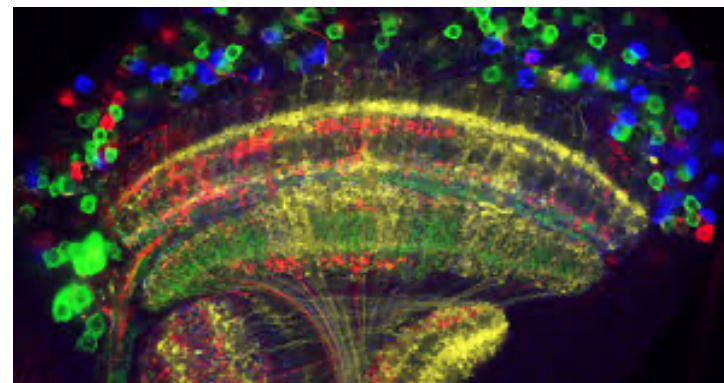
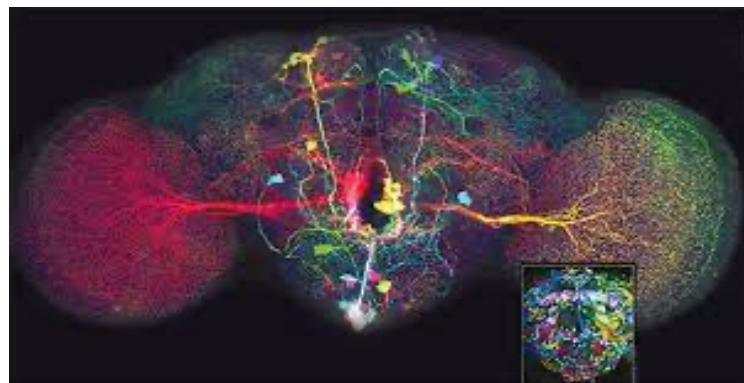
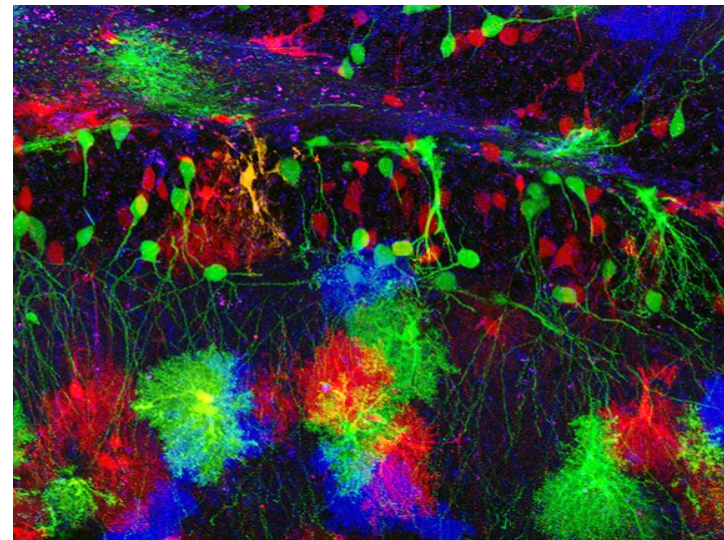
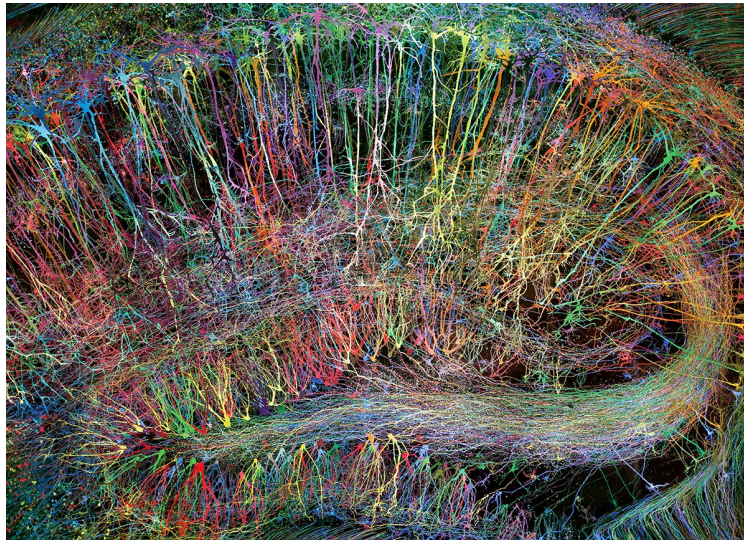


# Cell Cycle and Cell Determination

Michel.gho@sorbonne-universite.fr



# Cell Cycle and Cell Determination

Formation of organisms from a totipotential cell (the zygote), or the formation of body part from precursor cells, involves processes such as:

- Cell proliferation
- Cell determination
- Cell differentiation
- Cell shape changes
- Cell migration

All take place over time.

How these processes are temporally and spatially regulated/coordinated?

# Cell Cycle and Cell Determination

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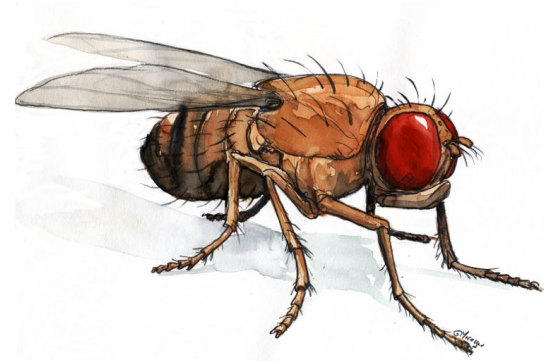
Cell proliferation  
Cell determination

Cell differentiation  
Cell shape changes  
Cell migration

All take place over time.

How these processes are temporally and spatially regulated/coordinated?

## Cell Cycle and Cell Determination



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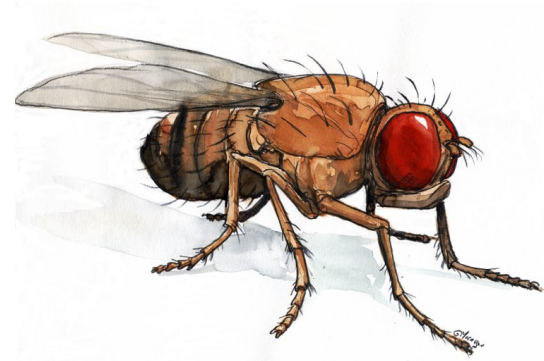
All take place over time.

How these processes are temporally and spatially regulated/coordinated?



