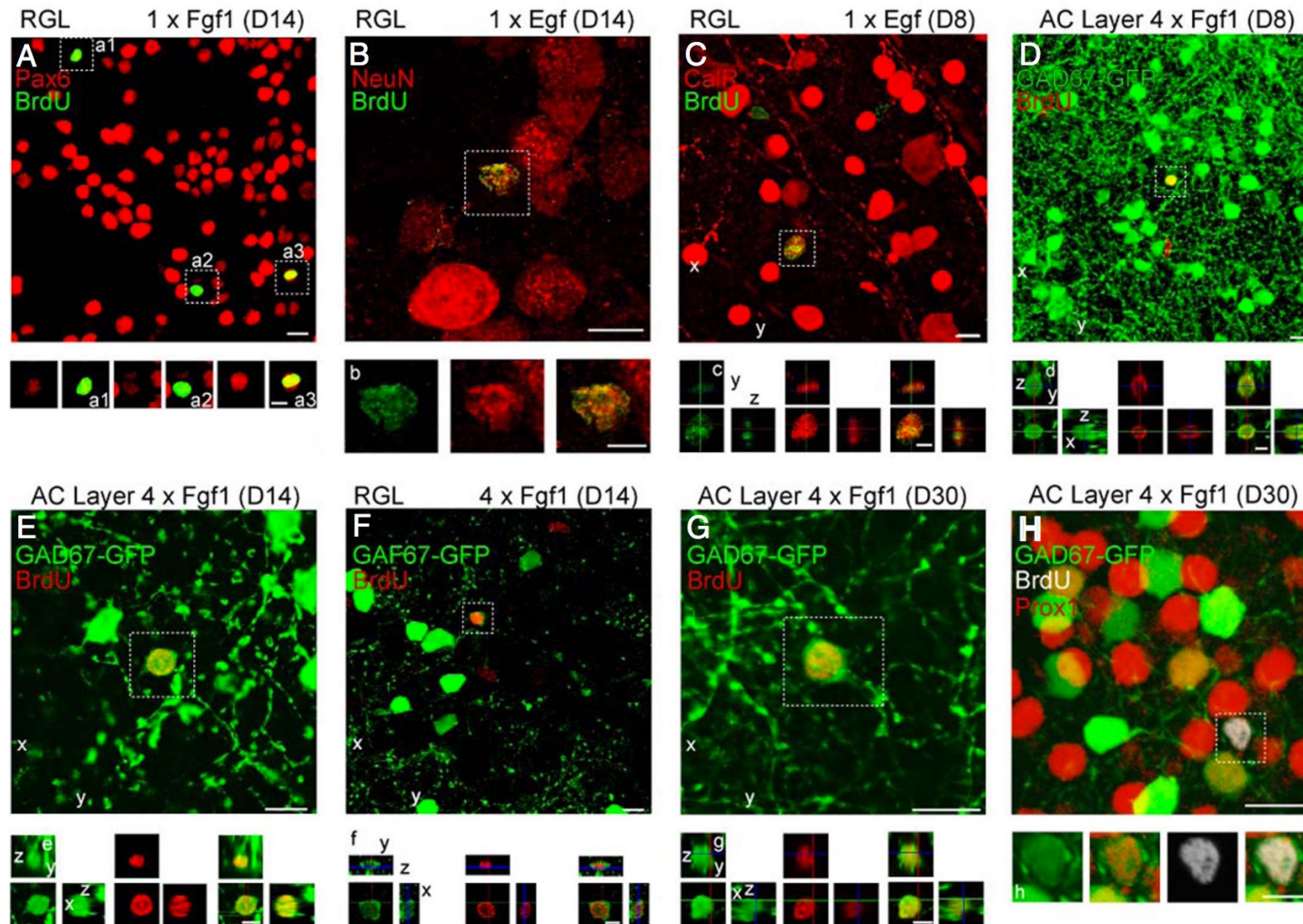
Müller glial cell potential for retinal tissue repair



EGF triggers Müller glia proliferation in adult mouse retina following NMDA damage

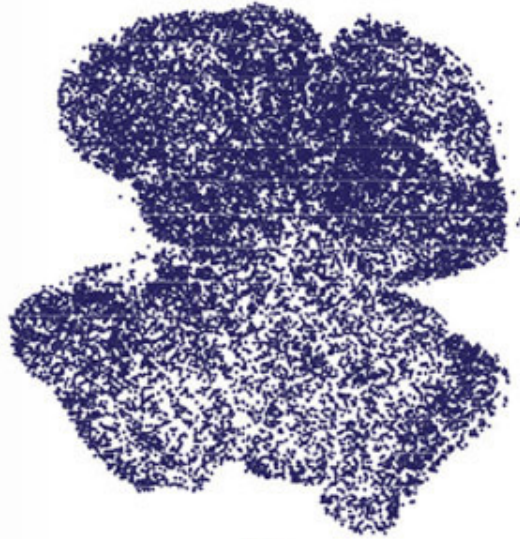


Some of these BrdU+ cells developed characteristics of retinal neurons



MÜLLER GLIA PROLIFERATIVE RESPONSE

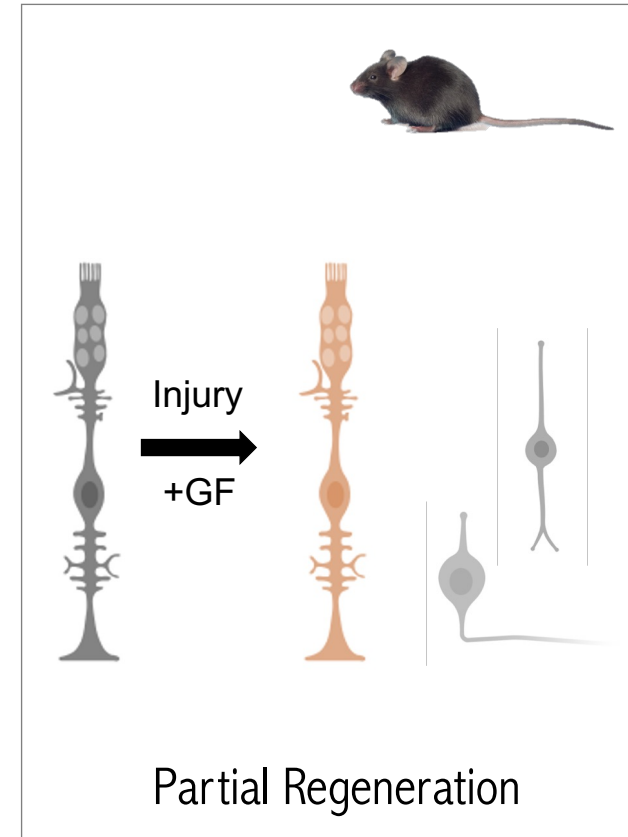
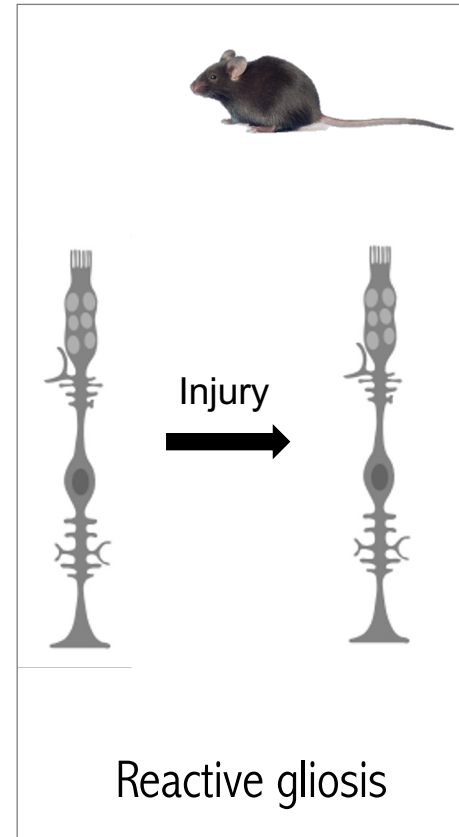
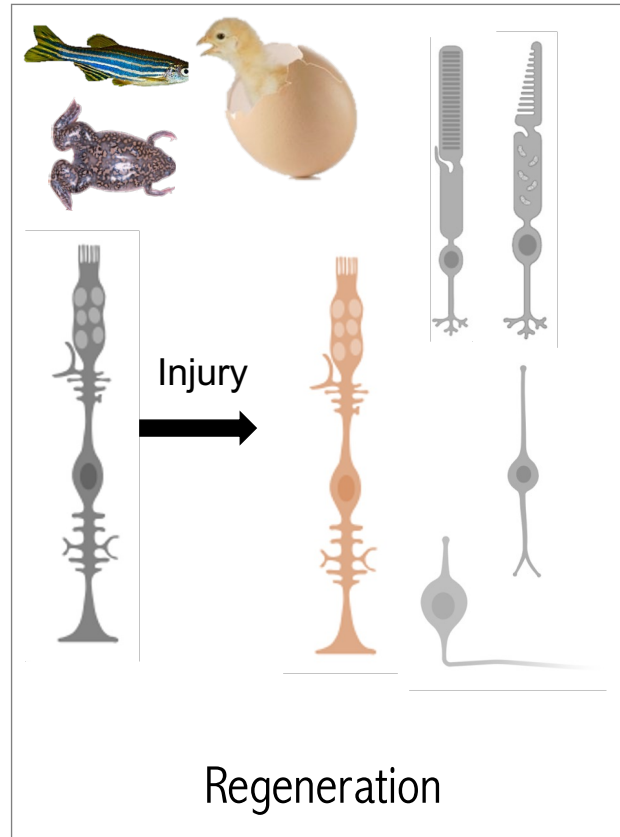
JUVENILE



ADULT

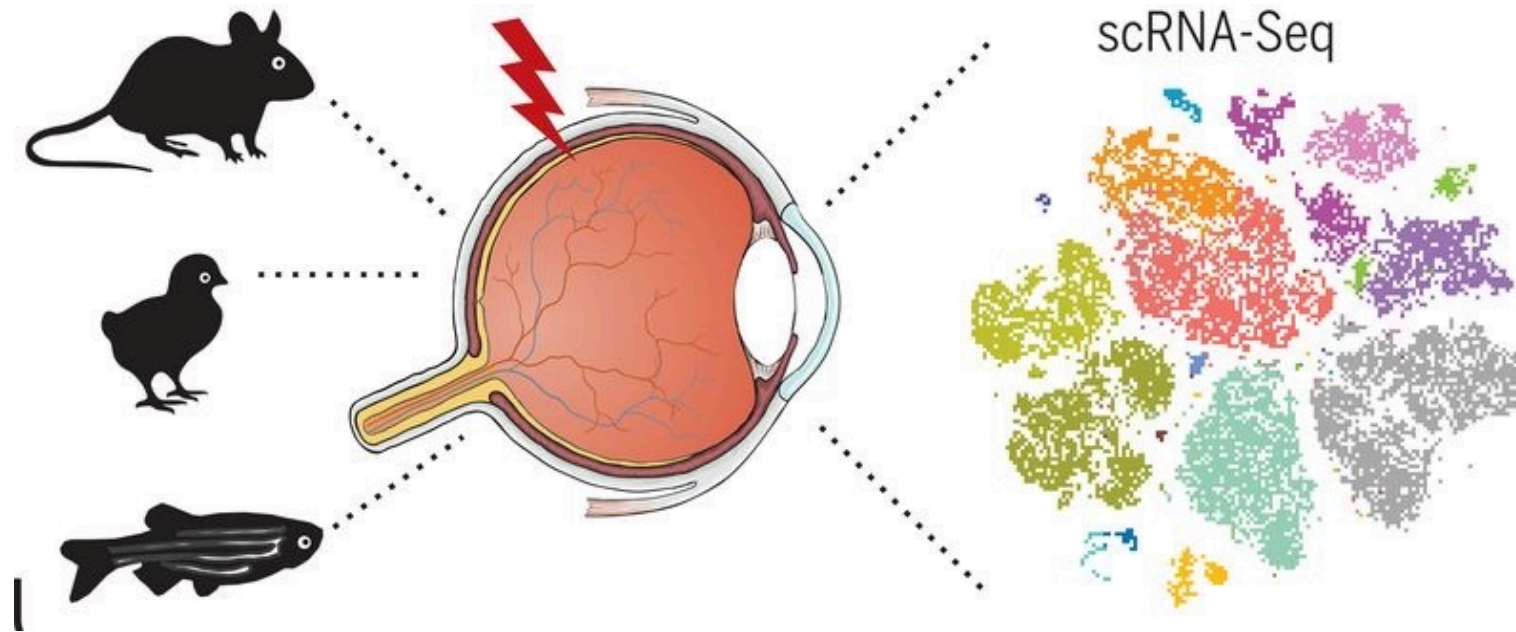


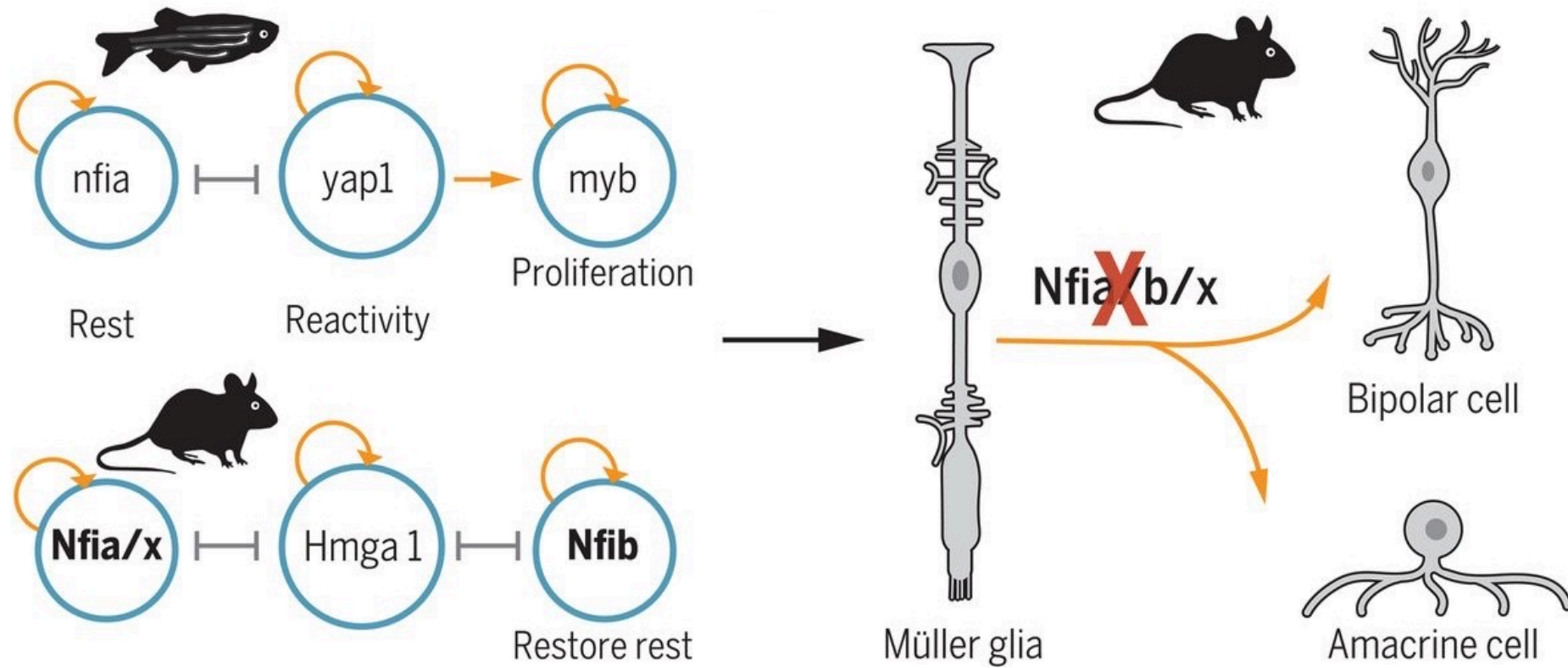
Müller glial cell potential for retinal tissue repair



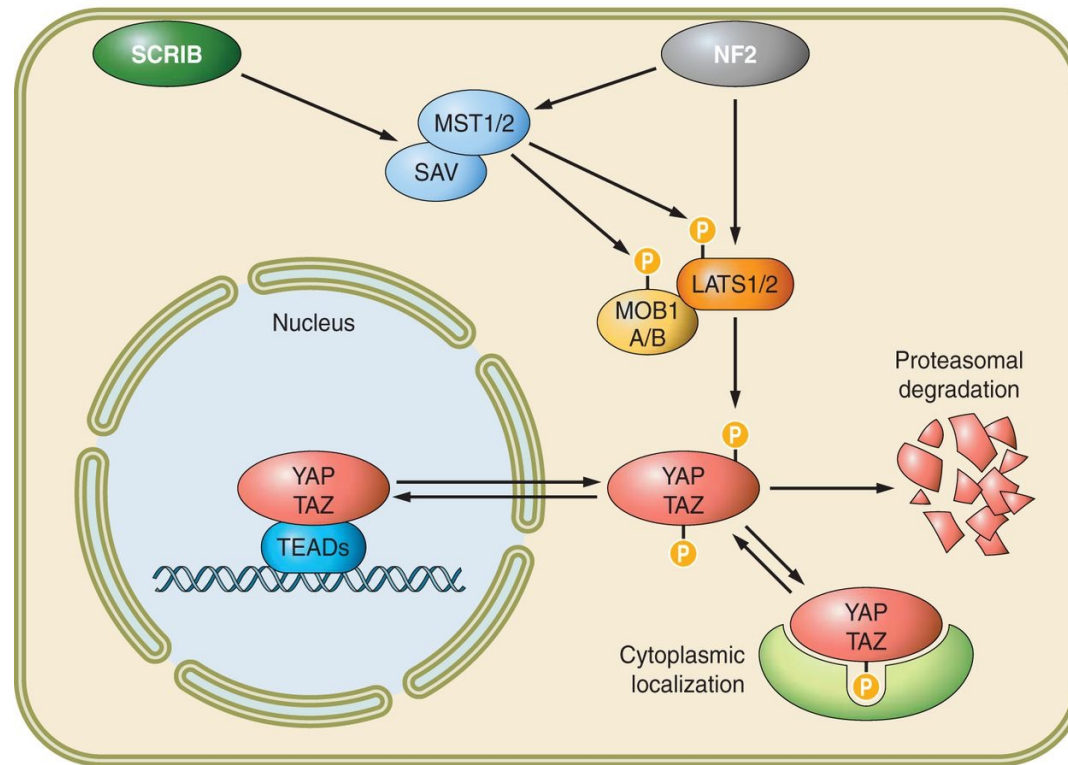


What are the mechanisms that sustain or constrain Müller cell proliferation upon injury?