# Neurogenesis in *Drosophila*

Cell cycle and neural cell determination in  
*Drosophila*



Life cycle

Neurogenesis

Mutual and lateral inhibition

Asymmetric cell division

Origin of neural precursor cell diversity

Spatial clues

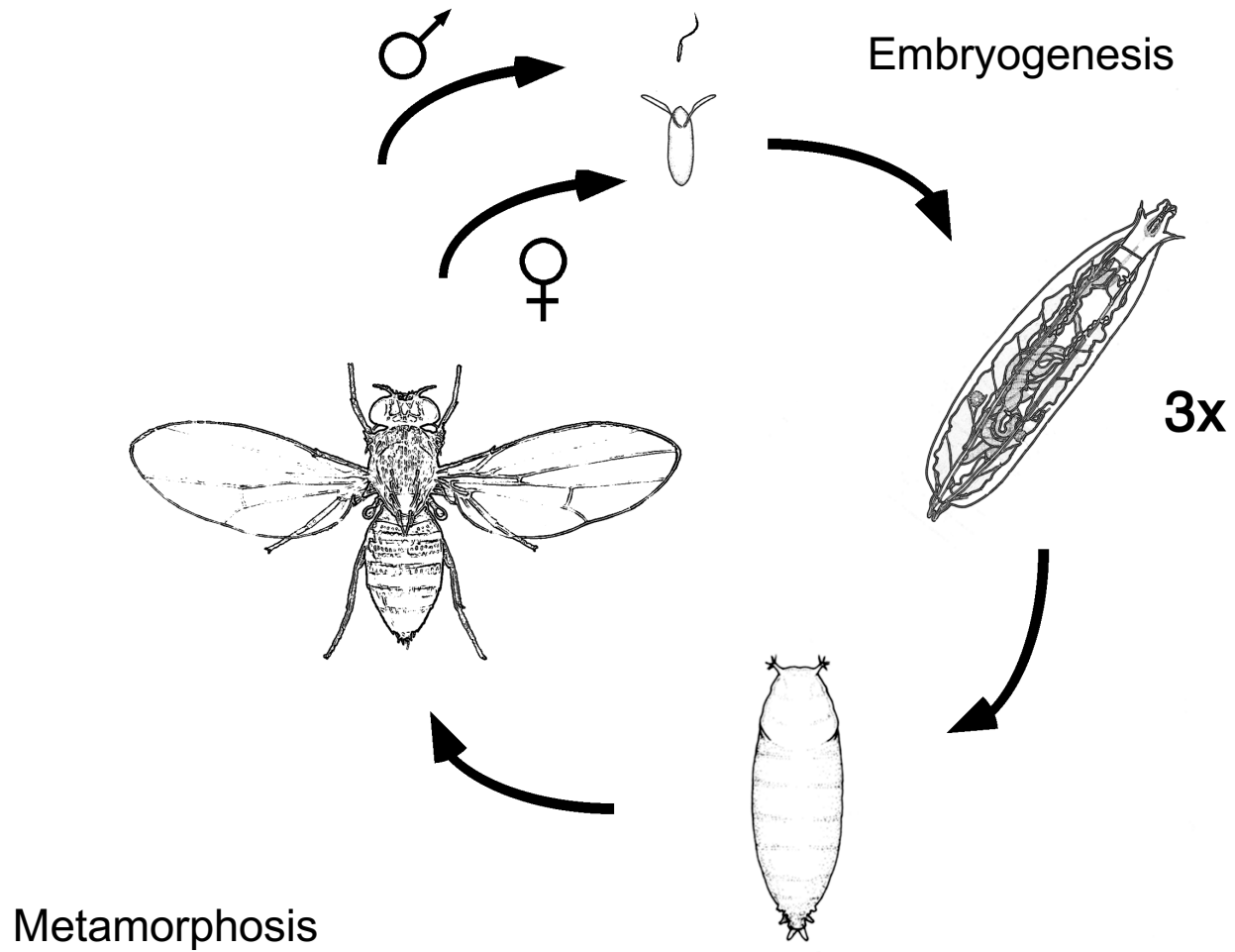
Temporal clues

Time in development

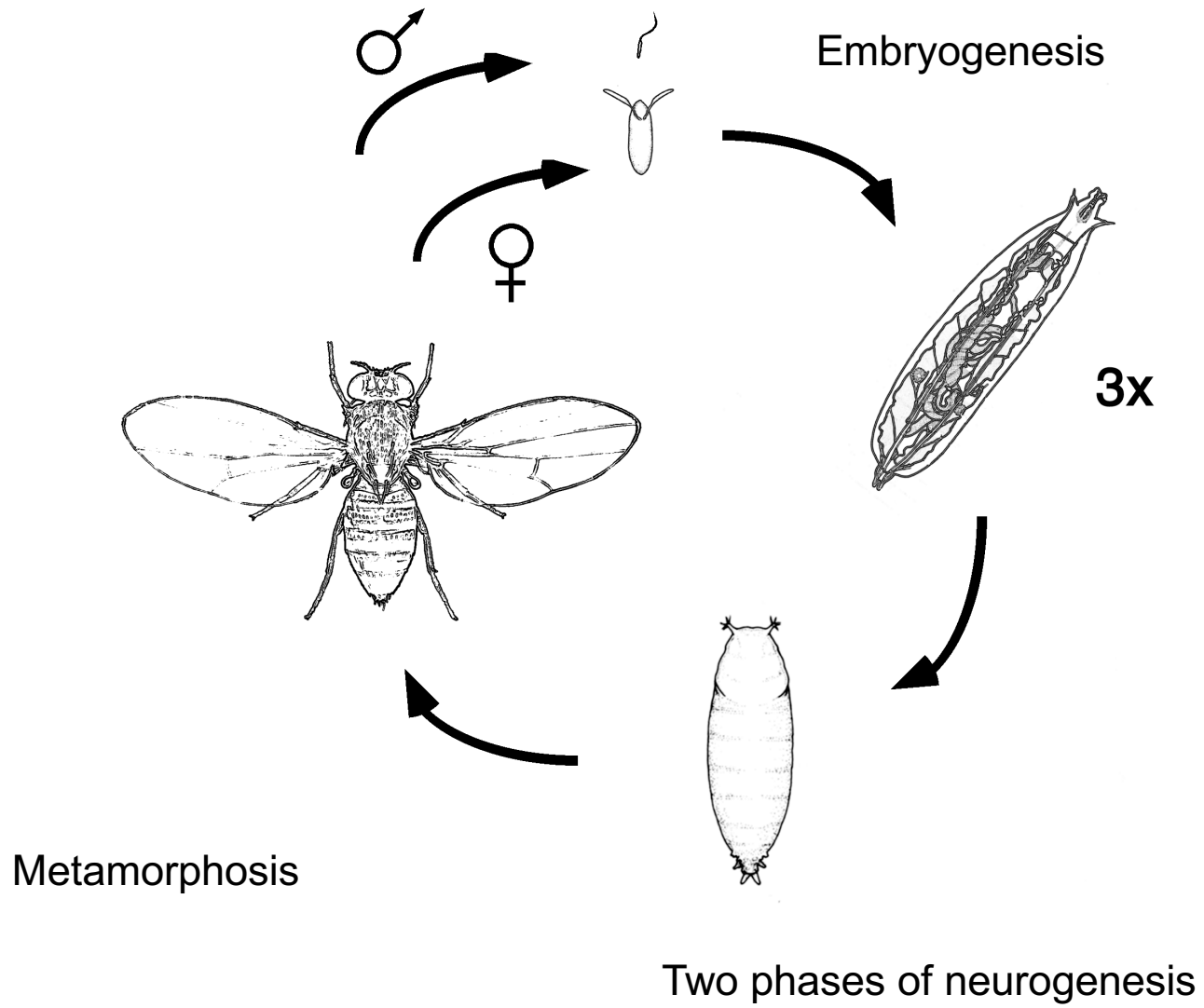
Temporal patterning

**Technical goals :** Molecular and genetic tools commonly used in *Drosophila*

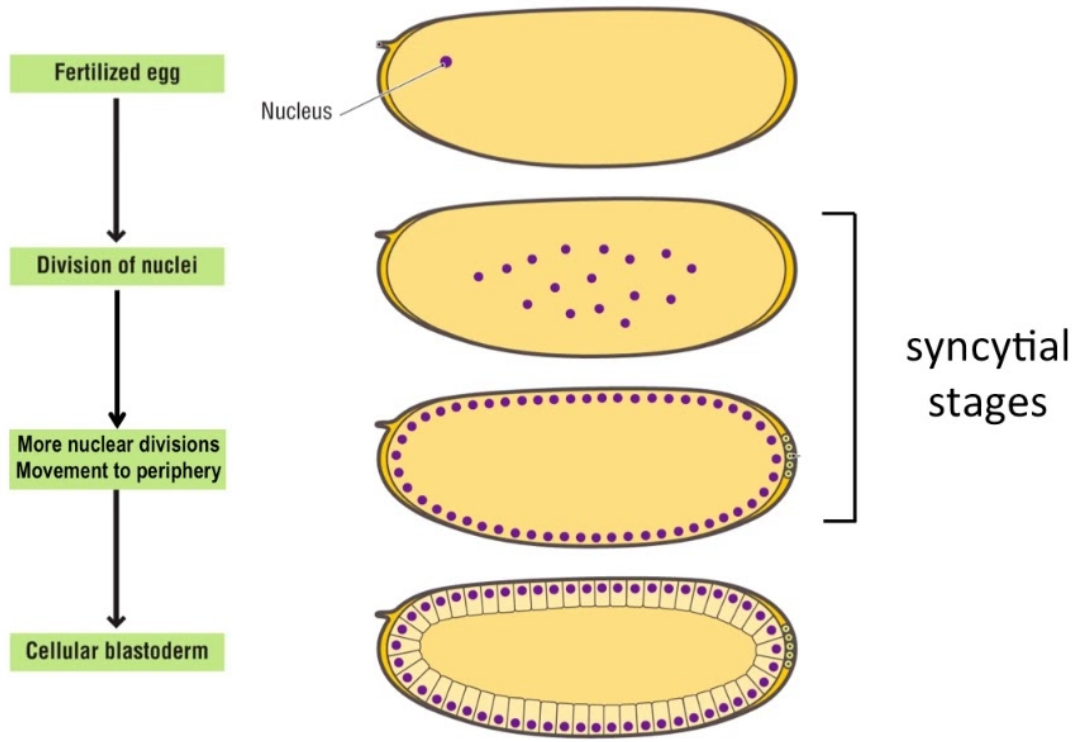
Life cycle



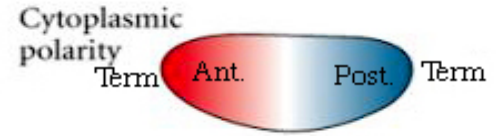
Life cycle



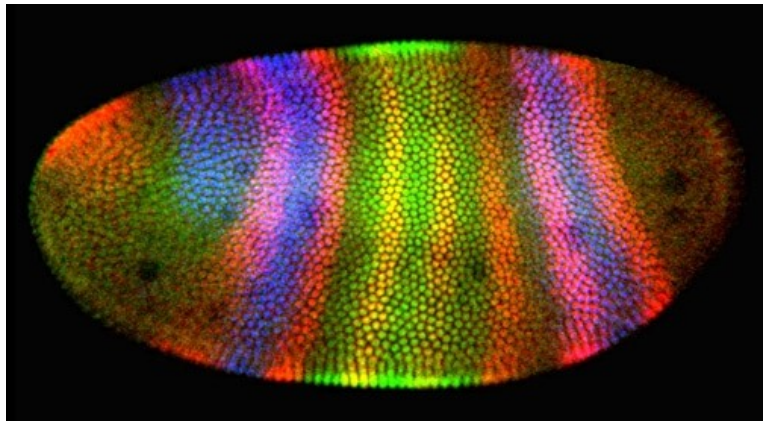
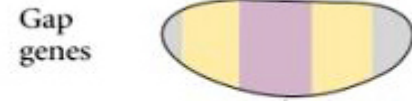
# Early stages of *Drosophila* development