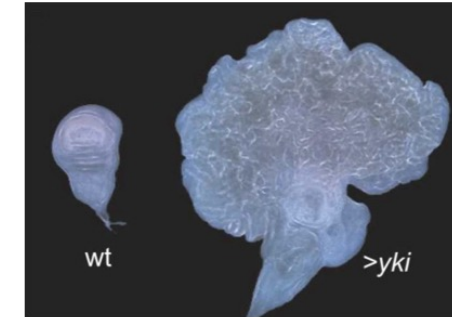




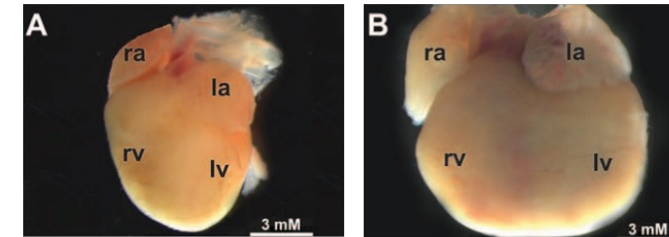
The Hippo pathway



From Piccolo et al., 2014



Todd Heallen et al. 2011

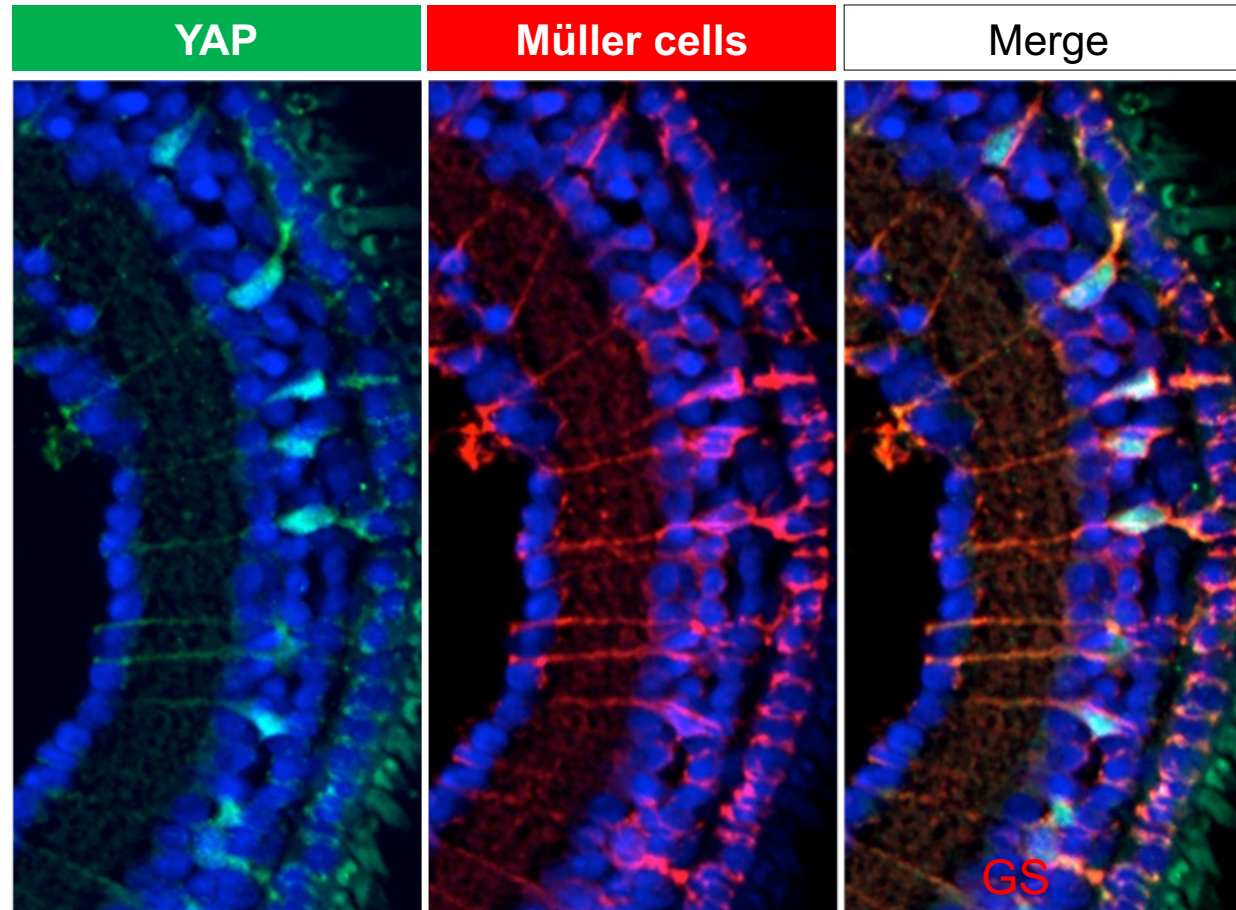


Pan et al. 2007

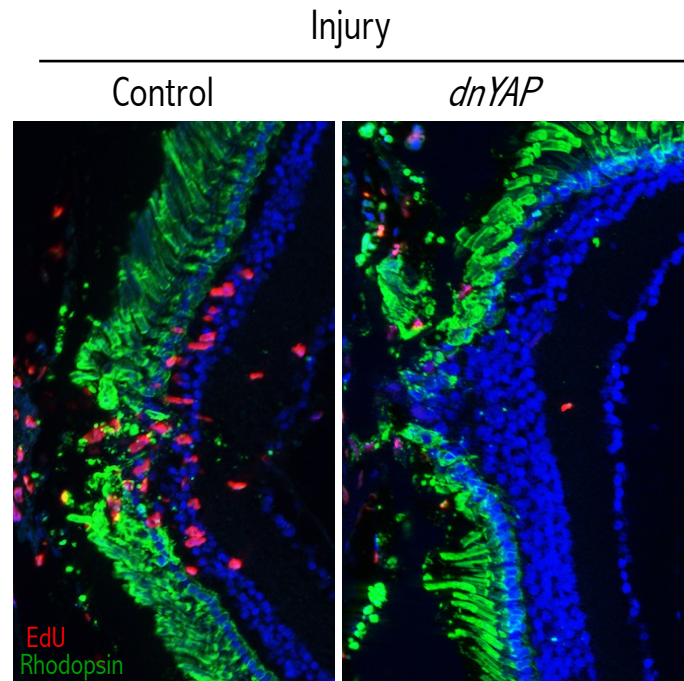
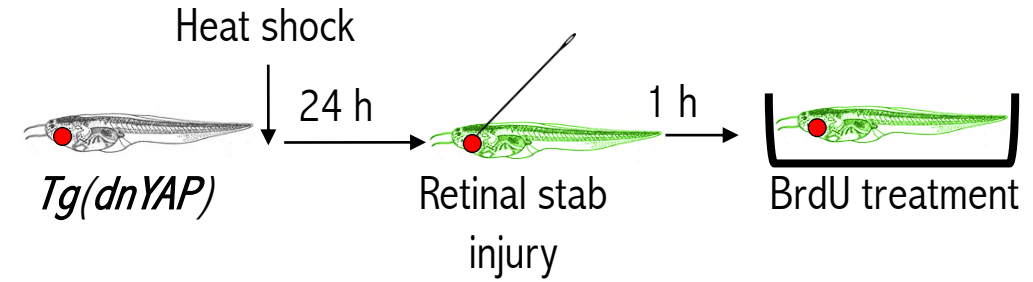
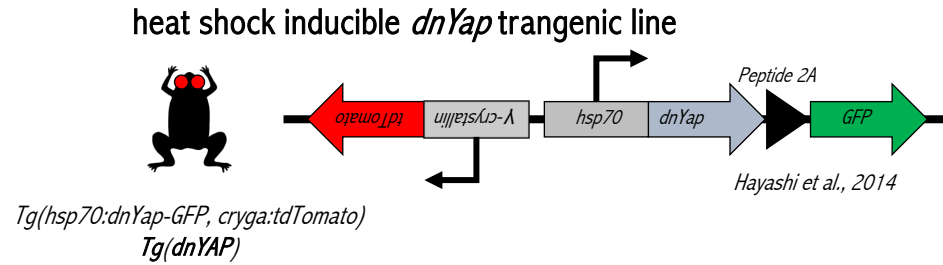


Pan et al. 2007

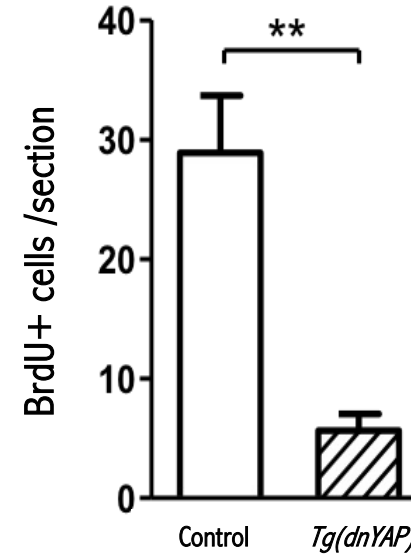
YAP is expressed in *Xenopus* Müller cells



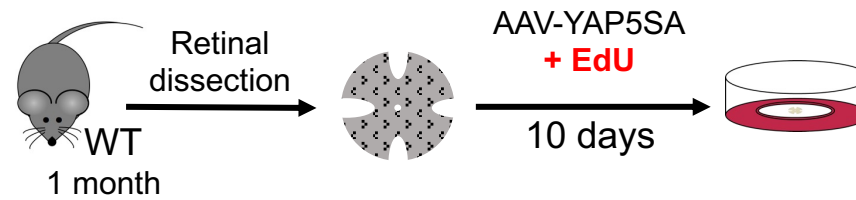
Inhibition of YAP prevents Müller glia proliferation upon acute retinal damage



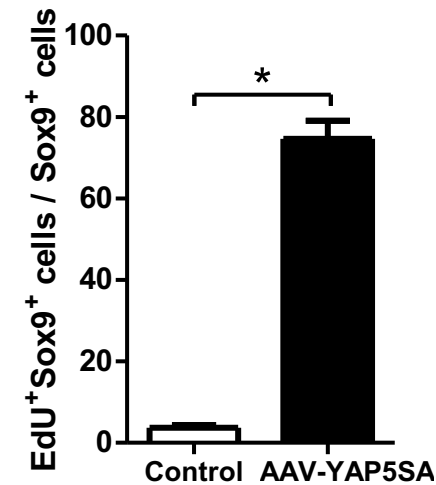
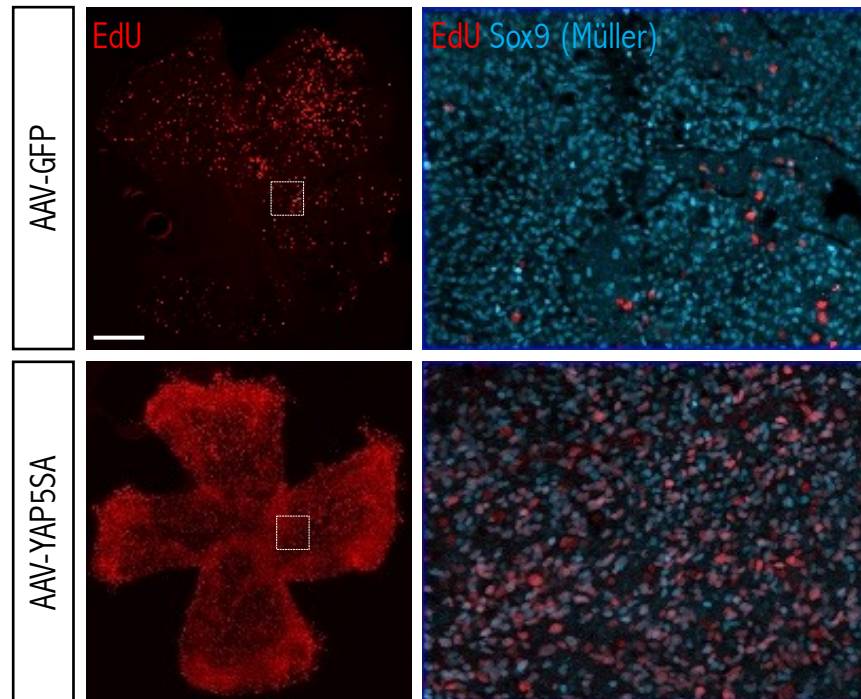
Photoreceptors Proliferation Nuclei



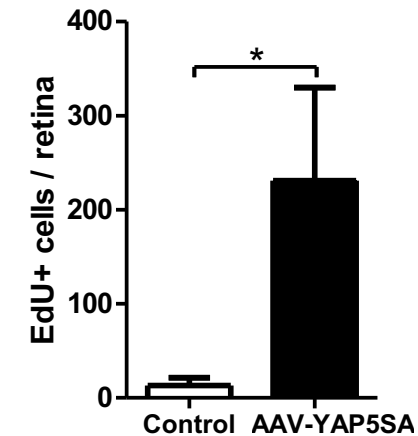
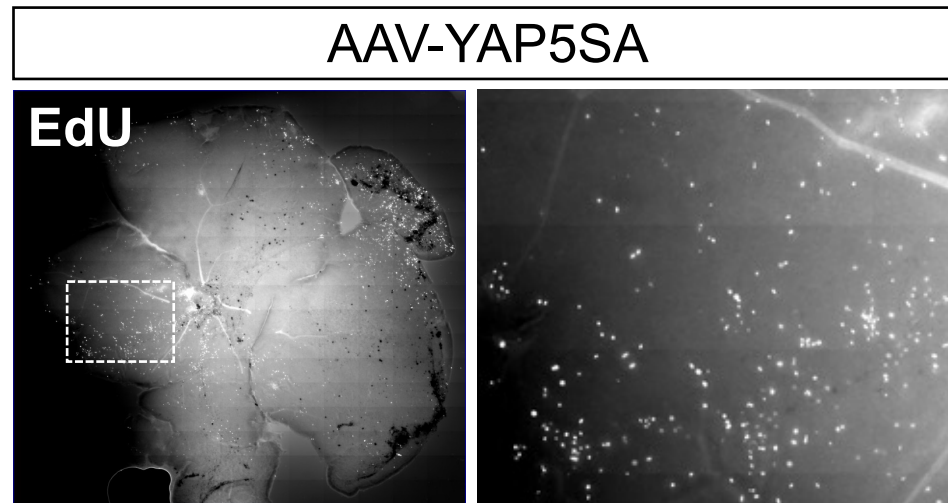
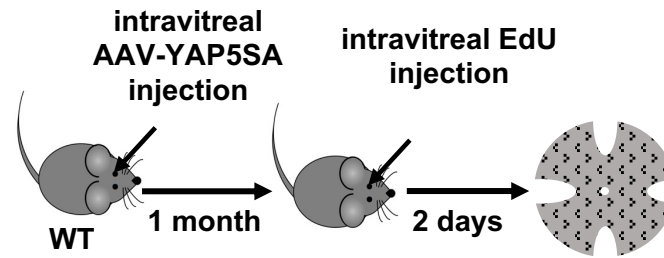
Forced YAP expression in mouse Müller glia cells stimulates their proliferation

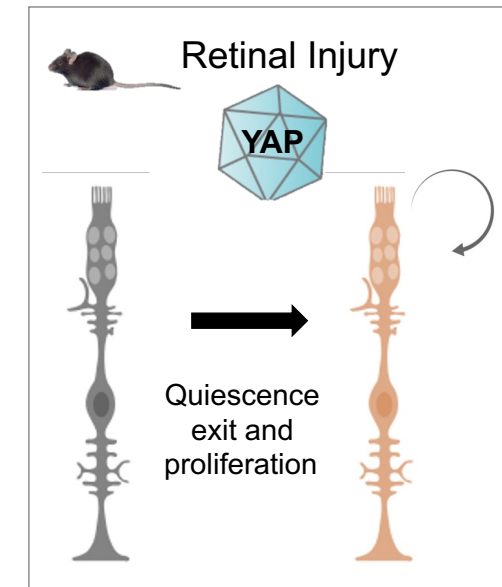
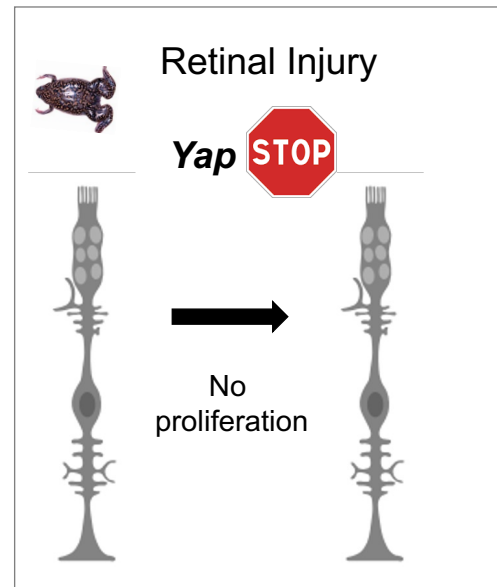
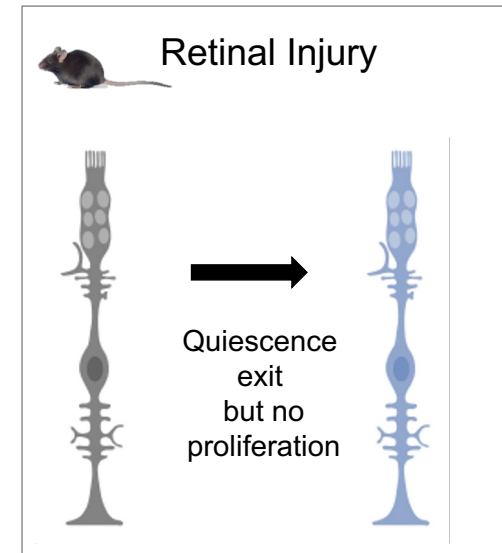
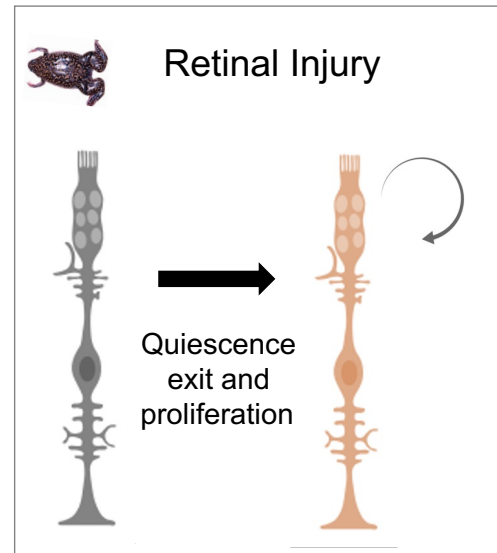


Klimczak et al., 2009



Forced YAP expression in mouse Müller glia cells stimulates their proliferation *in vivo*