Maternal Effect Genes



Zygotic Genes



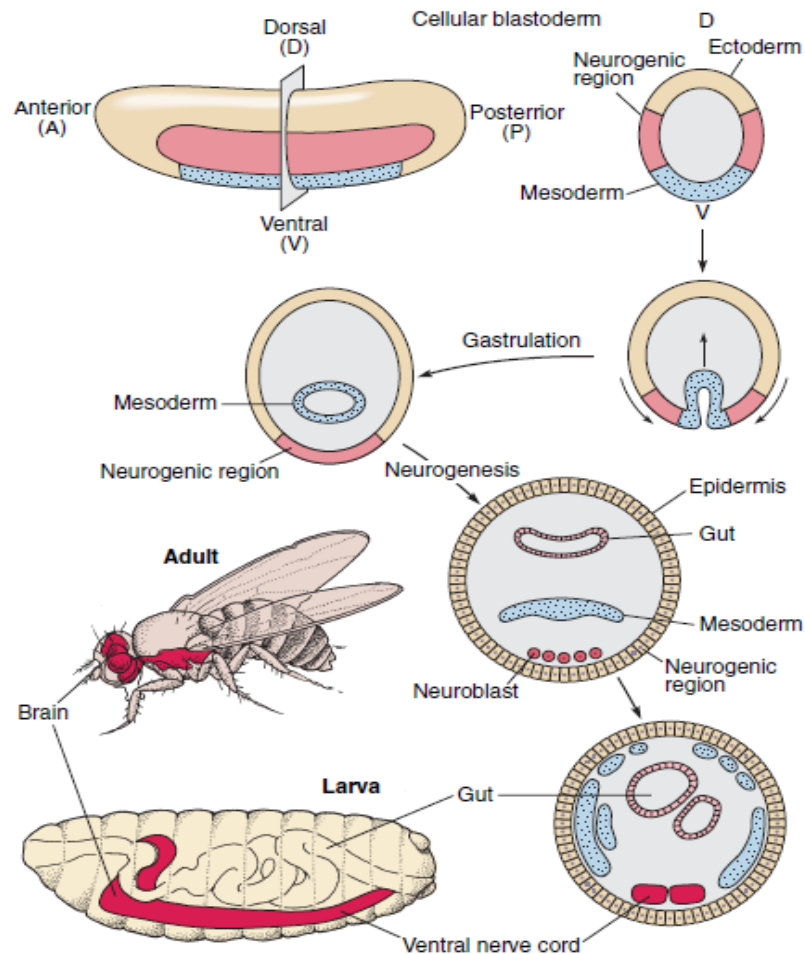
Hairy Krüppel Giant



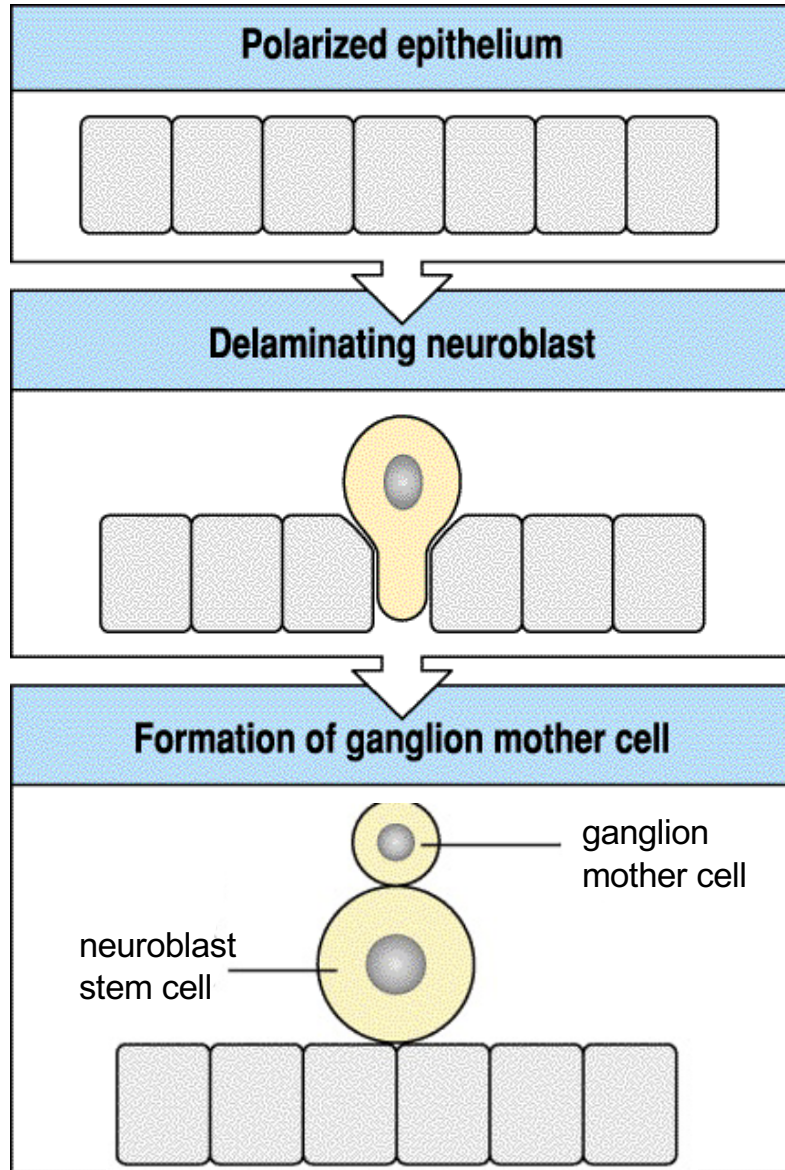
# Neurogenesis

## Neuroblast formation

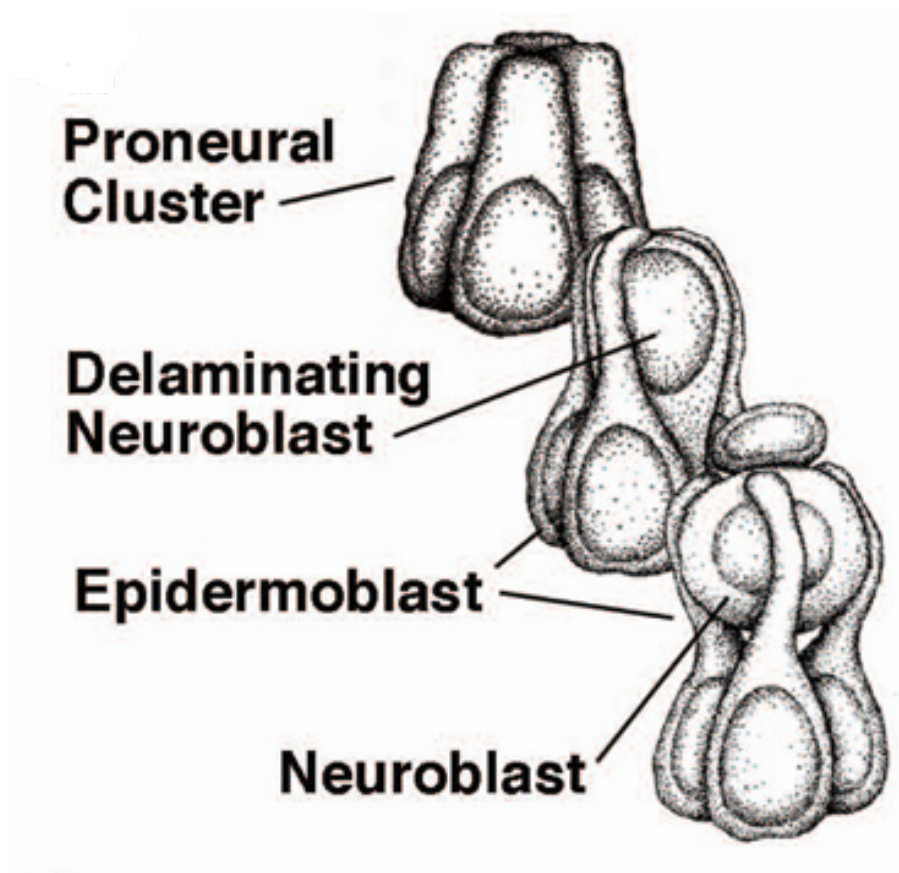
Neurons are generated from ventrolateral part of embryo (neuroectoderm)



- Cellularisation -> blastoderm
- Ventral furrow marks start of gastrulation
- Mesoderm invagination
- Ventral midline – future site of neurogenesis



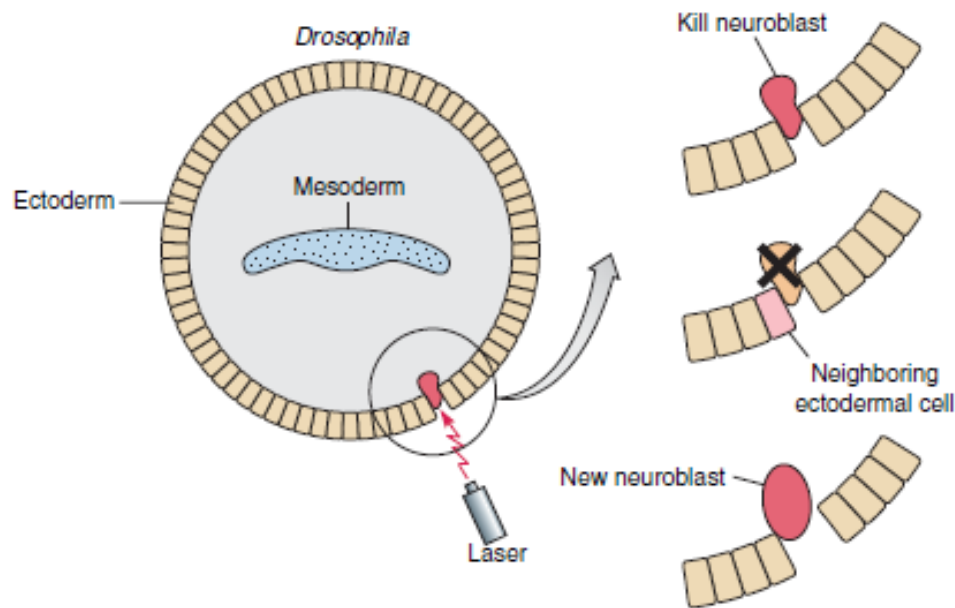
**Neuroblast formation**



## Neuroblast formation

Neuroblast selection is an active process

Neuroblasts inhibit neighboring cells to become a neuroblast



- The ablation of a delaminating neuroblast induces the redetermination of a neighbouring ectodermal cell to become a neuroblast