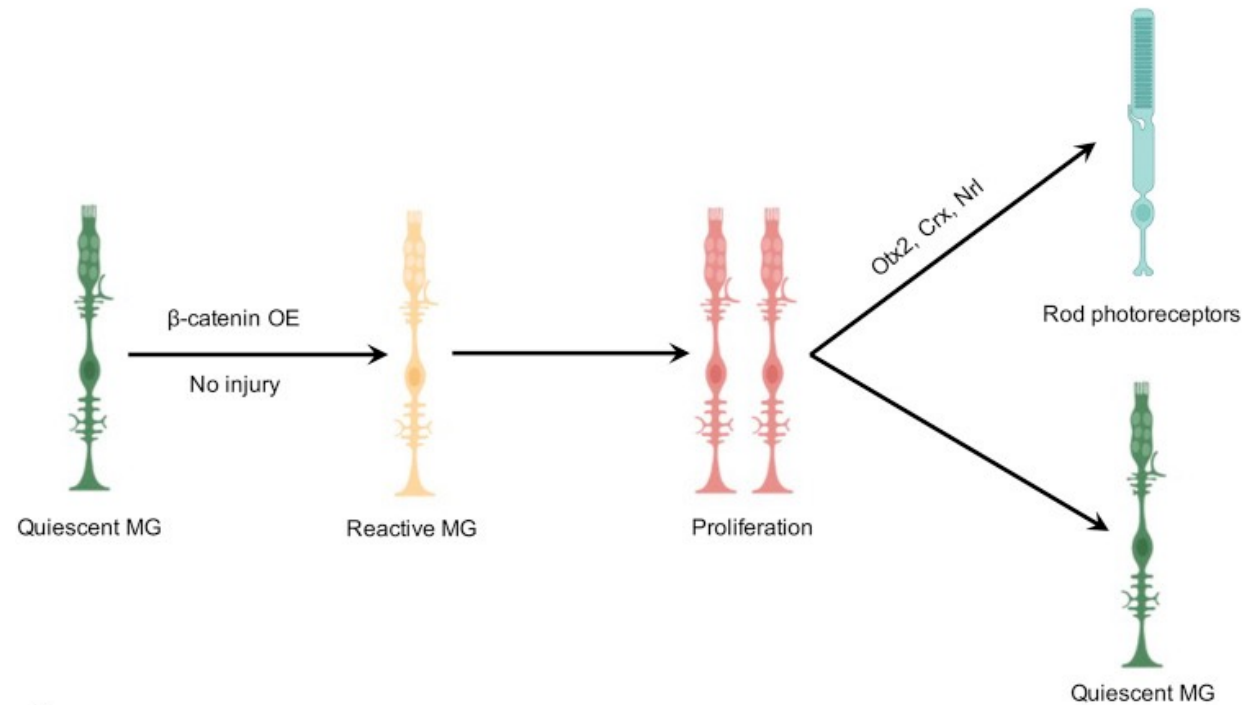




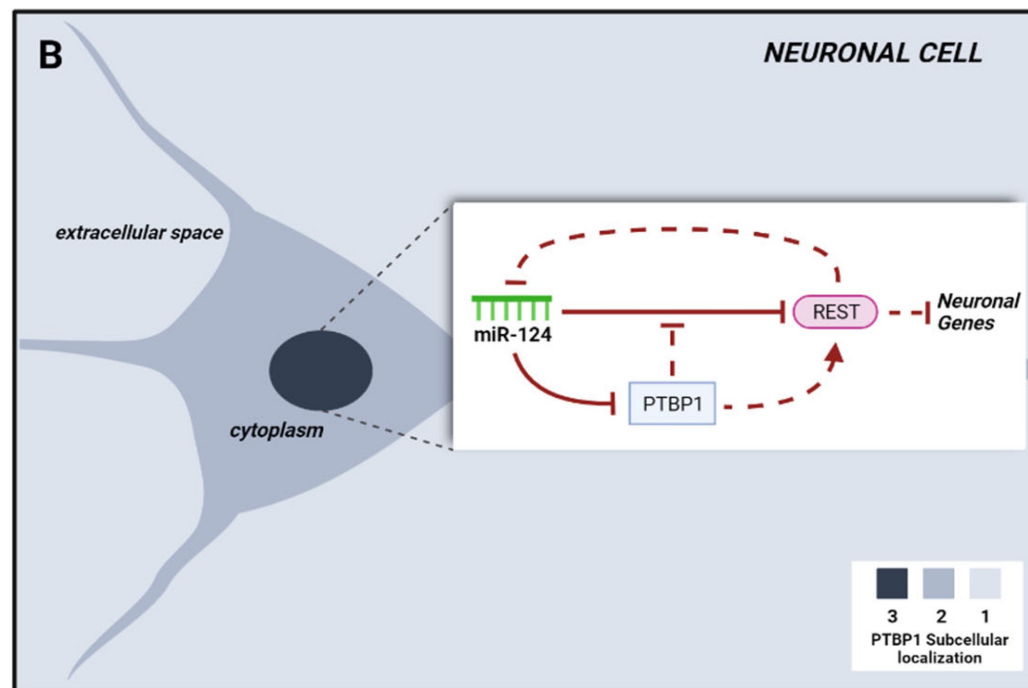
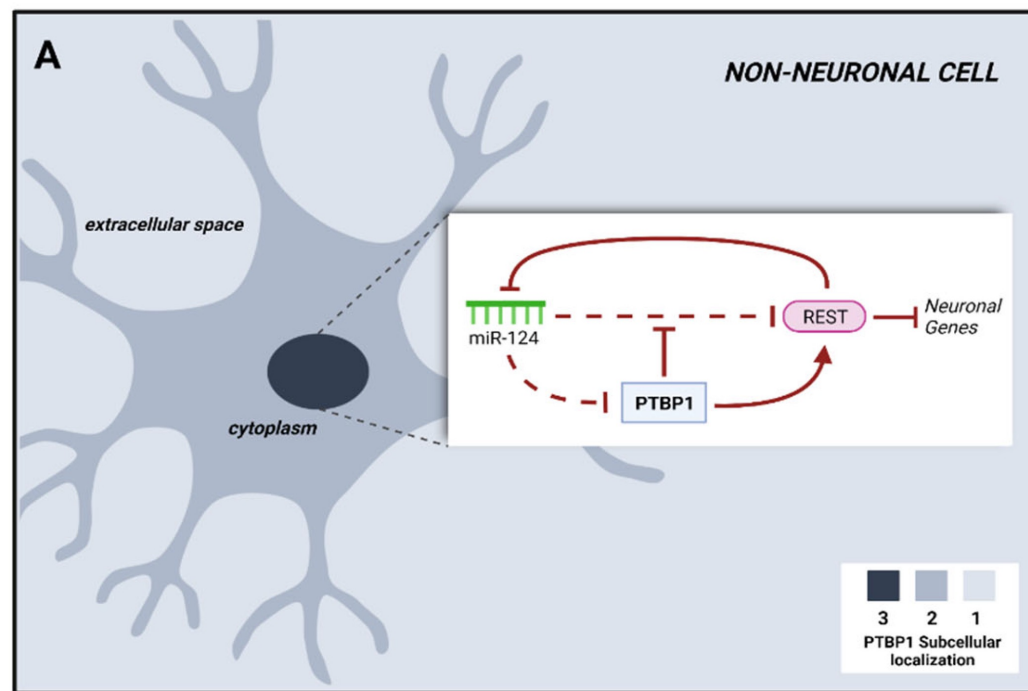
- What about neuron regeneration?

Restoration of vision after de novo genesis of rod photoreceptors in mammalian retinas

Kai Yao¹, Suo Qiu^{1,2}, Yanbin V. Wang^{3,4}, Silvia J. H. Park³, Ethan J. Mohns⁵, Bhupesh Mehta^{4,6}, Xinran Liu⁷, Bo Chang⁸, David Zenisek^{3,4}, Michael C. Crair^{3,5}, Jonathan B. Demb^{3,4} & Bo Chen^{1,9,10*}

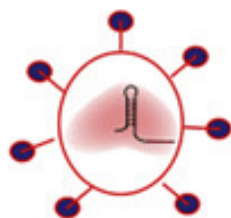


ShH10-GFAP-mediated gene transfer of Otx2, Crx, and Nrl

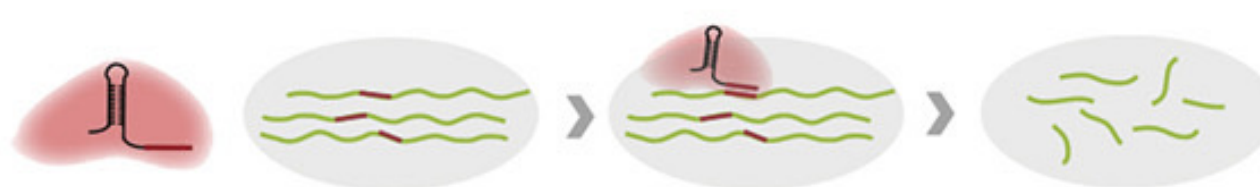


Glia-to-Neuron Conversion by CRISPR-CasRx Alleviates Symptoms of Neurological Disease in Mice

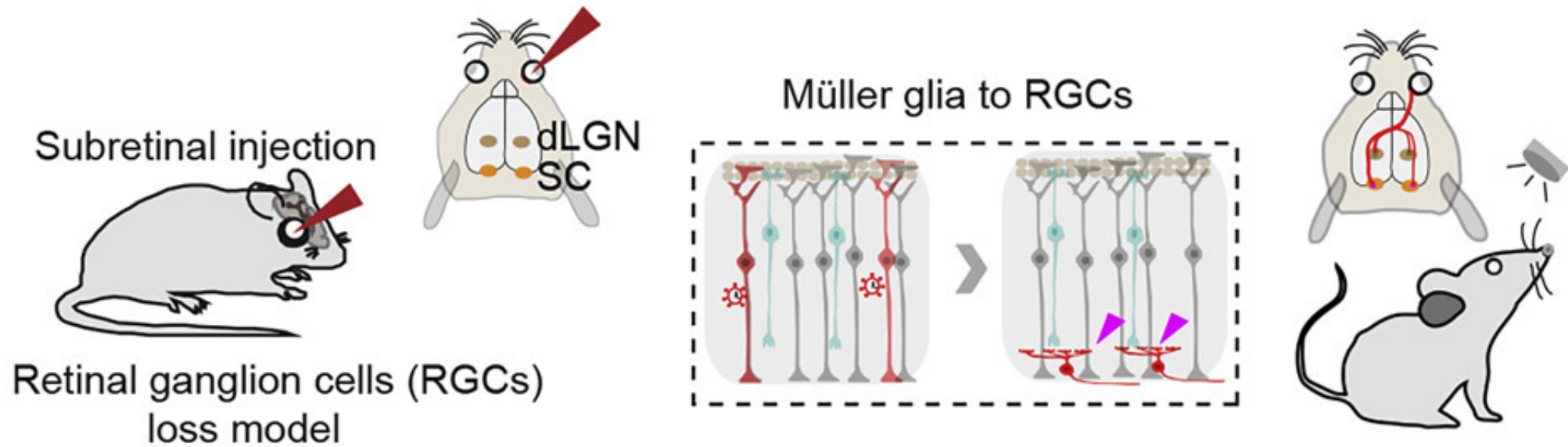
AAV-GFAP-CasRx-gRNA



Knockdown of *Ptbp1* mRNA

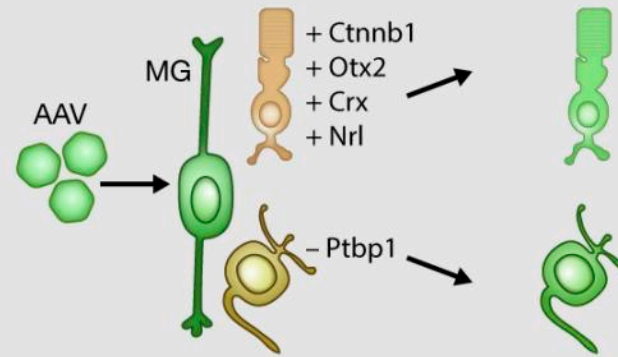


Ptbp1 downregulation converts MG into RGCs by direct transdifferentiation



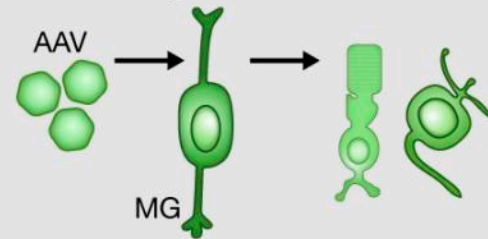
Cell targeting vs. cell generation

Observation



Interpretation

Only MG infected
MG generates neurons

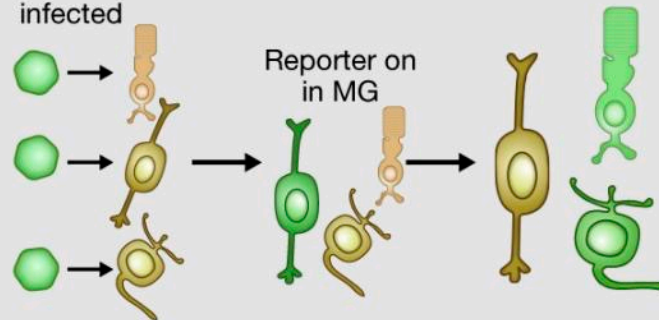


Alternative interpretation

MG and neurons infected

Reporter on in MG

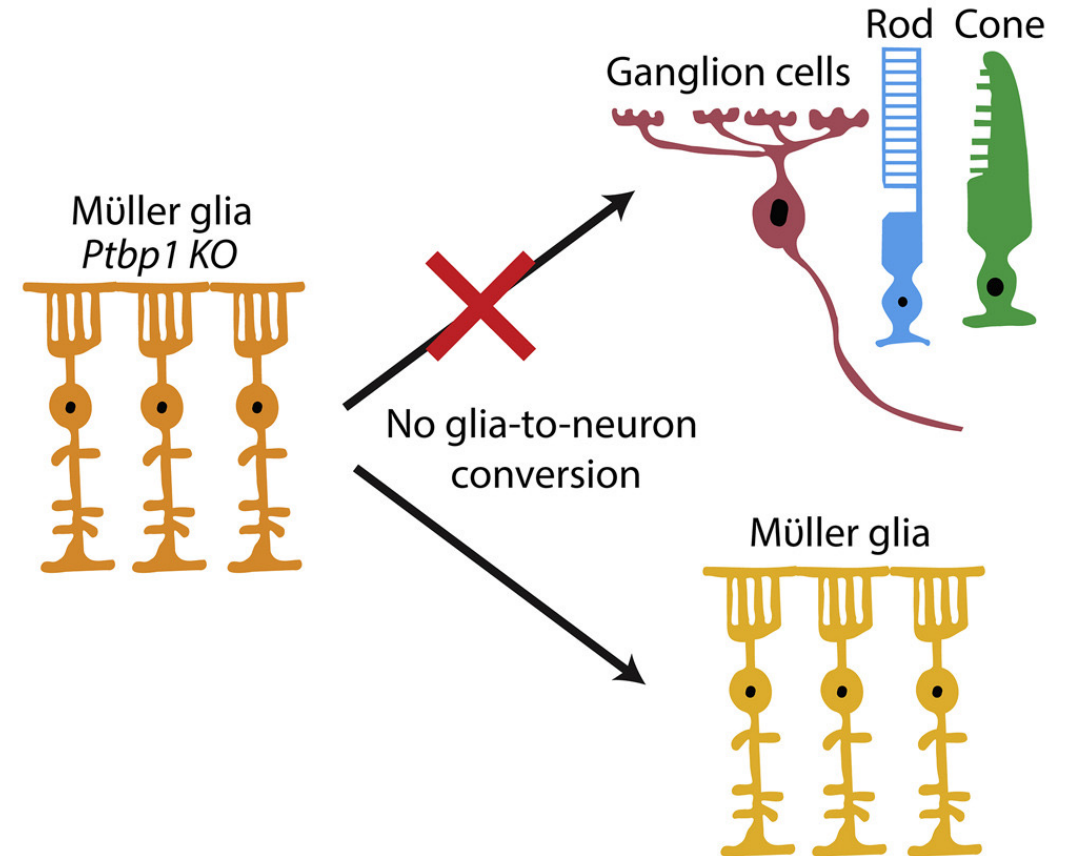
Reporter on in neurons



Thanh Hoang, Dong Won Kim,
Haley Appel, ..., Minzhong Yu,
Neal S. Peachey, Seth Blackshaw

Genetic loss of function of *Ptbp1* does not induce glia-to-neuron conversion in retina

- *Ptbp1* is genetically disrupted selectively in adult mouse Müller glia
- The fate of cells lacking *Ptbp1* is analyzed with lineage tracing and molecular markers
- *Ptbp1* deletion does not lead to glia-to-neuron conversion in retina
- scRNA-seq shows that glial identity is maintained after *Ptbp1* deletion

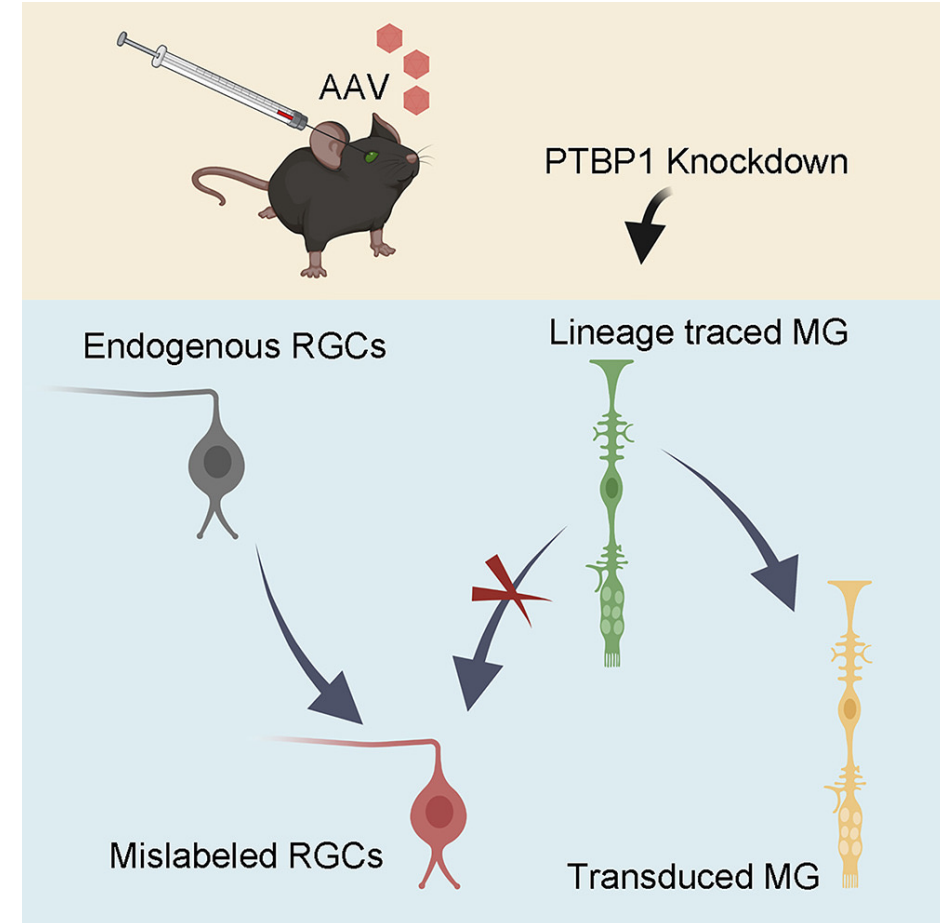


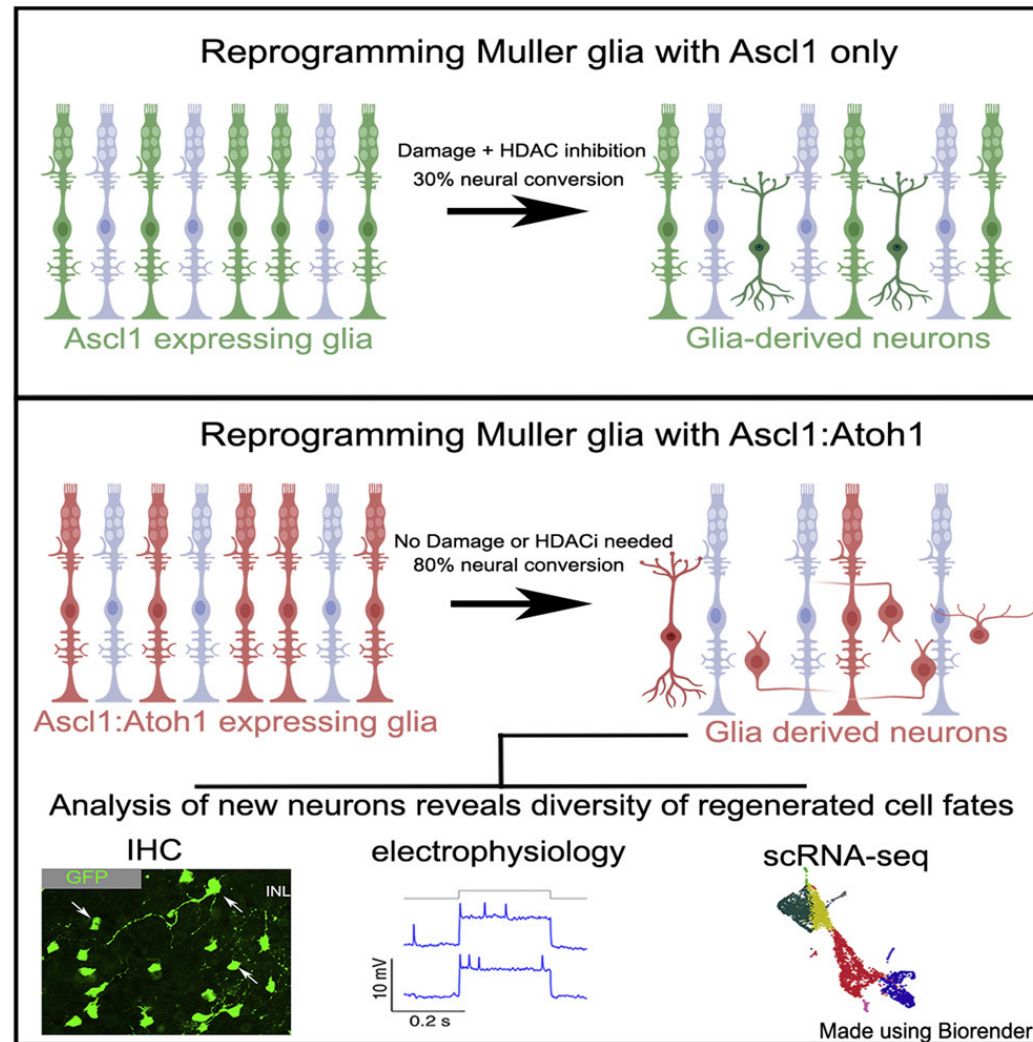
Critical examination of *Ptbp1*-mediated glia-to-neuron conversion in the mouse retina