- Neuroblast inhibits neighbouring cells from becoming neuroblasts

## Neuroblast formation

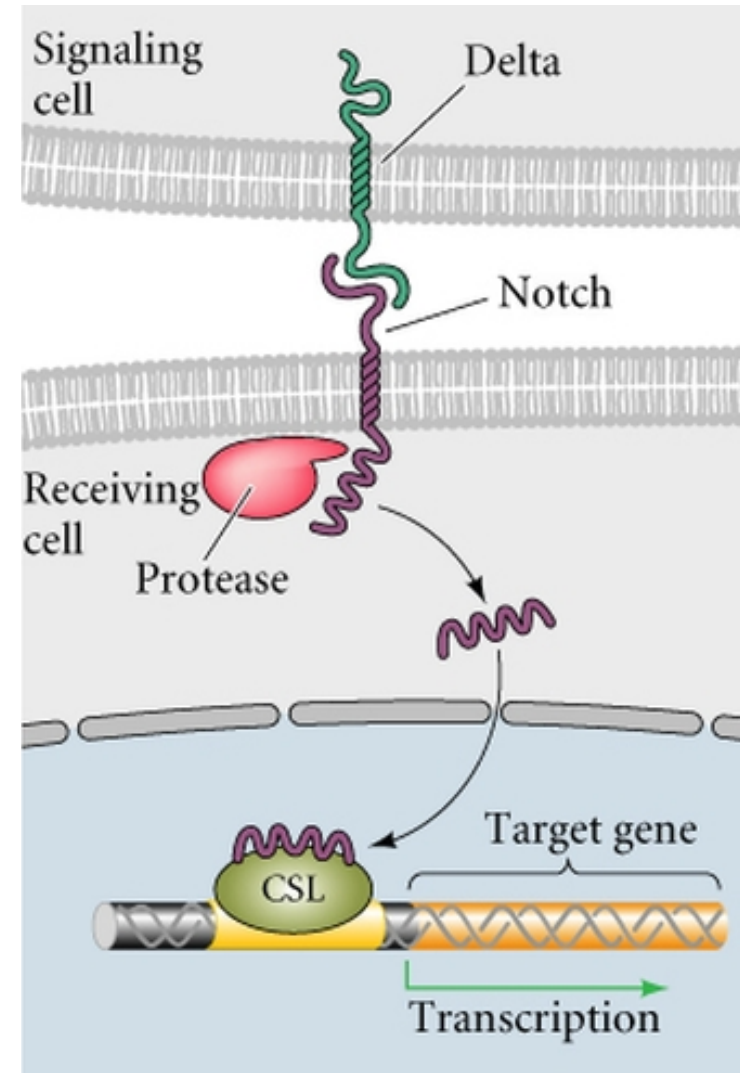
Neuroblast selection is an active process

### Proneural Genes

- Achaete-Scute complex of genes (AS-C)
  - Achaete (ac), scute (sc), lethal of scute (lsc), asense (as)

### Neurogenic Genes

- Notch-Delta pathway
  - Notch (N), Delta (DI), Enhancer of split genes (E(spl)), Hairless (H), Suppressor of Hairless (Su(H)), brainiac (brn), mastermind (mam), neuralized (neu), kuzbanian (kuz), big brain (bib)...



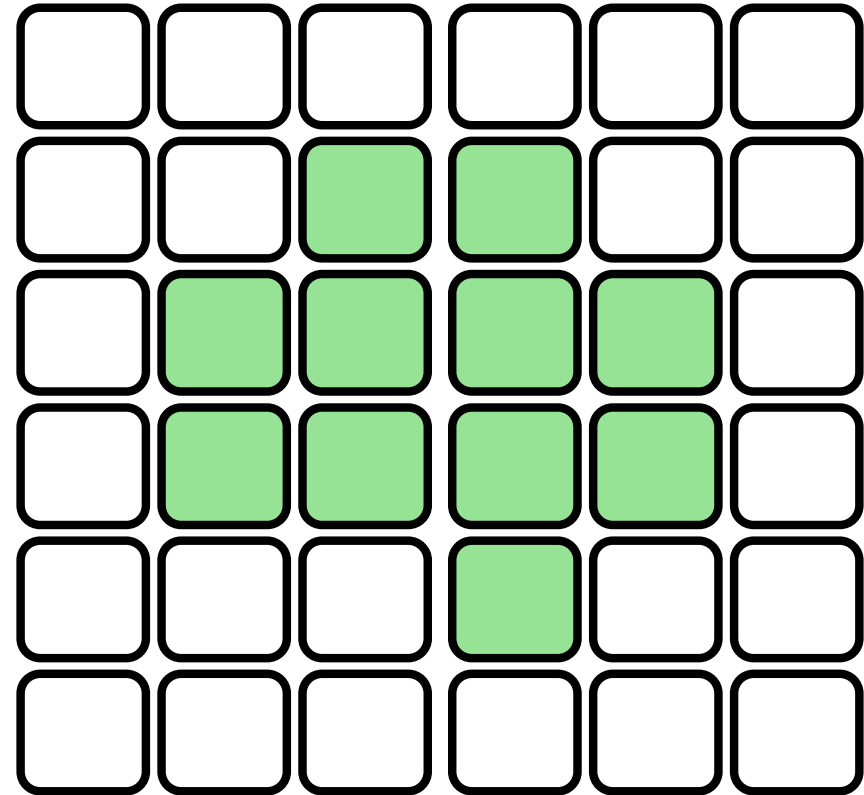


Neuroblast selection is an active process

### Proneural Genes

- Achaete-Scute complex of genes (AS-C)
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### Neuroblast formation