Authors

Ye Xie, Jing Zhou, Bo Chen

- AAV-based Cre recombination is unsuitable for examining MG-to-RGC conversion
- Lineage-traced MG are not converted into RGCs after *Ptbp1* downregulation
- NMDA-induced injury does not facilitate MG-to-RGC conversion after *Ptbp1* downregulation
- Stringent fate mapping is required for critical examination of glia-to-neuron conversion

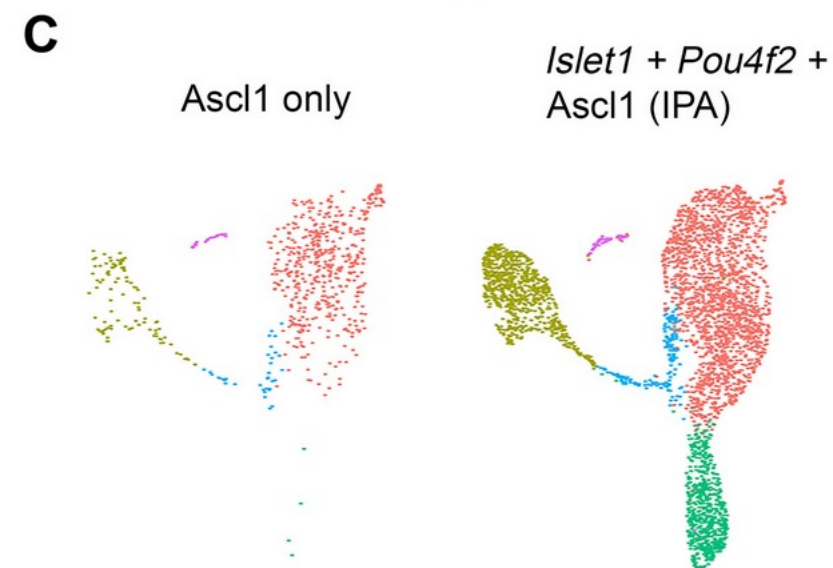
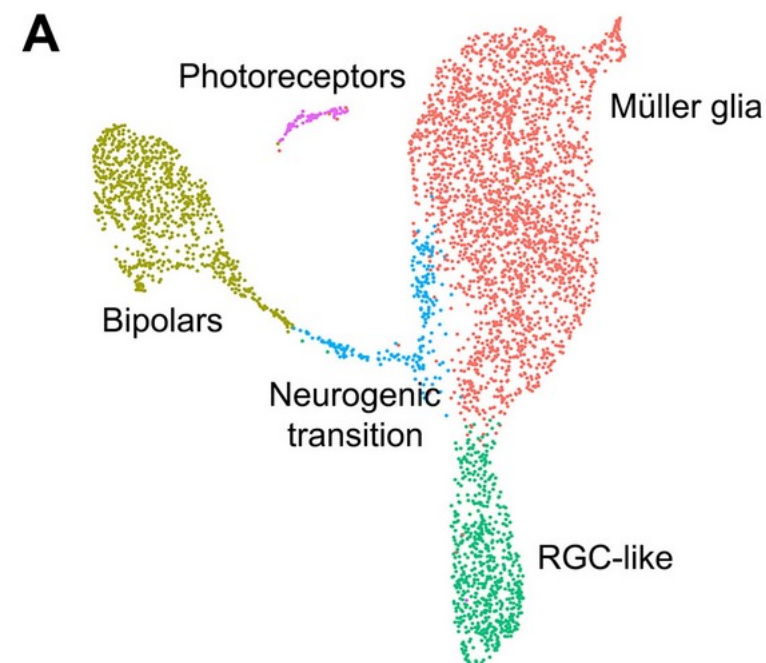




Reprogramming Müller glia to regenerate ganglion-like cells in adult mouse retina with developmental transcription factors

LEVI TODD ^{ID}, WESLEY JENKINS ^{ID}, CONNOR FINKBEINER, MARCUS J. HOOPER ^{ID}, PHOEBE C. DONALDSON, MARINA PAVLOU ^{ID}, JULIETTE WOHLSCHEGEL ^{ID},
NORIANNE INGRAM ^{ID}, FRED RIEKE, AND THOMAS A. REH ^{ID} [Authors Info & Affiliations](#)

SCIENCE ADVANCES • 23 Nov 2022 • Vol 8, Issue 47 • DOI: 10.1126/sciadv.abq7219

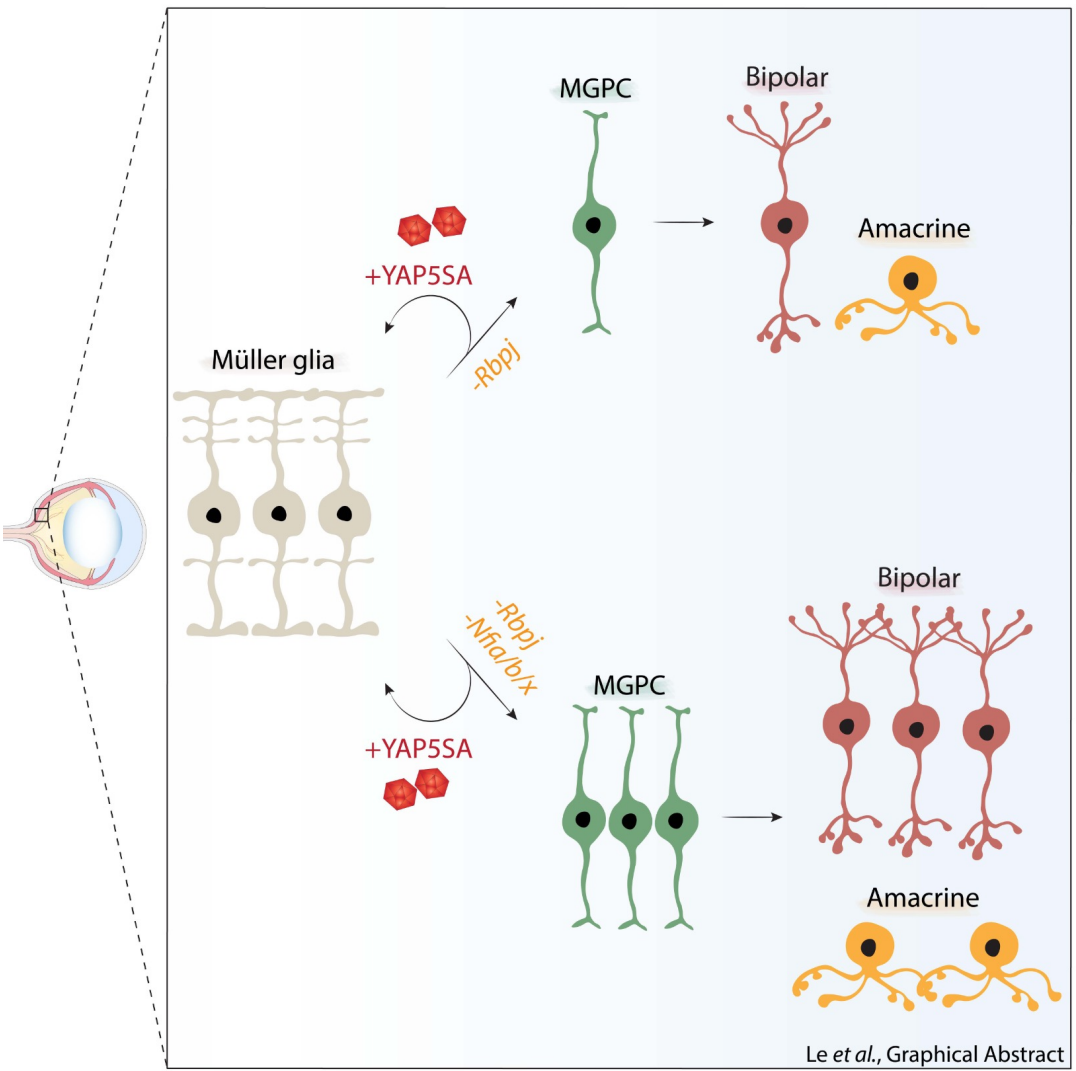
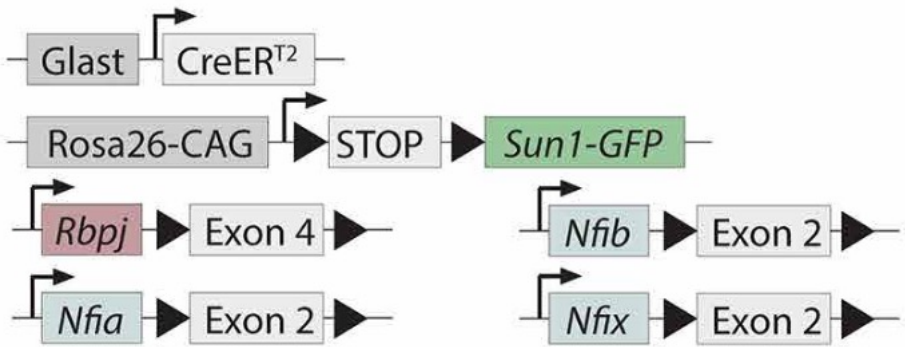


REGENERATION

Robust reprogramming of glia into neurons by inhibition of Notch signaling and nuclear factor I (NFI) factors in adult mammalian retina

Nguyet Le¹, Trieu-Duc Vu^{2,3}, Isabella Palazzo¹, Ritvik Pulya¹, Yehna Kim¹, Seth Blackshaw^{1,4,5,6,7*}, Thanh Hoang^{2,3,8*}

July 2024



AAV-mediated expression of proneural factors stimulates neurogenesis from adult Müller glia *in vivo*

 Marina Pavlou, Marlene Probst, Elizaveta Filippova,  Lew Kaplan, Aric R. Prieve,
 Fred Rieke,  Thomas A. Reh

bioRxiv
THE PREPRINT SERVER FOR BIOLOGY

Posted September 15, 2024

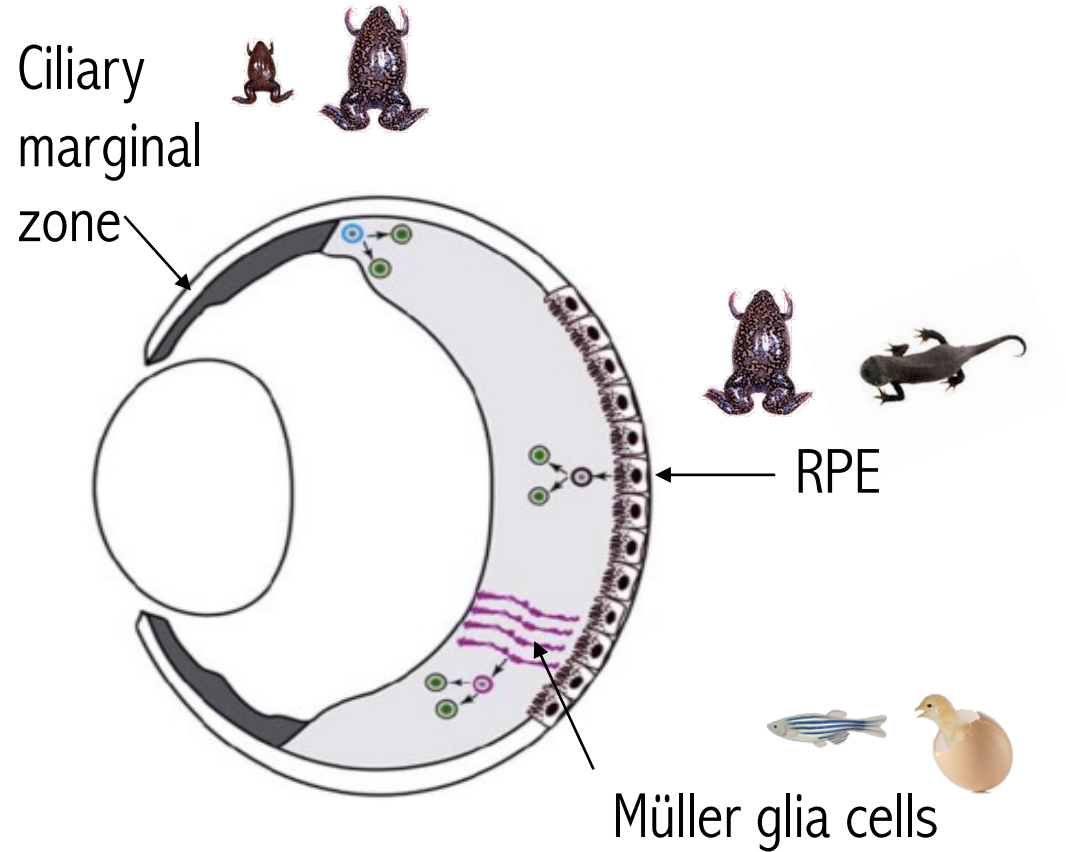
Viral-mediated Oct4 overexpression and inhibition of Notch signaling synergistically induce neurogenic competence in mammalian Muller glia.

Nguyet Le, Sherine Awad, Isabella Palazzo, Thanh Hoang,  Seth Blackshaw

bioRxiv
THE PREPRINT SERVER FOR BIOLOGY

Posted September 19, 2024

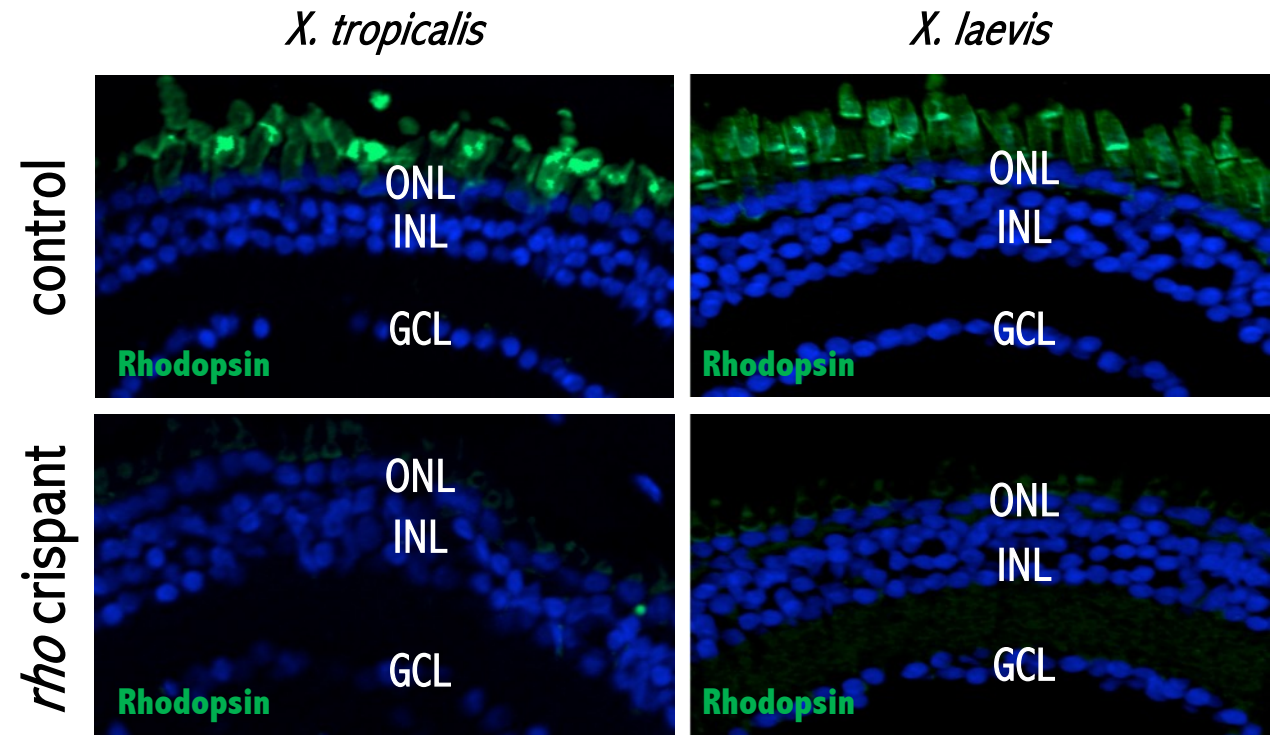
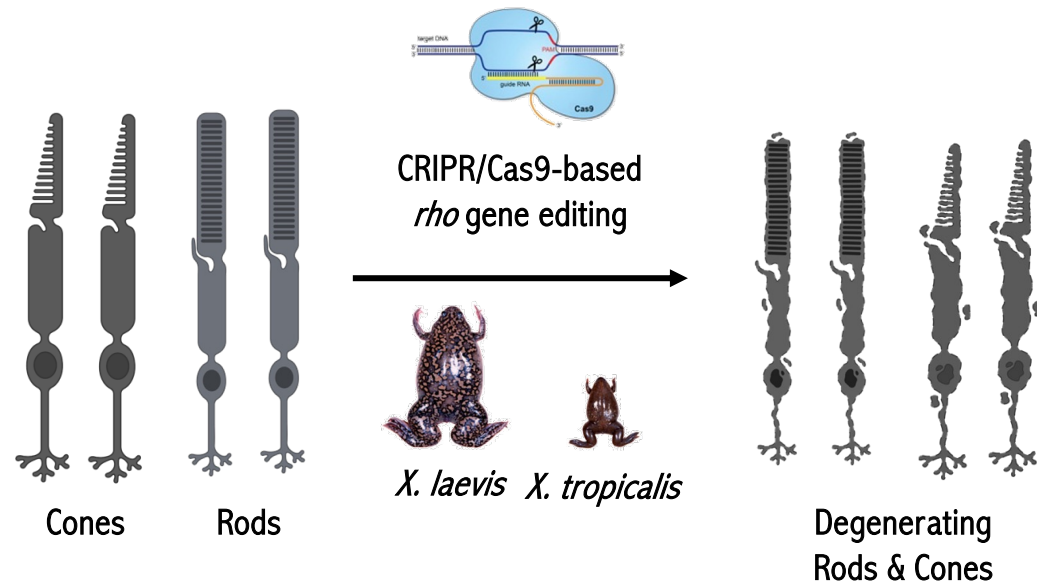
key step forward in developing a cellular reprogramming approach to regenerative medicine