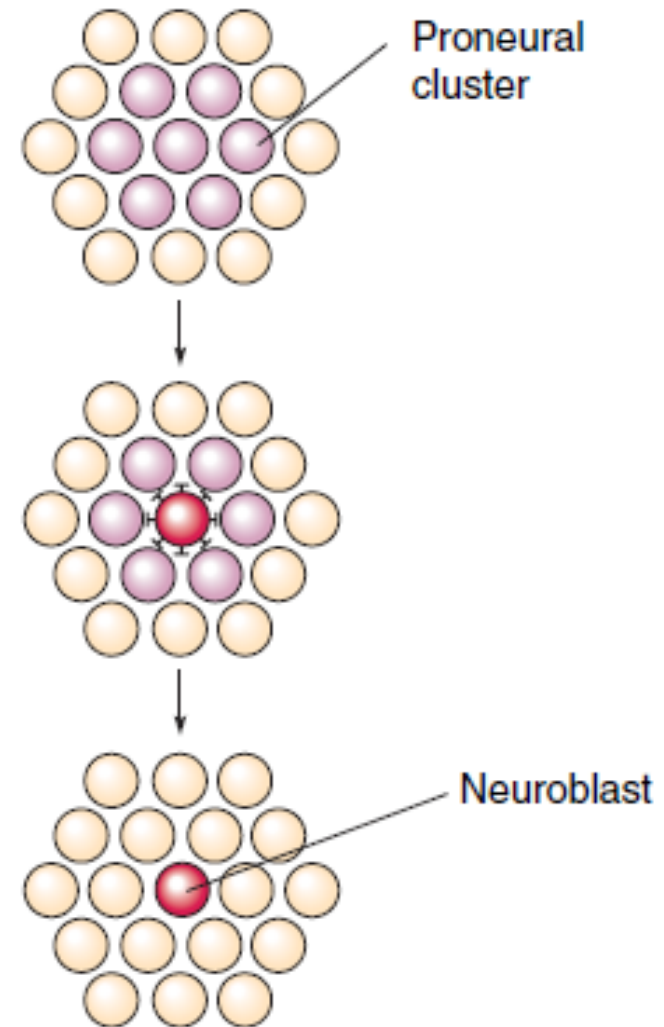
Equivalent cells

Cells expressing proneural genes become competent to acquire a neuroblast fate

Neuroblast selection is an active process

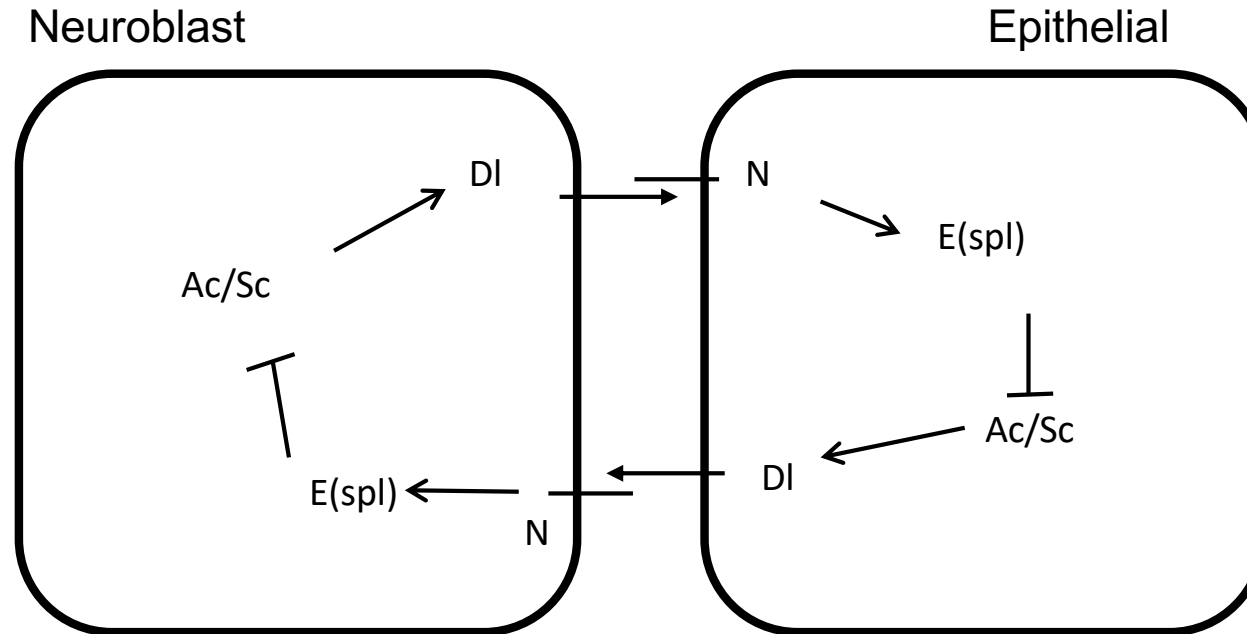
Example of an emergent property since it is not possible to know which cell will be neurogenic and which will be epithelial

### Neuroblast formation



## Neuroblast formation

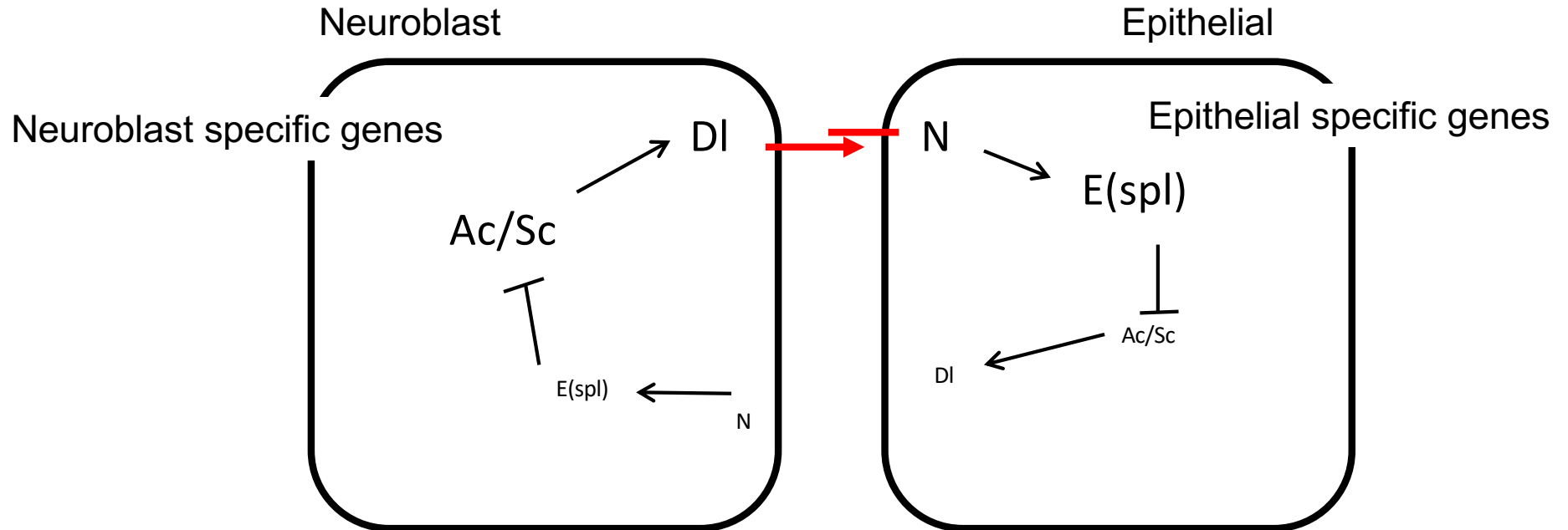
How does lateral inhibition take place?