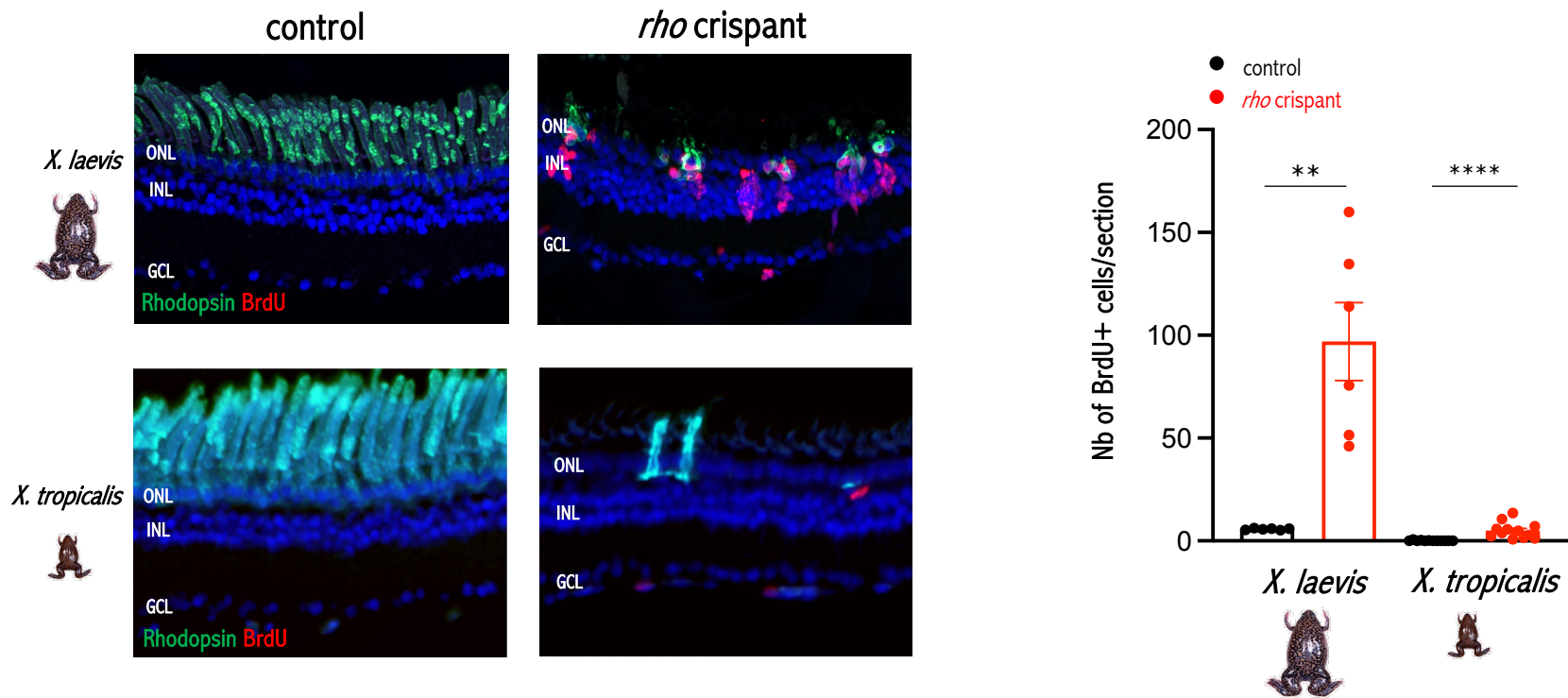
Retinectomy $\begin{cases} \rightarrow X. laevis \rightarrow \text{RPE} + \text{CMZ} \\ \rightarrow X. tropicalis \rightarrow \text{CMZ} \end{cases}$

Do Müller cells from *X. laevis* and *X. tropicalis* respond similarly to retinal injury?

CRISPR-dependent photoreceptor degeneration as a model of retinitis pigmentosa

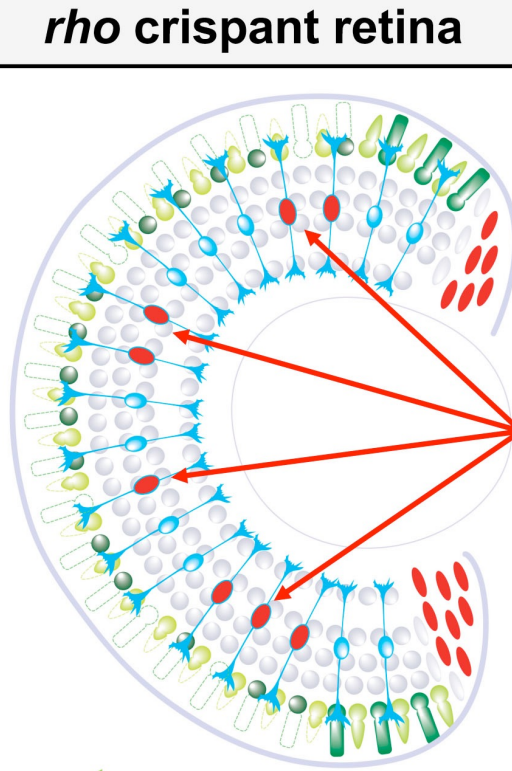
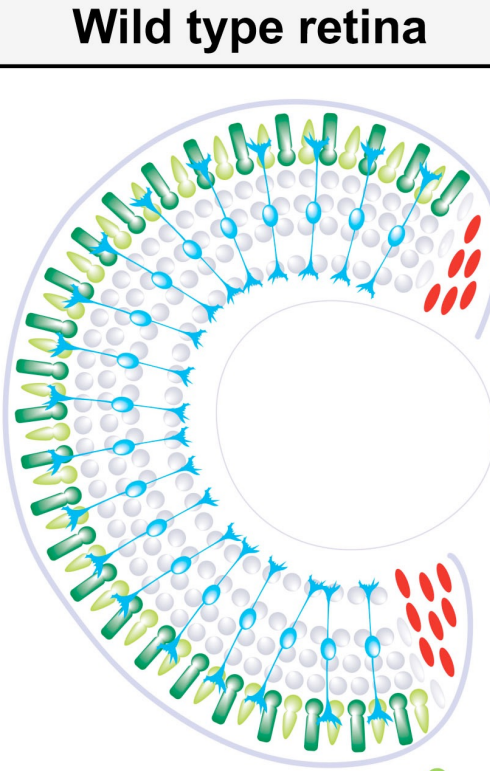


Müller cell response to photoreceptor degeneration in *X. laevis* and *X. tropicalis*



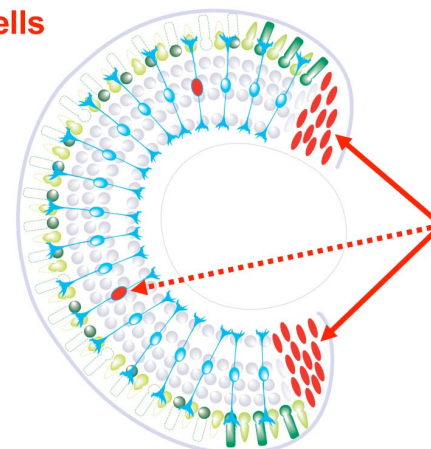
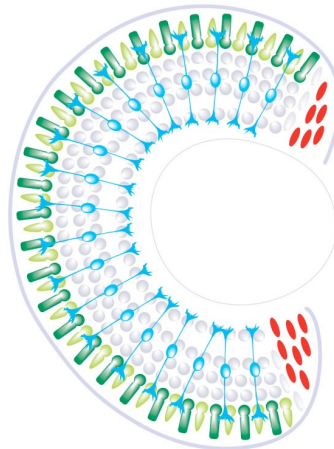


Xenopus laevis



Cone photoreceptors
Rod photoreceptors
Müller glial cells
Proliferative cells

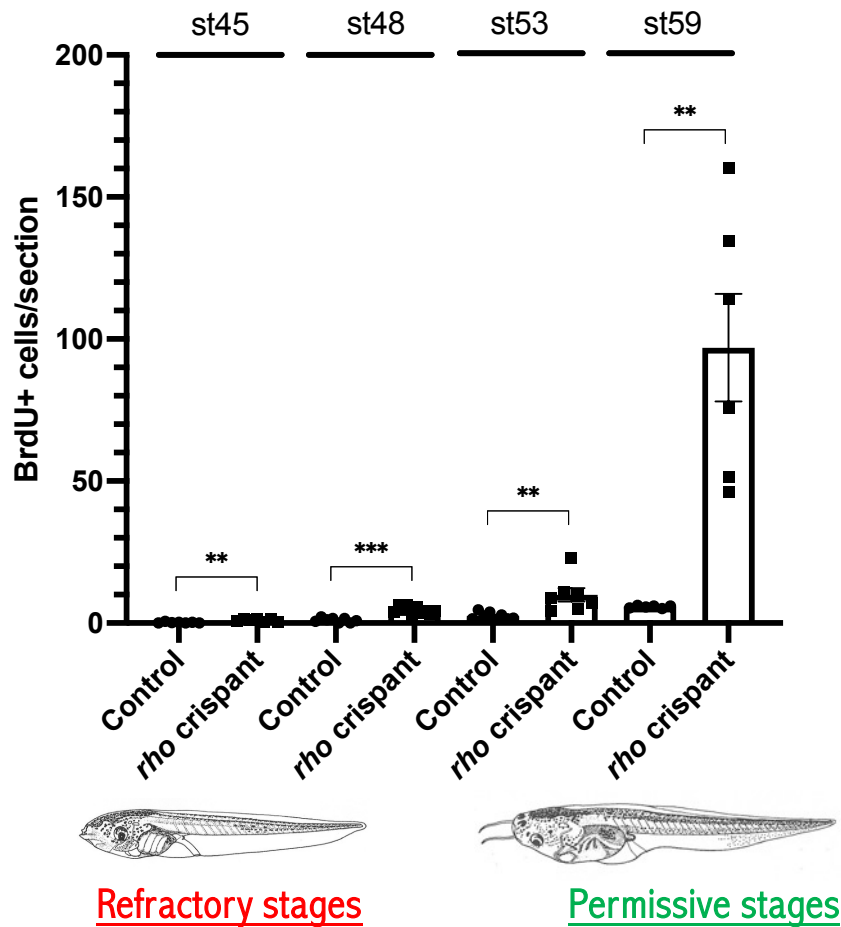
Xenopus tropicalis

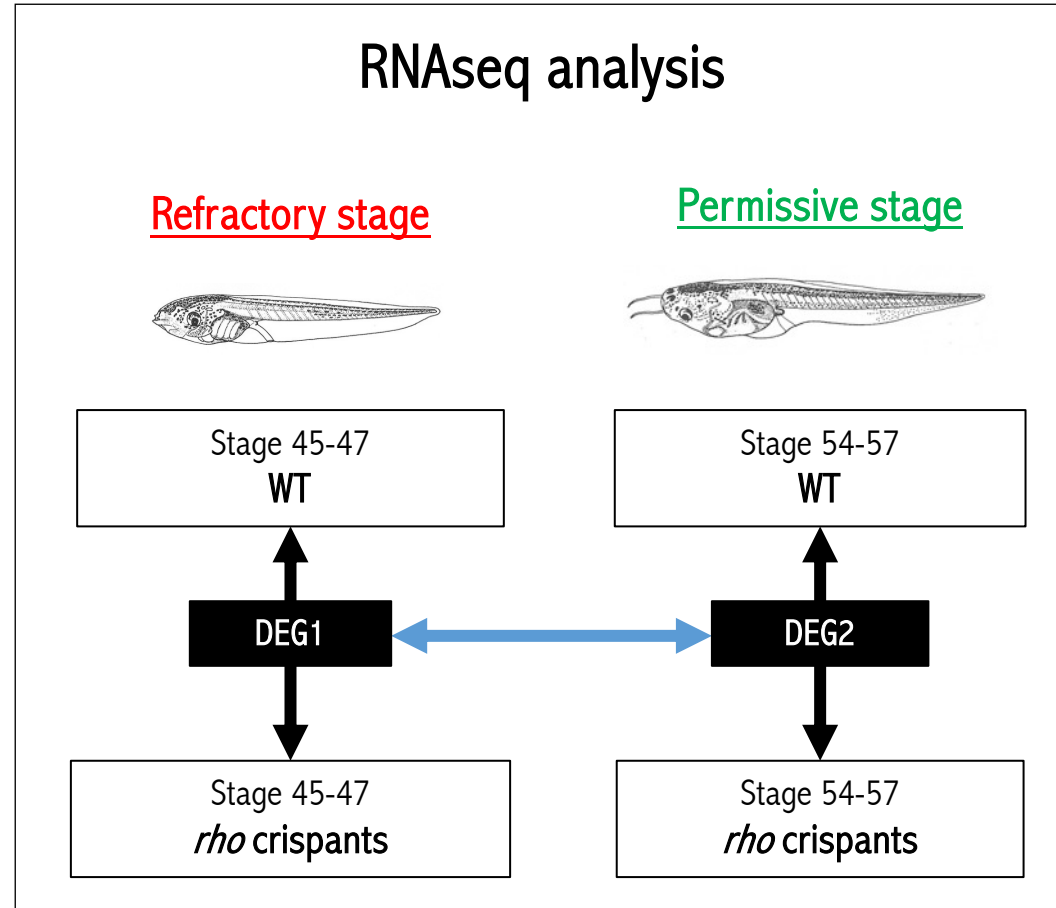


Müller cell response also differs at different stages in *X. laevis*



X. laevis

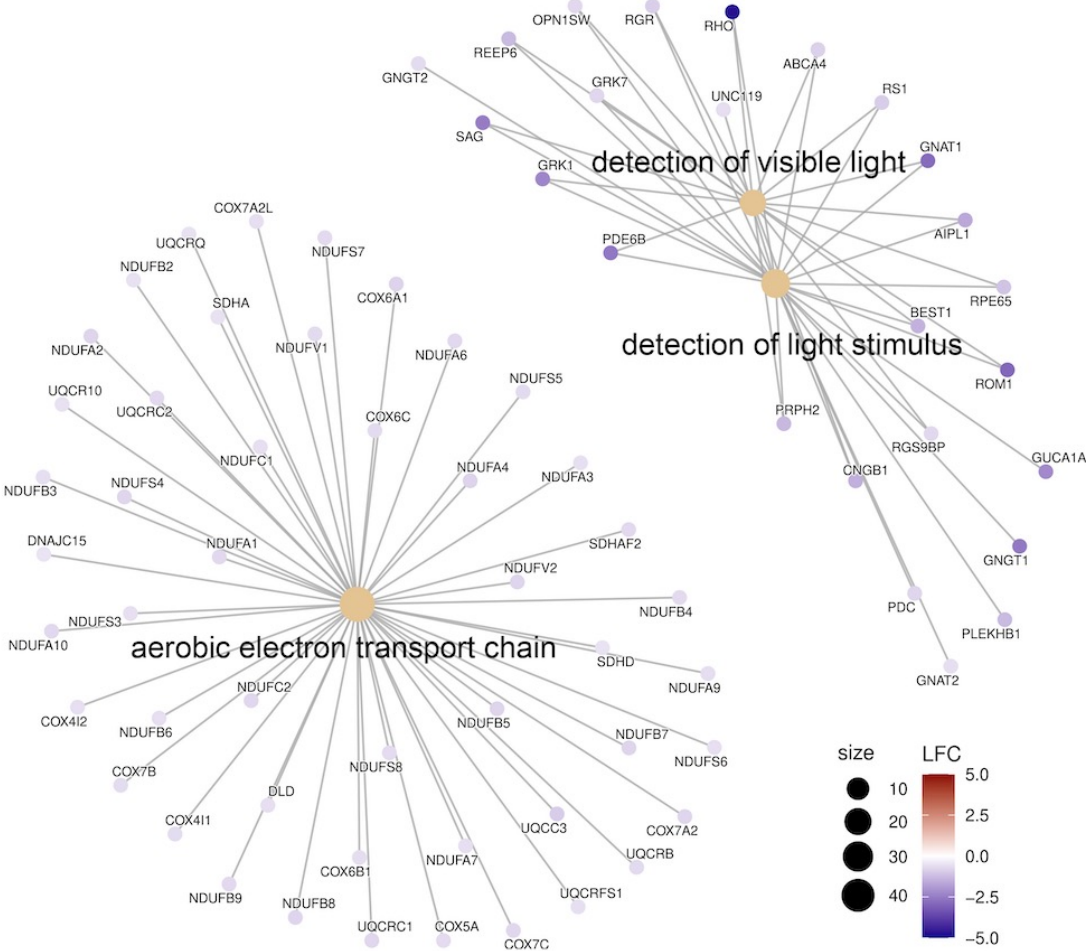




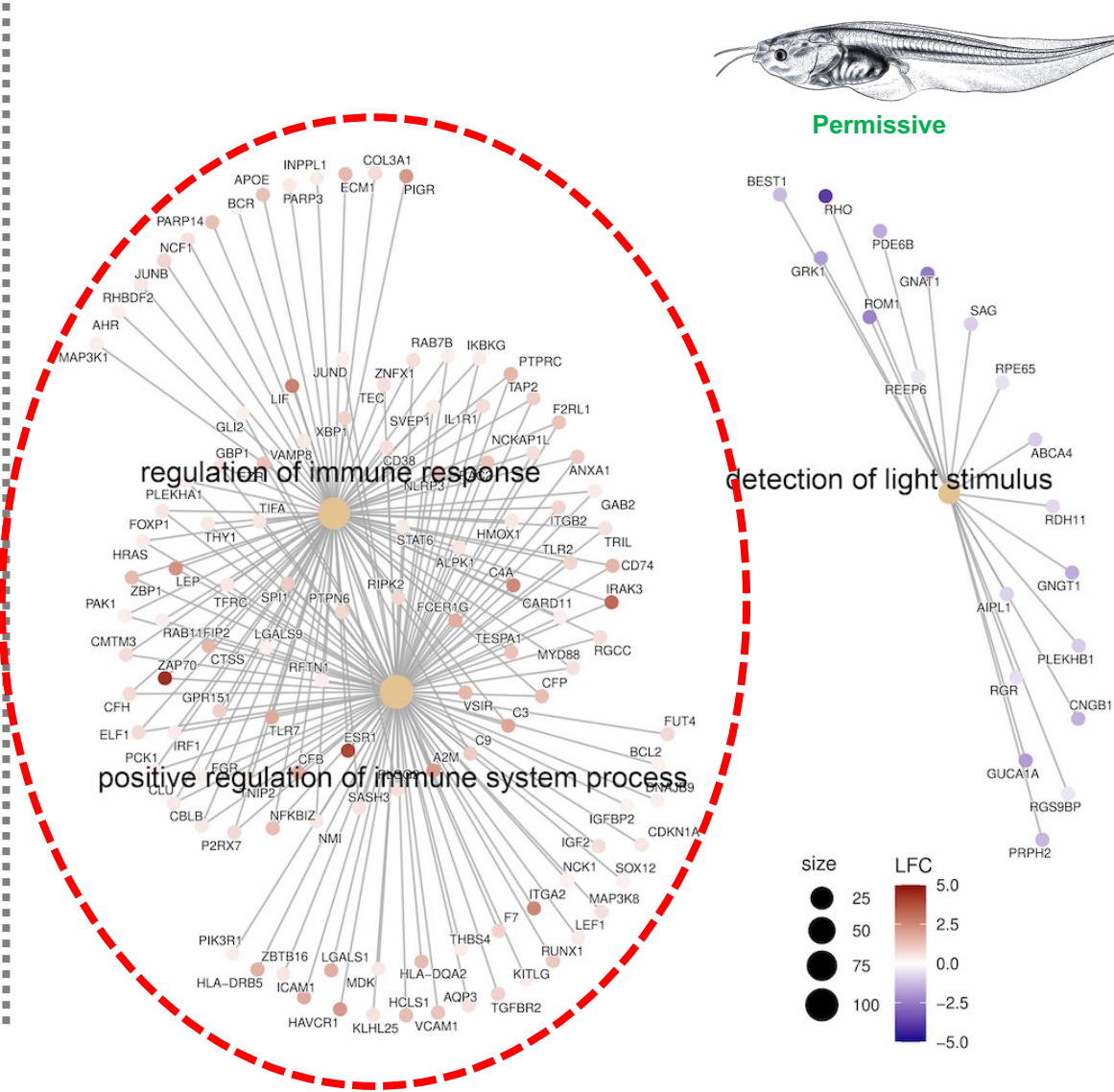
Immune response is upregulated only at permissive stages following injury