Cells present similar levels of Notch activity  
(Mutual inhibition)

## Neuroblast formation

How does lateral inhibition take place?



Due to a stochastic (random) events one cell expresses more Delta ligand than the other.

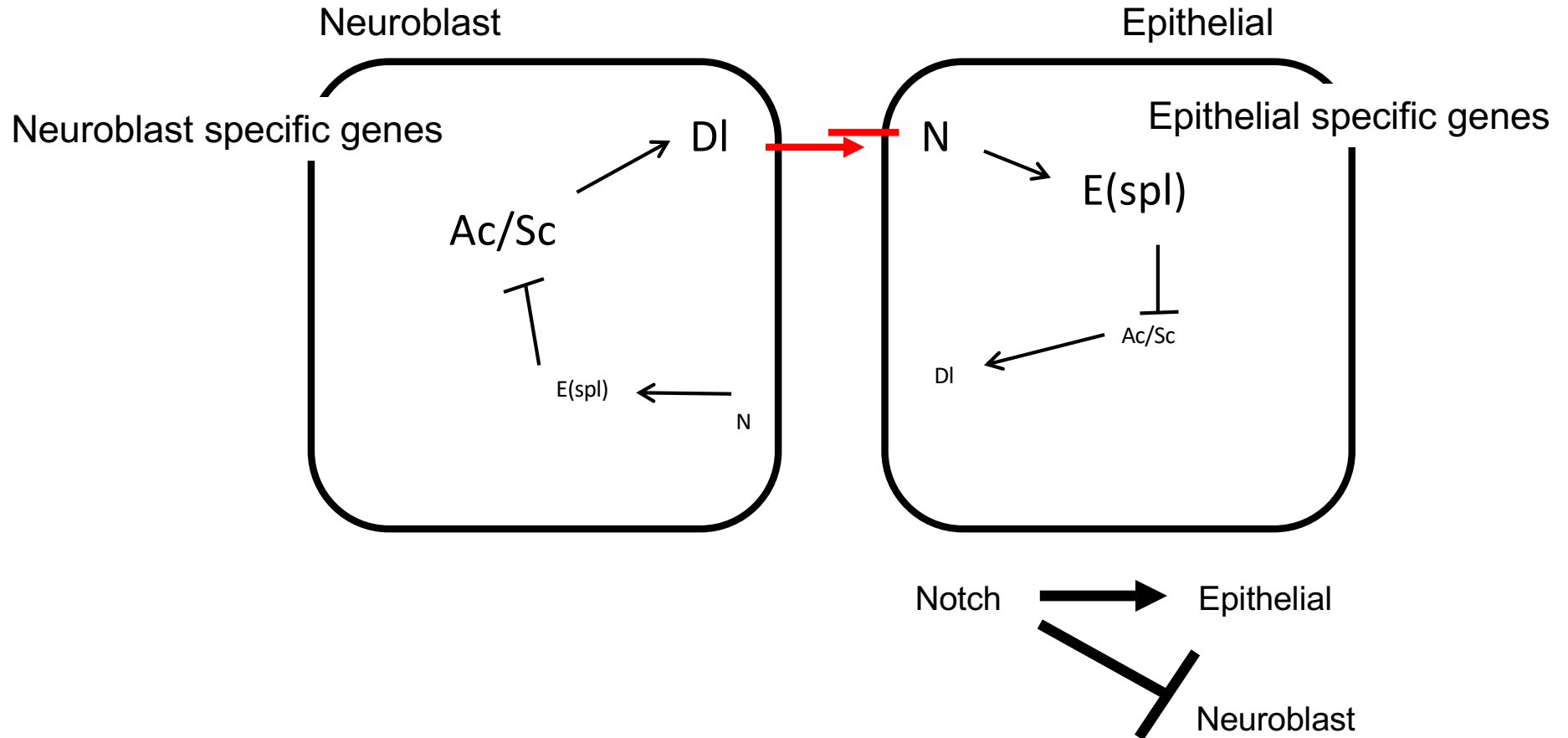
The net signal is unidirectional.

The activation of Notch receptor results in decreased Delta expression in cell that receive the signal.



## Neuroblast formation

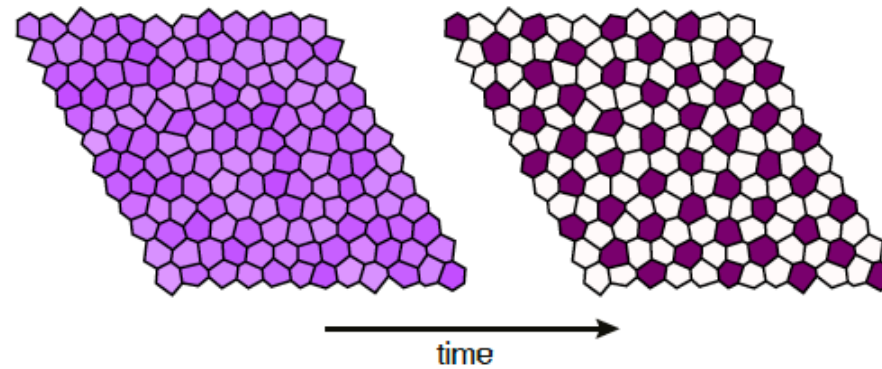
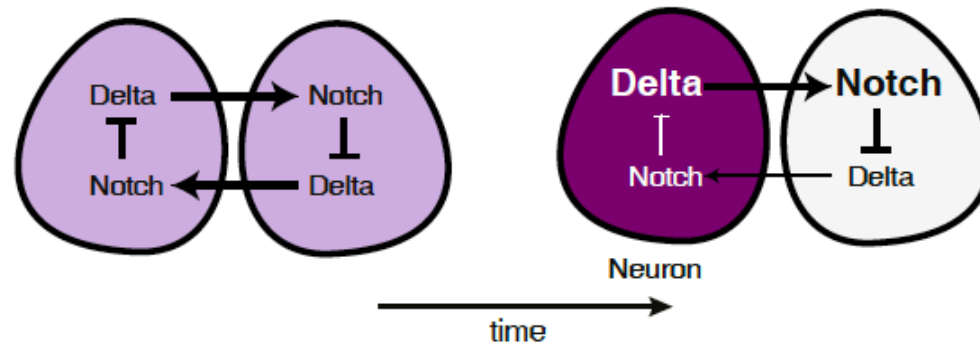
How does lateral inhibition take place?



When Notch or Delta are absent, the ectoderm cells become neurogenic. The cell that becomes the neuroblast inhibits the surrounding cells from this fate and become epithelial cells instead (lateral inhibition).

## Neuroblast formation

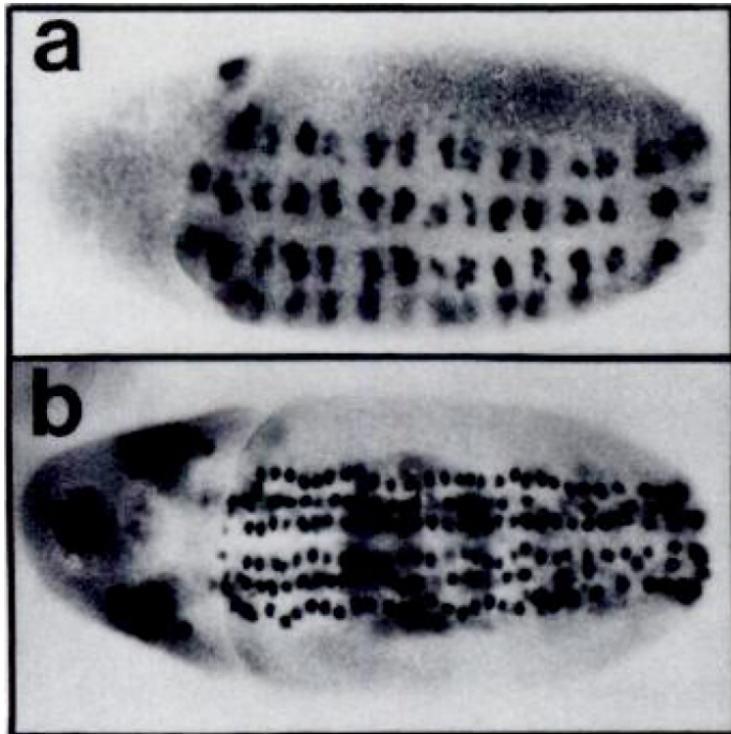
How does lateral inhibition take place?



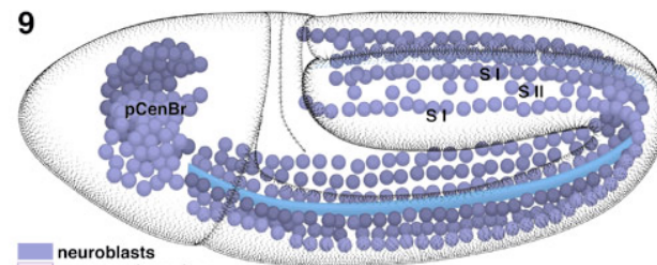
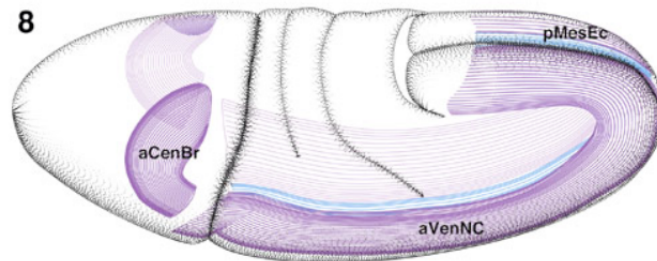
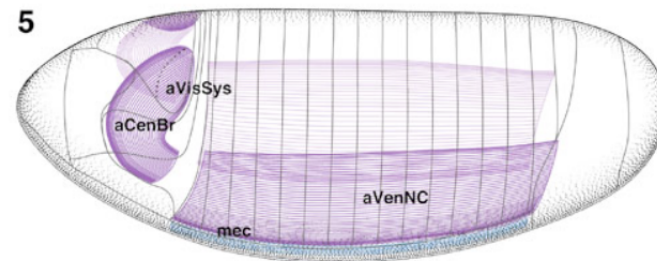
Neurogenic selection through lateral inhibition

## Neuroblast formation

Lateral inhibition during neurogenesis in *Drosophila*

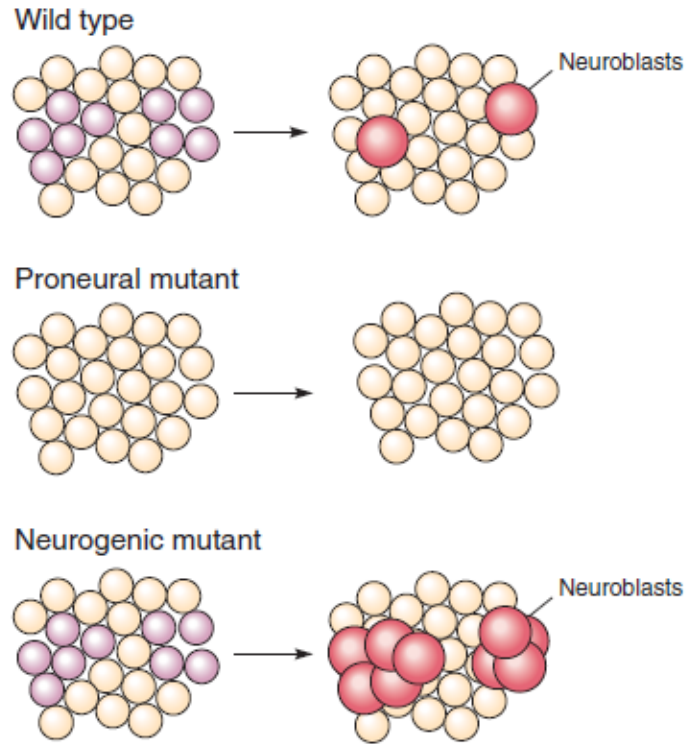


### Central Nervous System

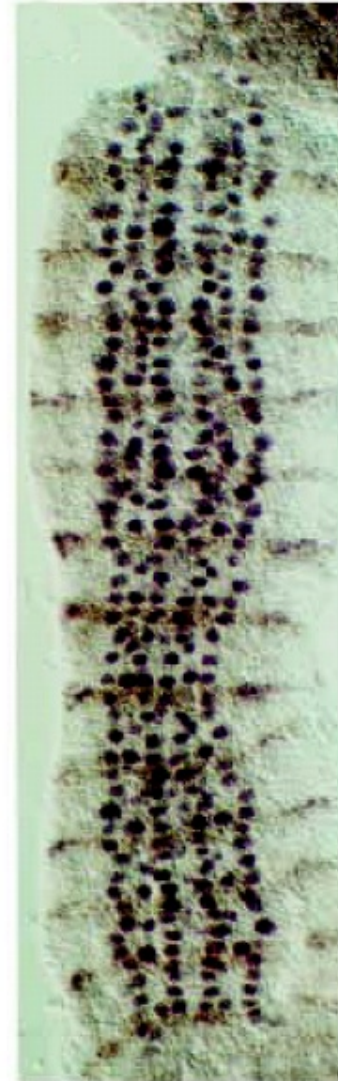


neuroblasts  
neurogenic region  
mesectoderm

*achaete-scute* complex genes and the Notch pathway are required for neuroblasts formation



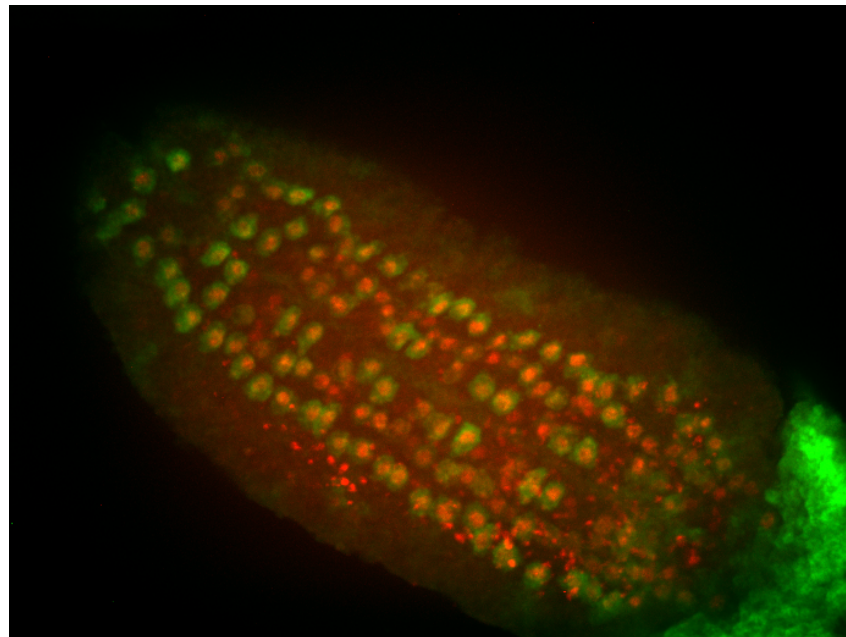
- Only one of proneural cells becomes a neuroblast in a normal conditions
- Flies mutant for proneural genes (*achaete/scute*) don't form neuroblasts
- Flies mutant for Notch or Delta – form ectopic neuroblasts