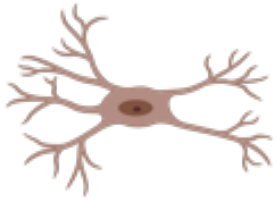
Refractory



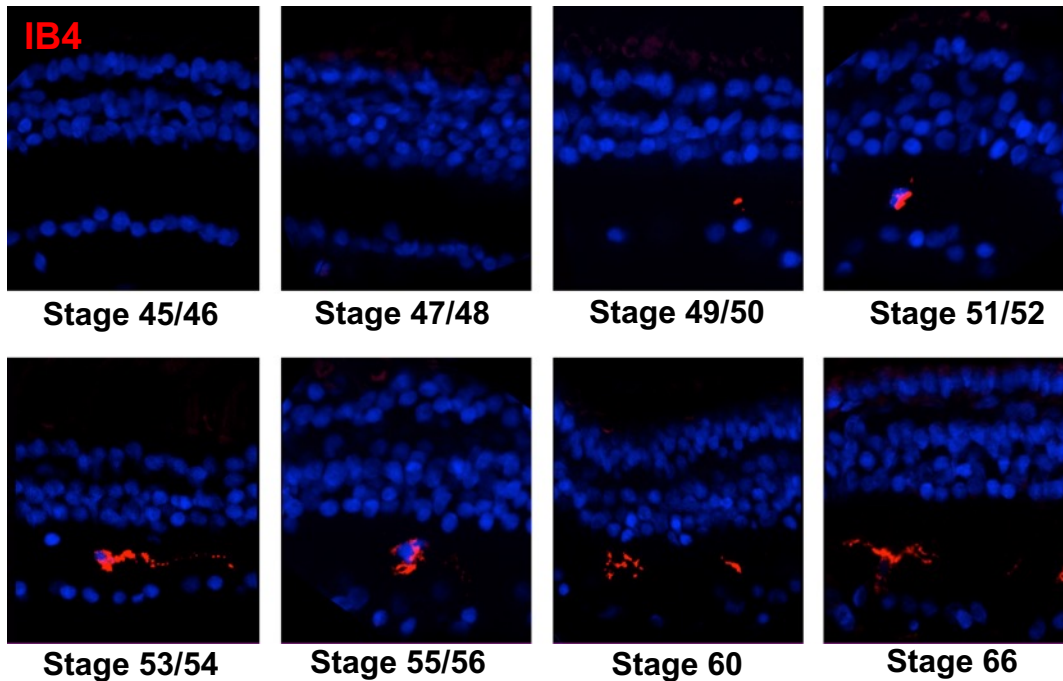
Permissive



Evolution of microglia at different stages in physiological conditions



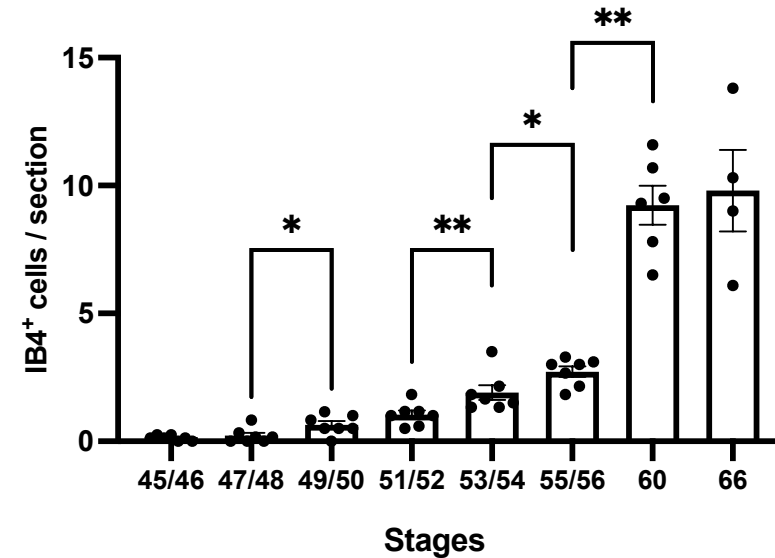
Microglia



Refractory stages



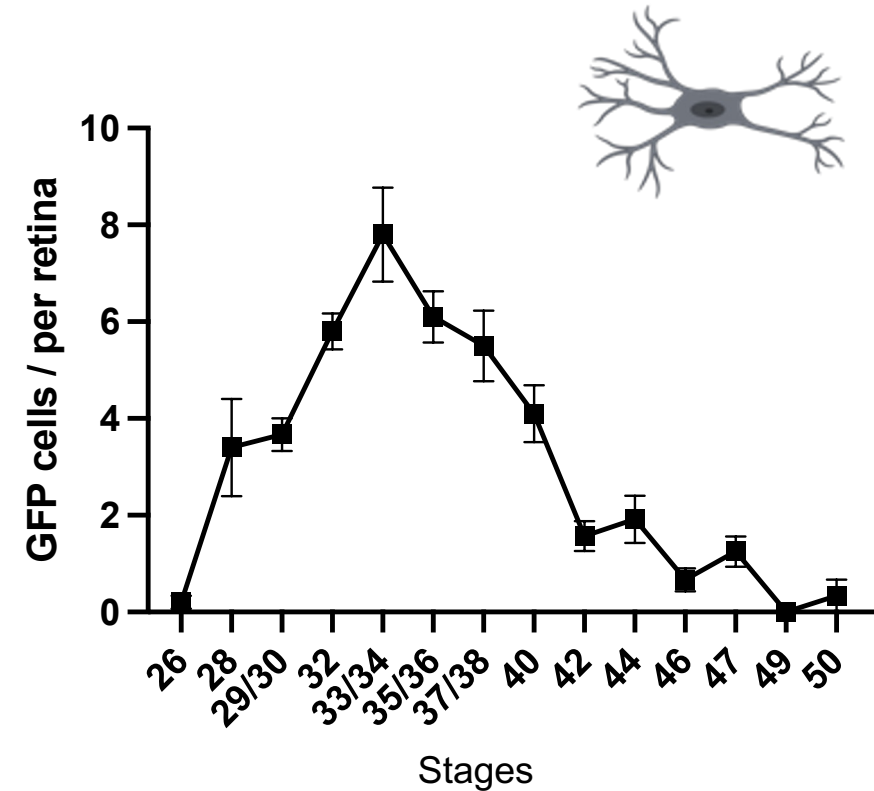
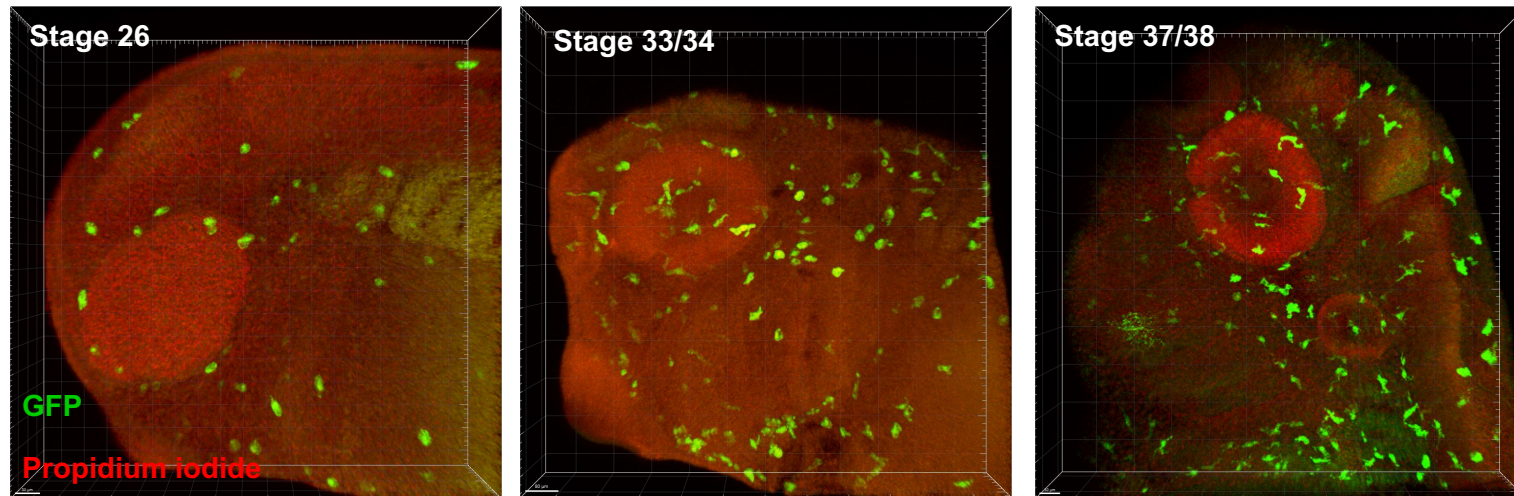
Permissive stages



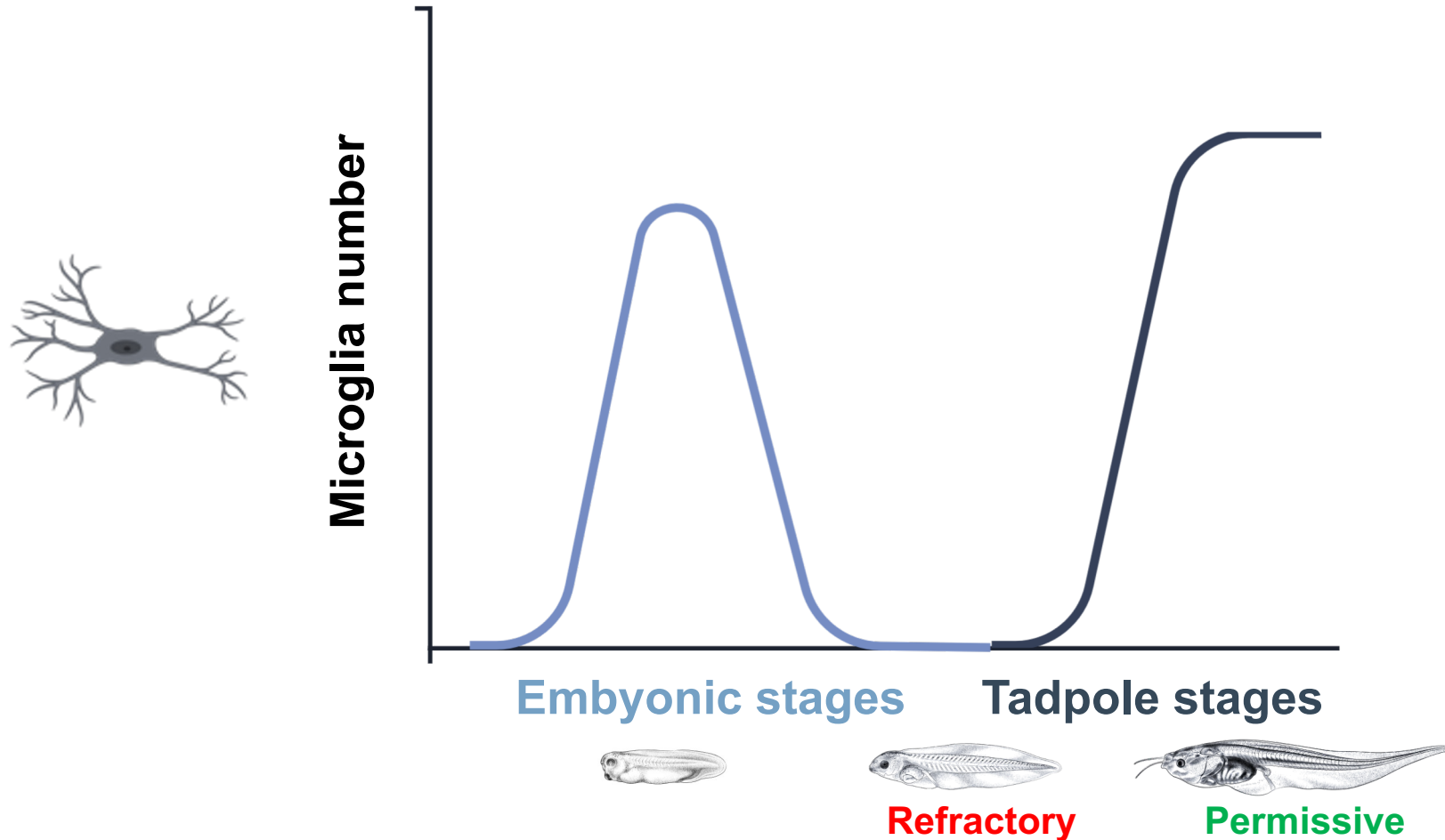
Retinal infiltration with microglia coincides with a shift in the proliferative capacity of Müller cells

Dynamic retinal colonization of microglia during development

lurp1:GFP (from Jacques Robert, University of Rochester)



Two waves of microglia colonization in the *Xenopus* retina



Are Müller cells refractory because of a limited inflammatory microenvironment?

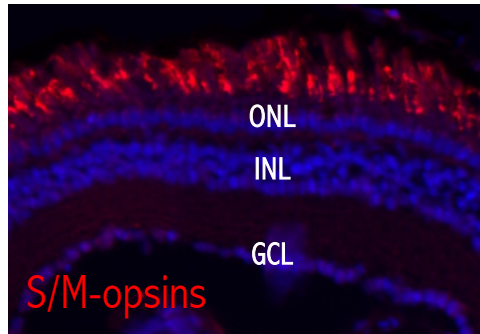
Can we trigger Müller cell proliferative response at the refractory stage by generating neuroinflammation?

CoCl₂ : a novel model to induce retinal degeneration in *Xenopus*

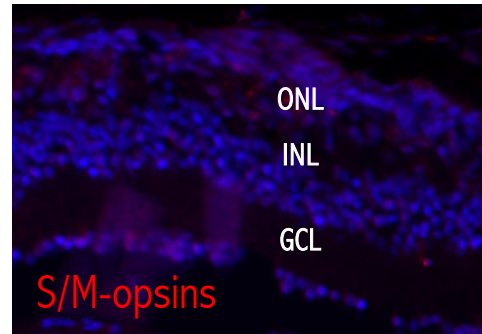
CoCl₂



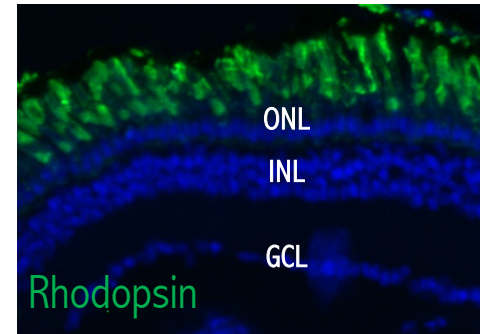
Control



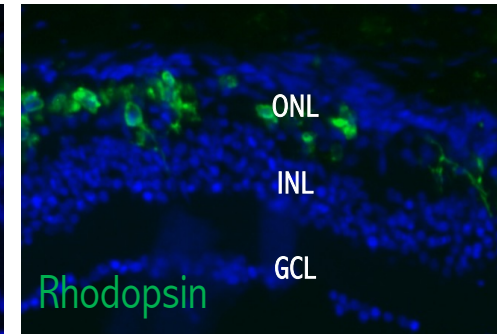
CoCl₂



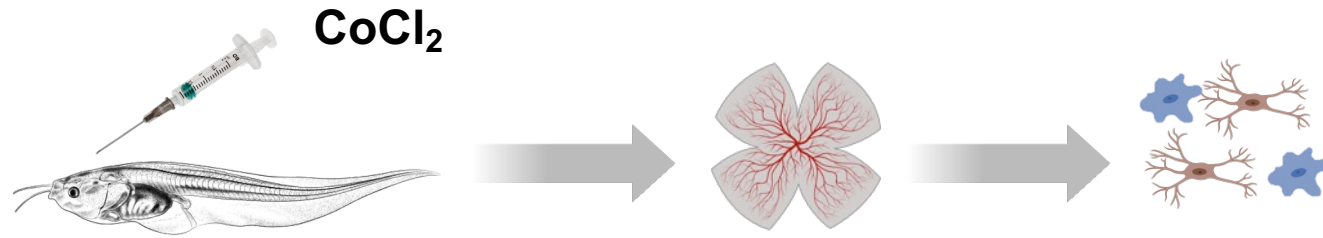
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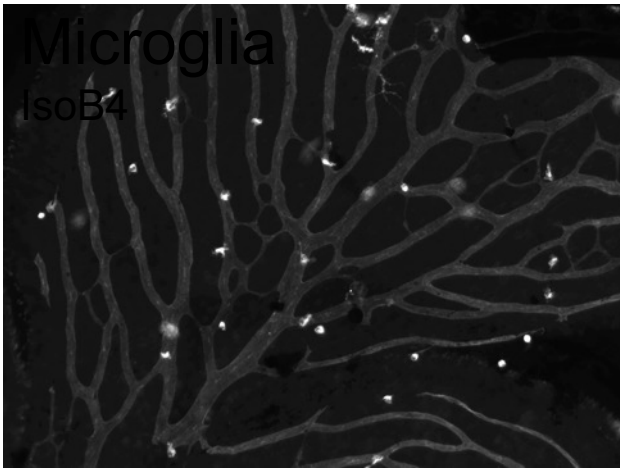
CoCl₂



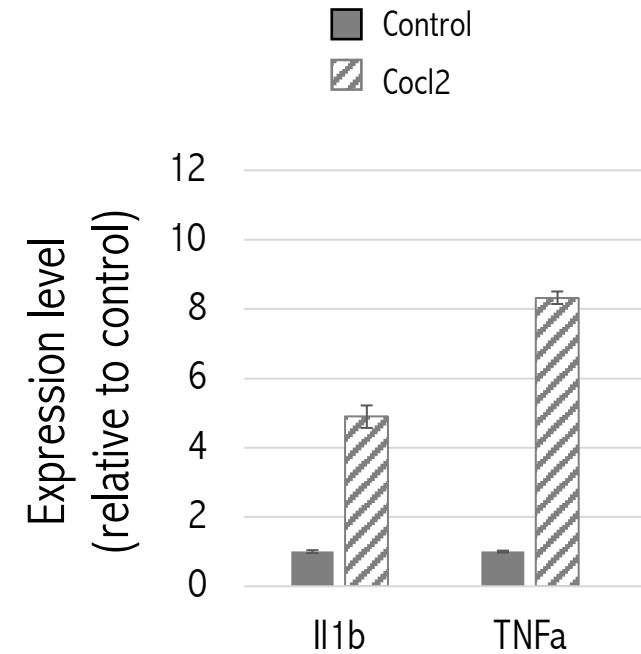
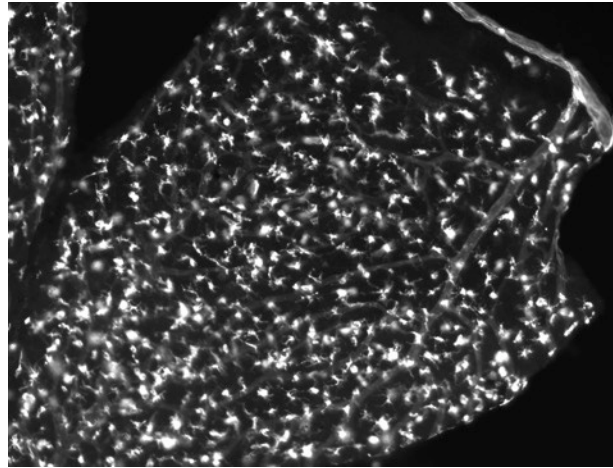
CoCl₂ intraocular injections leads to a severe inflammatory response



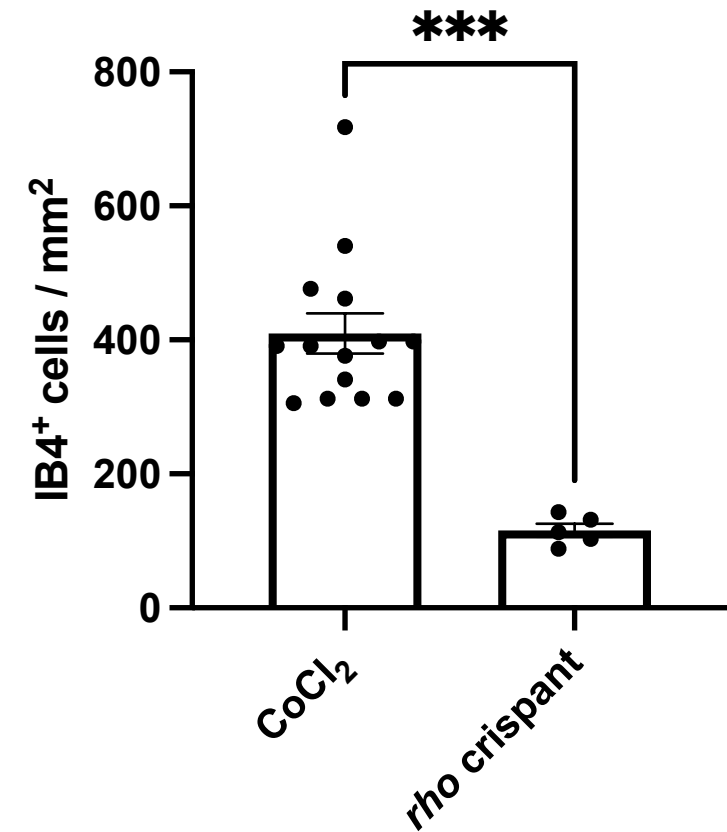
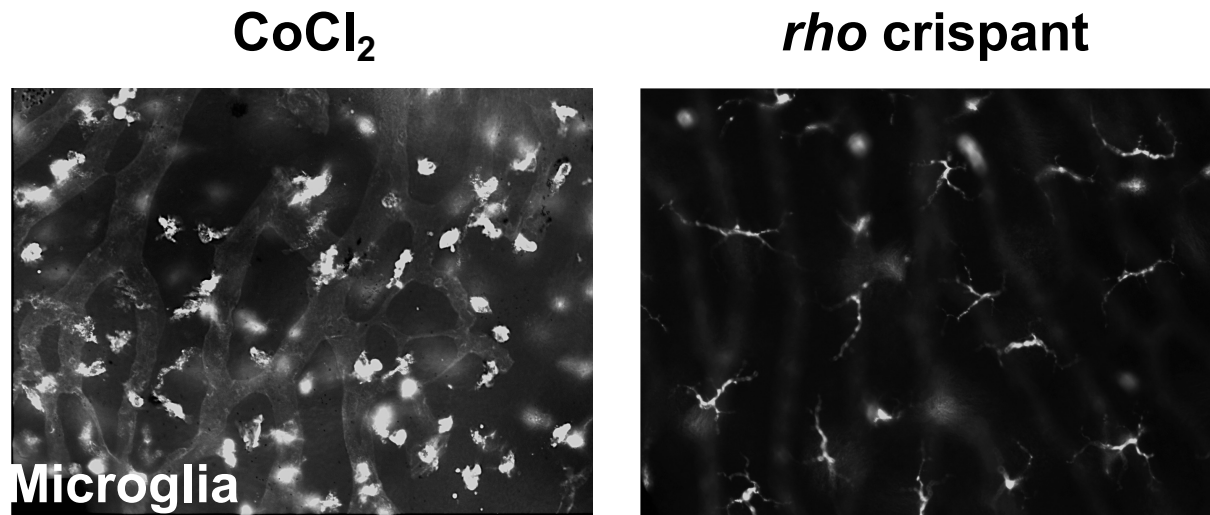
Control



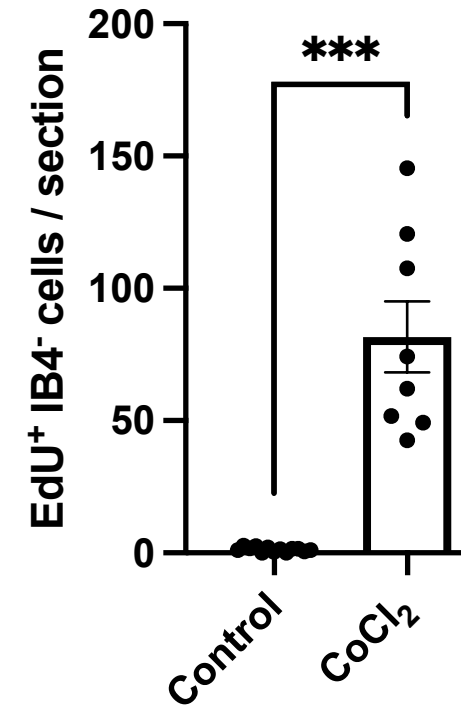
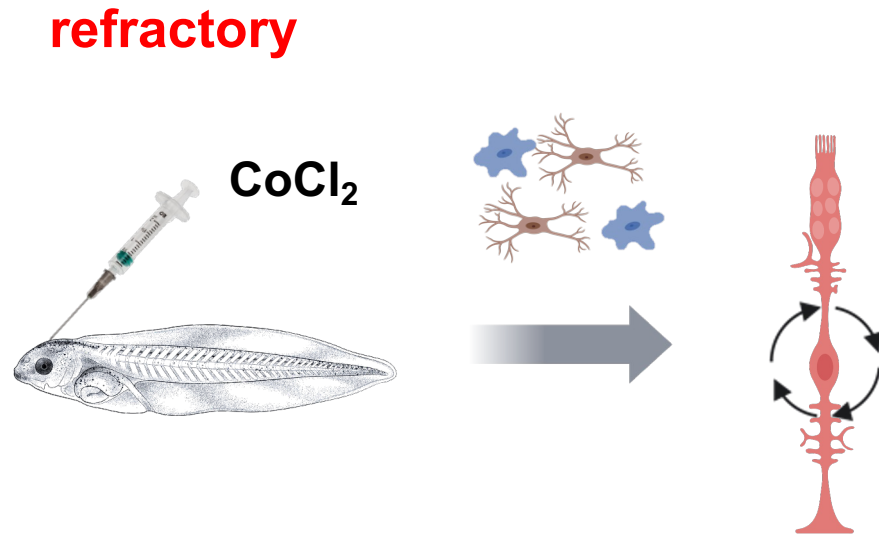
CoCl₂



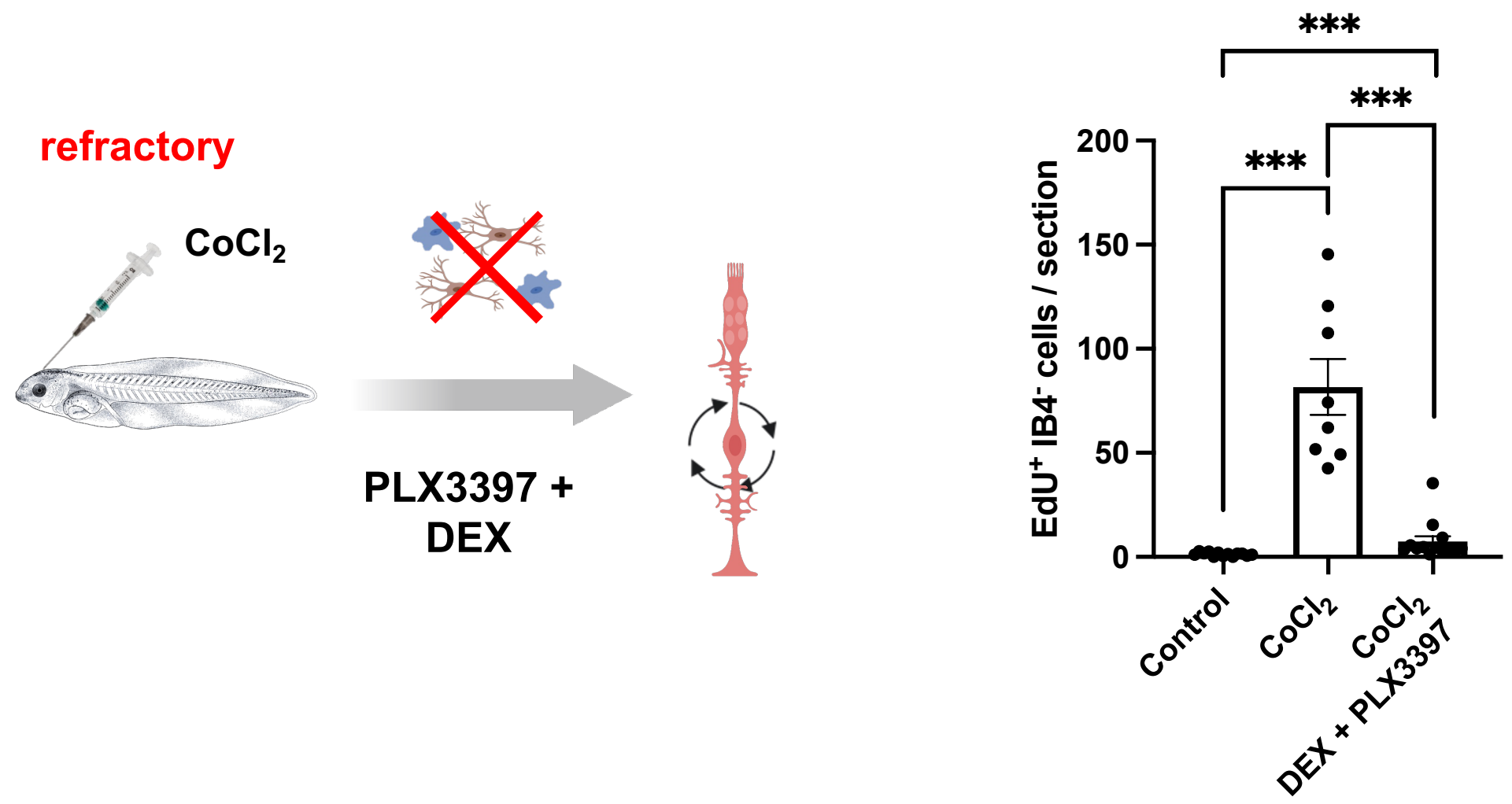
The increase in microglia is higher in CoCl_2 tadpoles than in *rho* crispant

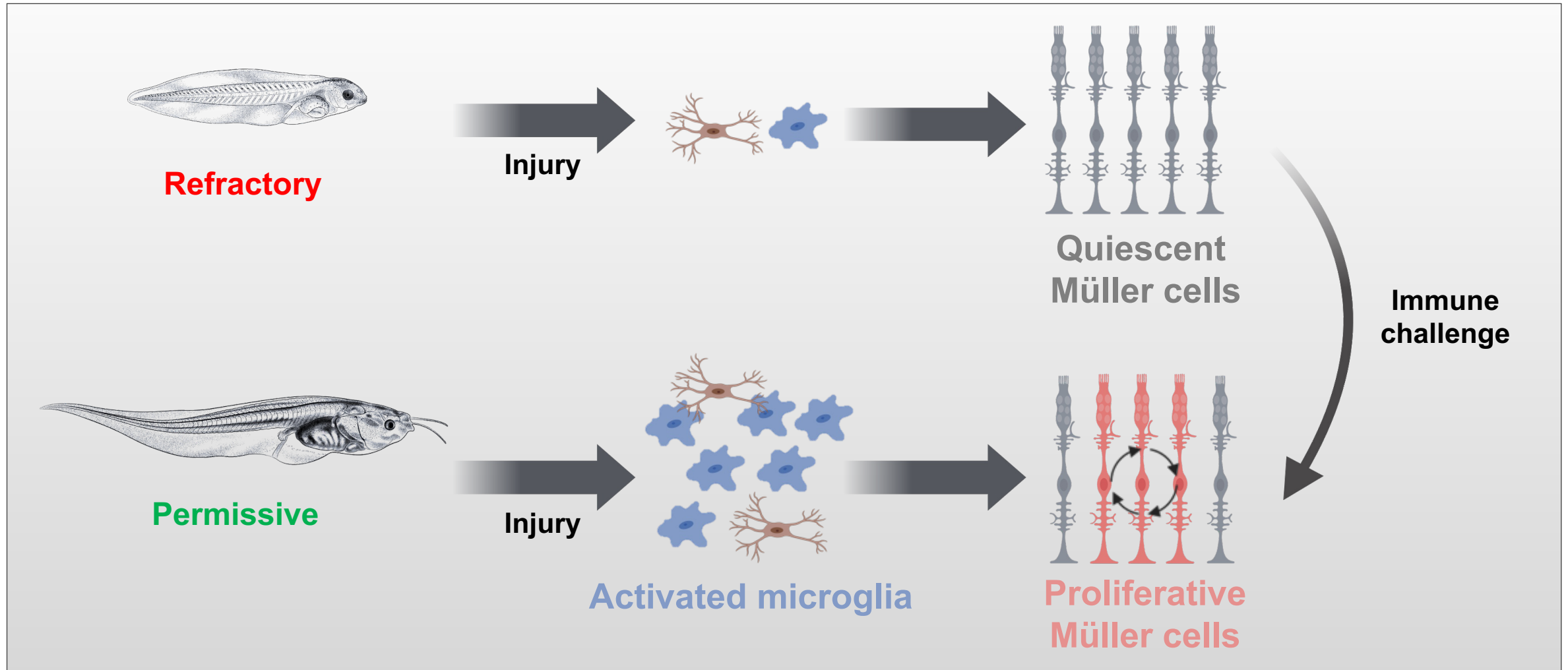


Microglia activation triggers a proliferation response of refractory Müller cells



Microglia activation mediates CoCl_2 dependent proliferative response of refractory Müller cells





Can an immune challenge awake refractory Müller cells
in other species?

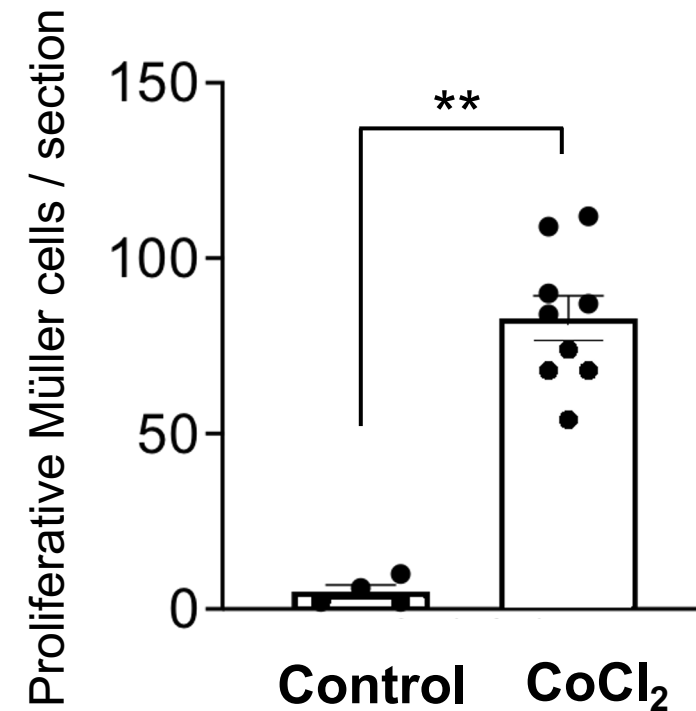
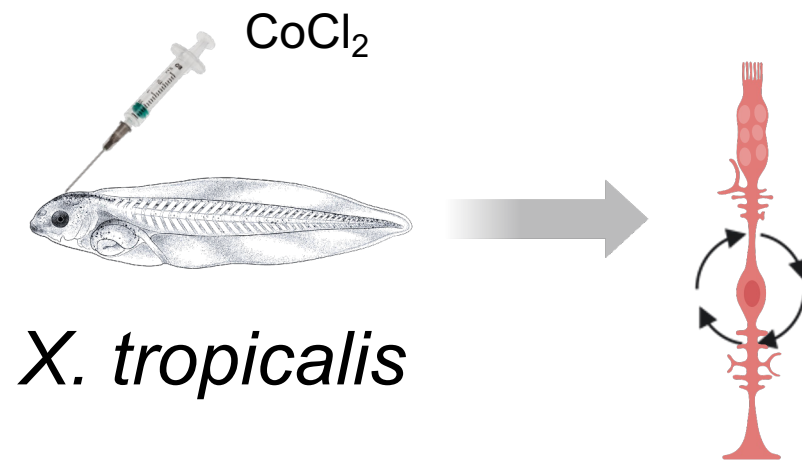
X. tropicalis



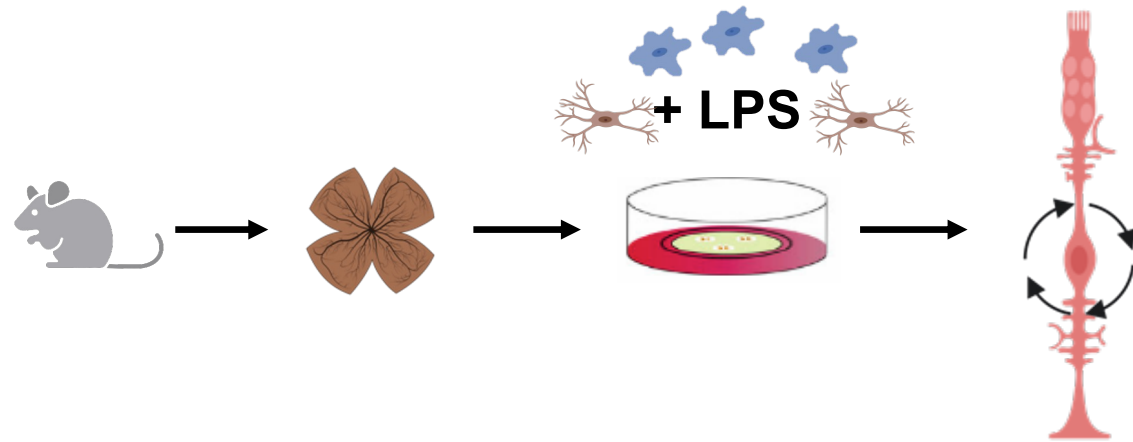
Mouse



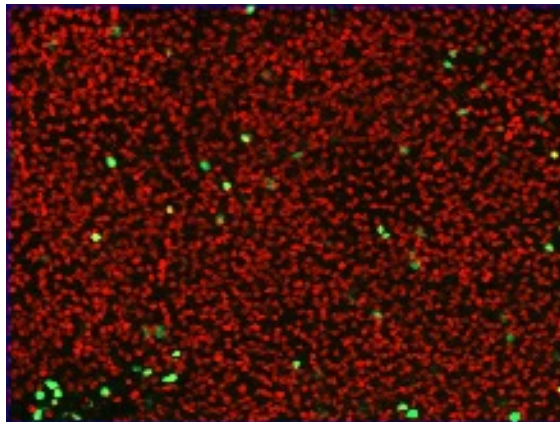
CoCl₂ triggers a proliferation response of *X. tropicalis* Müller cells



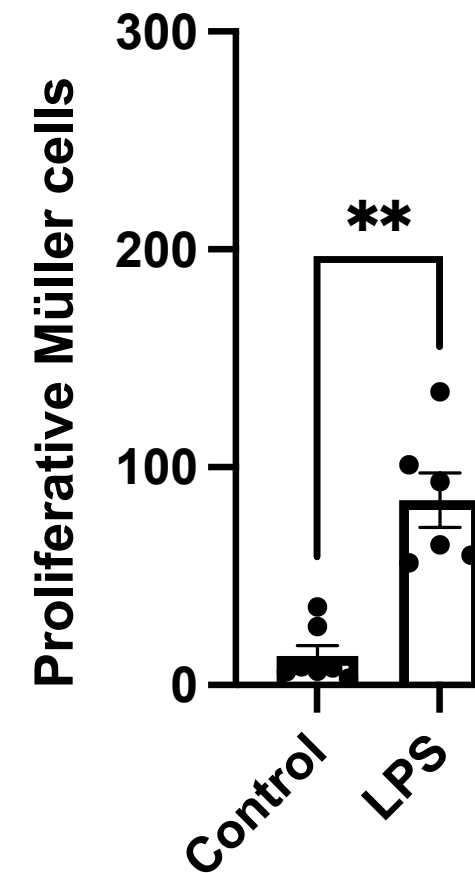
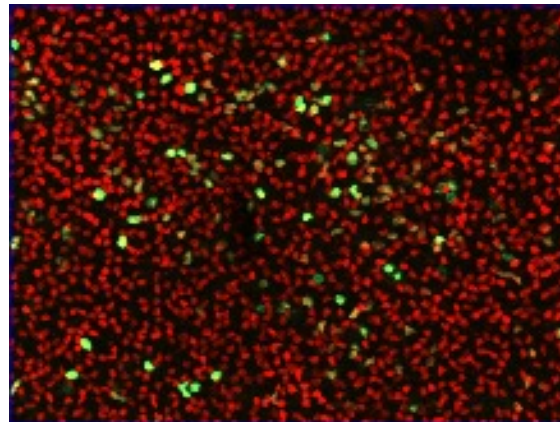
An immune challenge promotes mouse Müller cell proliferation



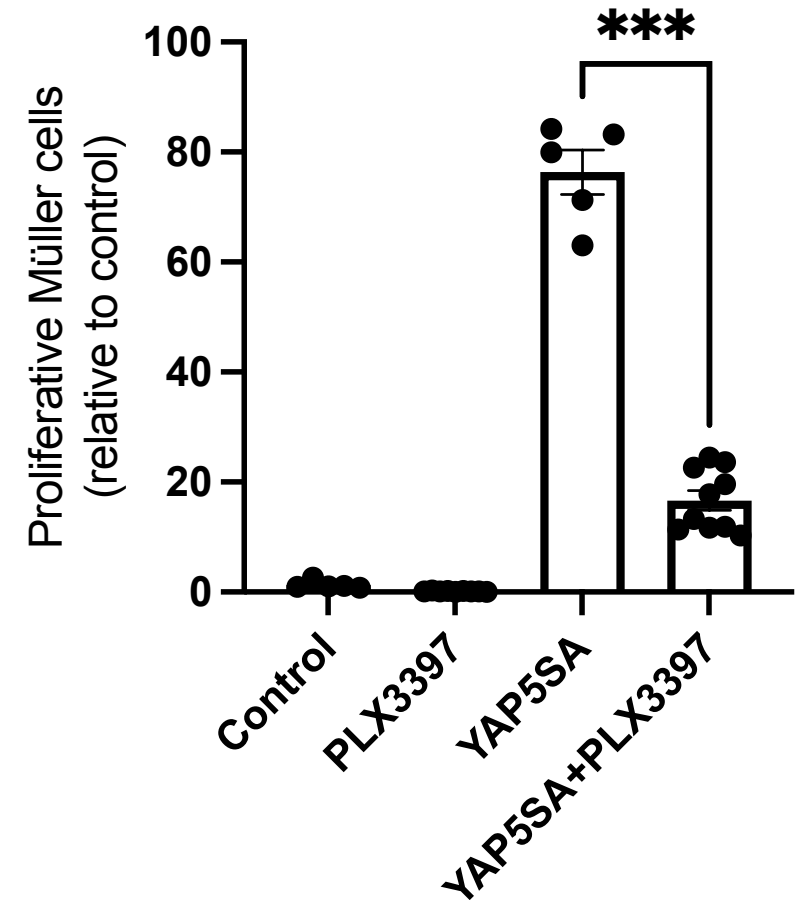
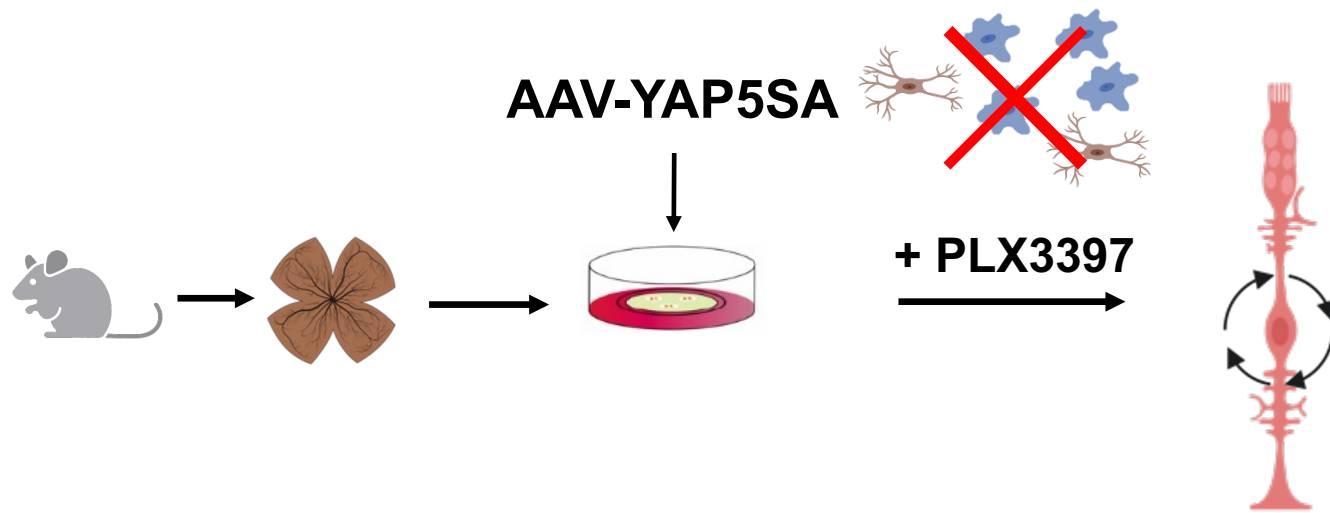
Control



LPS

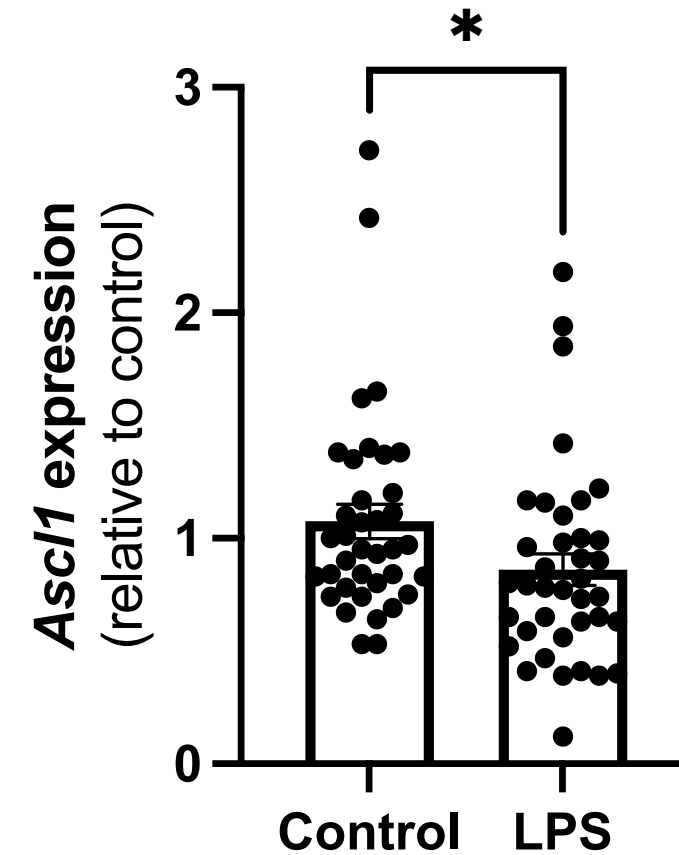
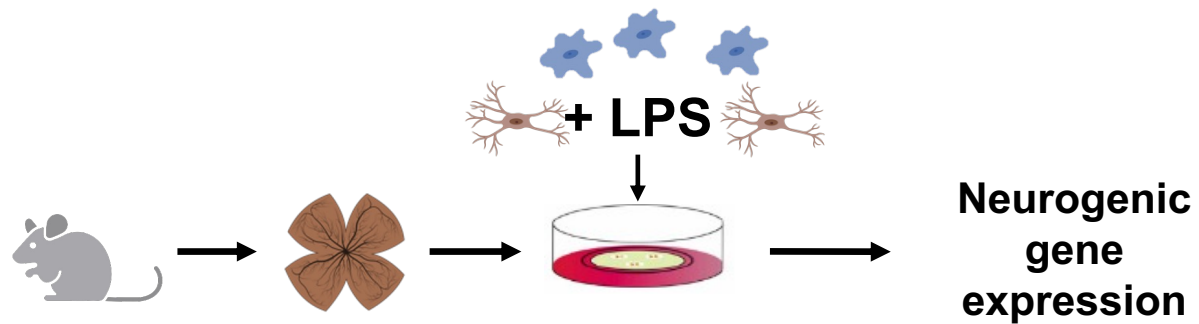


Microglia are necessary for YAP-dependent proliferative effects on Müller cells

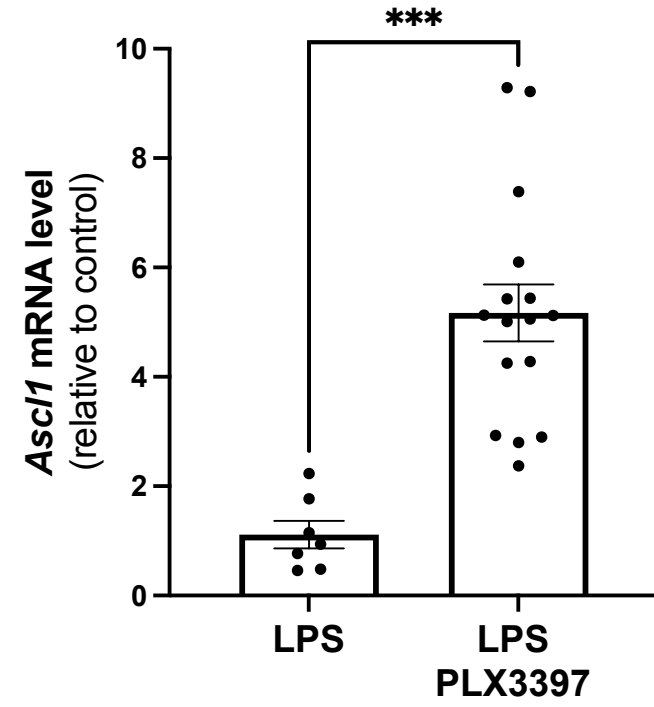
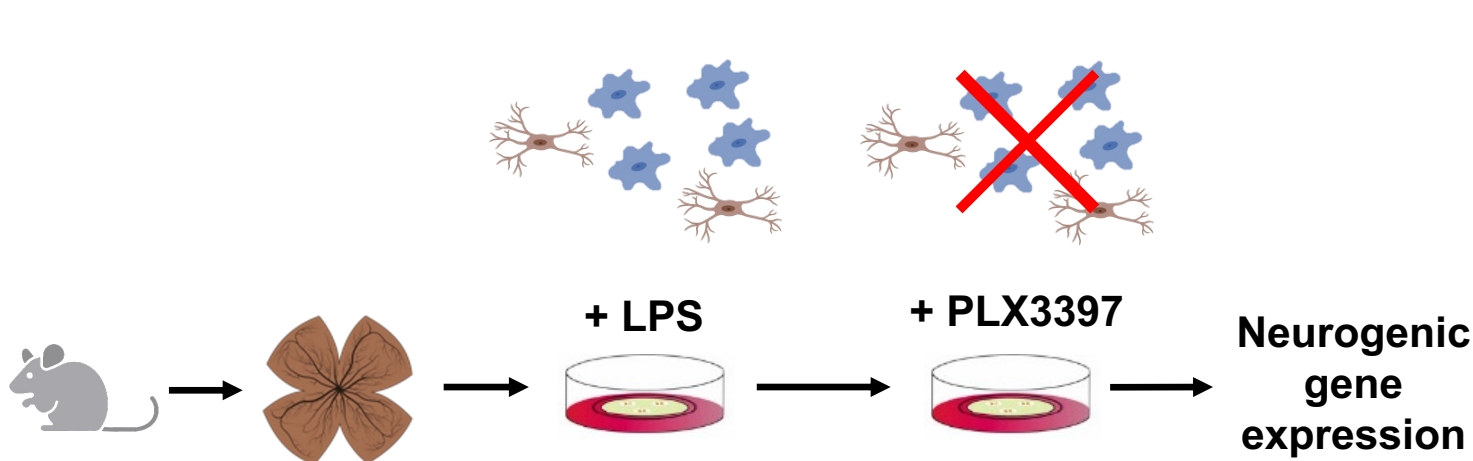


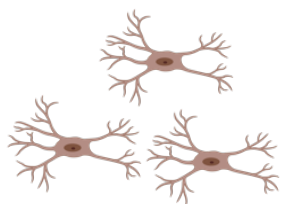
What about the neurogenic potential of LPS-induced proliferative Müller glia?

LPS-induced proliferative Müller cell are not neurogenic

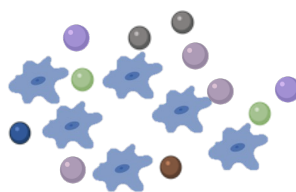


Ablation of microglia following an immune challenge triggers *Asc/1* expression





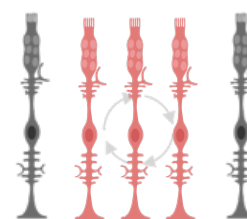
Microglia



Activated microglia



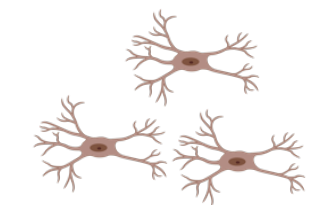
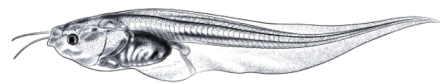
Quiescent
Müller cells



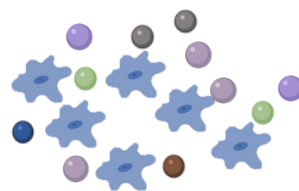
Proliferative
Müller cells



Neurogenic
Müller cells



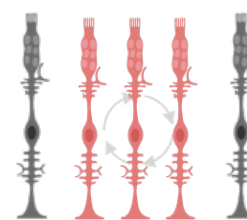
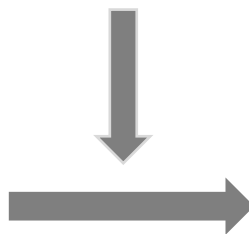
Resting microglia



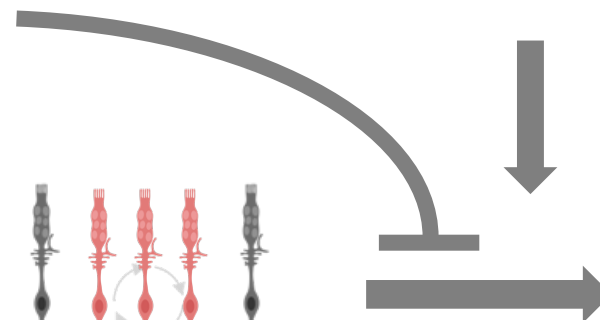
Activated microglia



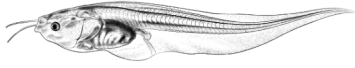
Quiescent
Müller cells



Proliferative
Müller cells



Neurogenic
Müller cells



Caroline Borday
Odile Bronchain
Albert Chesneau
Alicia Donval
Morgane Locker
Léa Siron
Jing Xian Lun
Karine Parain



Juliette Bitard
Sophie Lourdel
Patrick Pla
Julien Ricard
Jérôme Roger
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