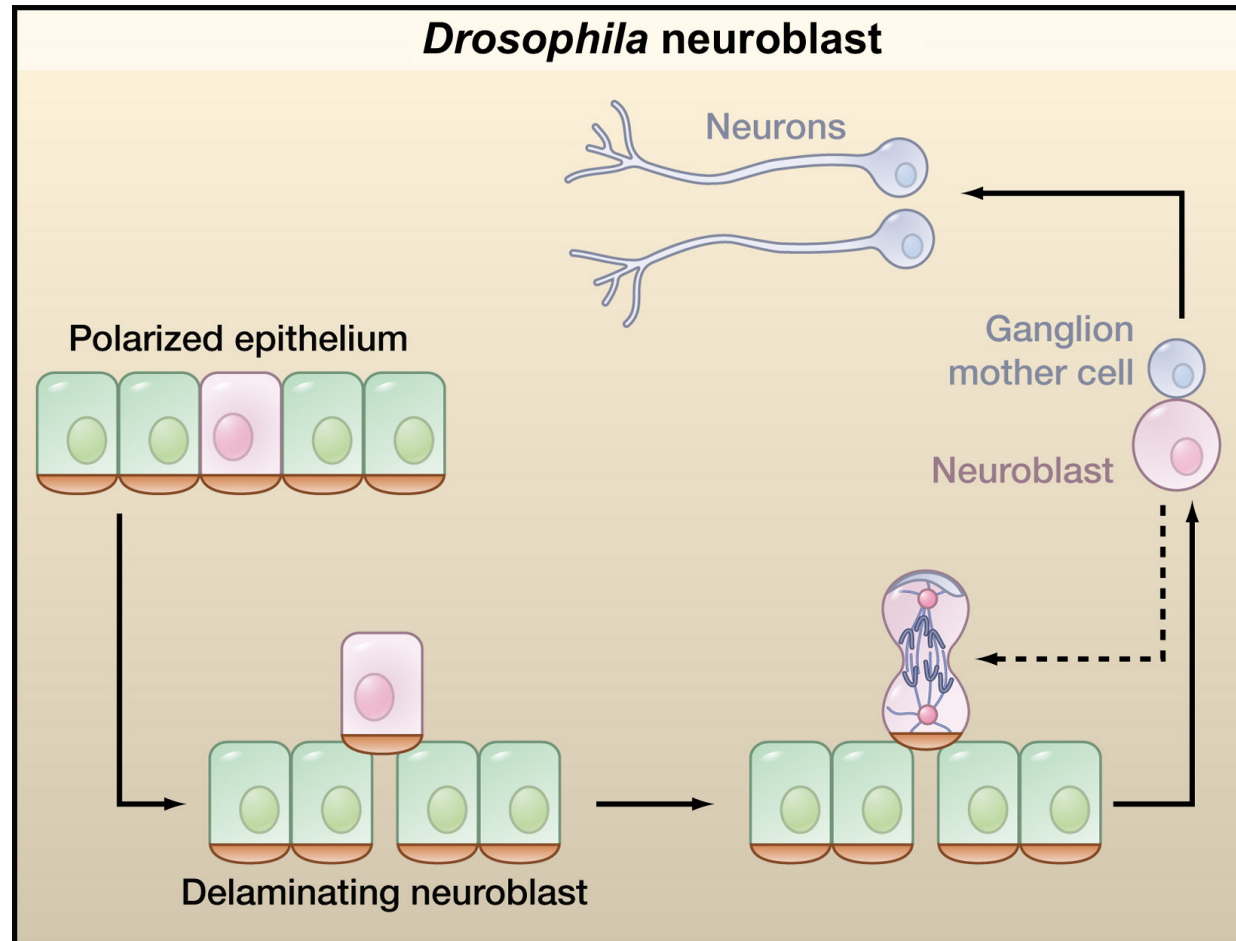


## Genetic control of neuroblasts formation

- 1) Specific genes expression pattern the neuroectoderm and pre-define territories with neural competence (pre-pattern genes)
- 2) In this territories, the expression of so-called proneural genes specified proneural fate in all cells in these clusters (proneural clusters). These genes belong to the Achaete-Scute complex (AS-C) and encode conserved basic-helix-loop-helix transcription factors
- 3) Single neuroblasts are specified within proneural clusters via the activation of so-called neurogenic genes –the Notch pathway (mutual followed by lateral inhibition)

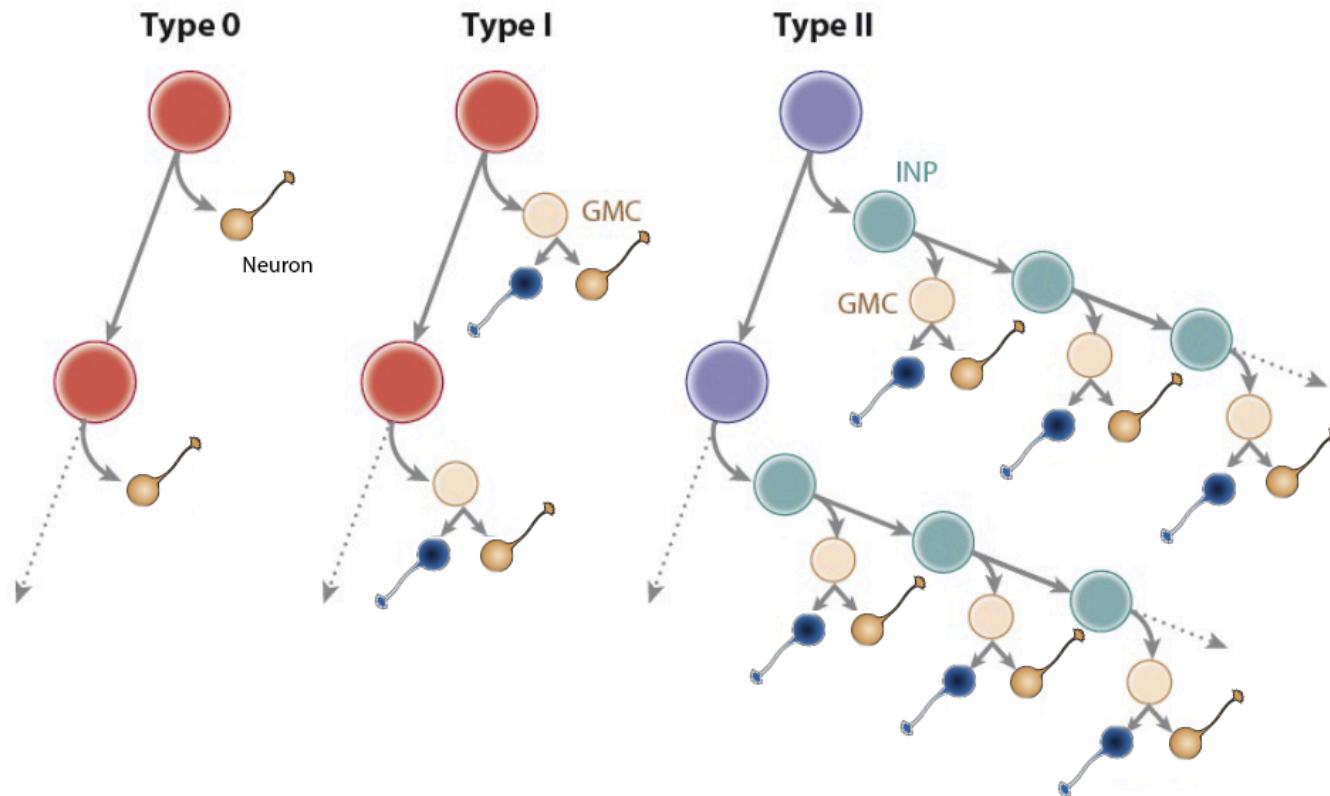


Neuroblasts undergo a self-renewing asymmetric division



(Simons & Clevers 2011)

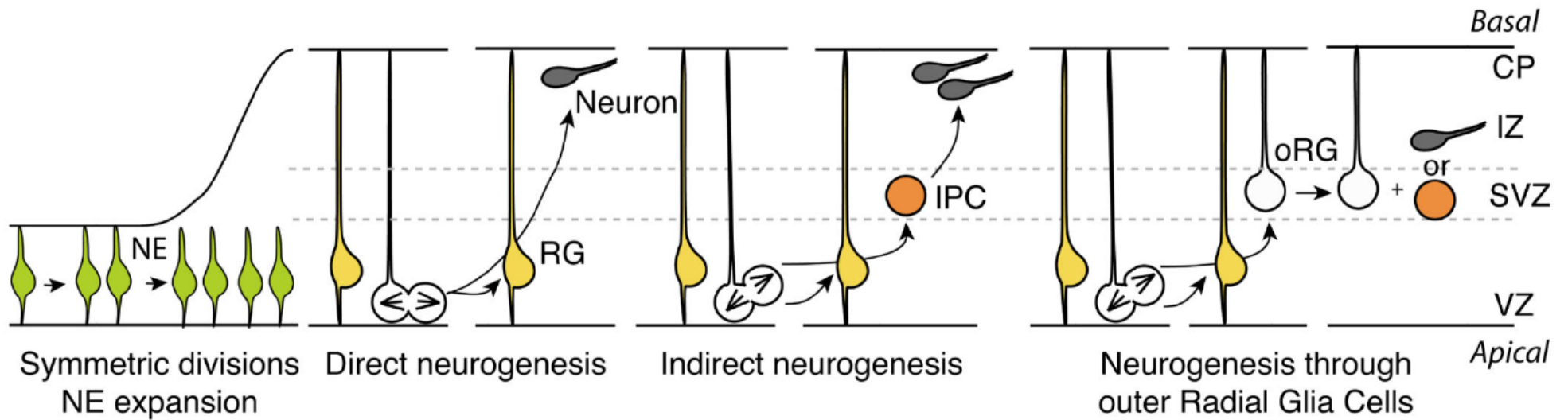
# Neuroblast cell lineage



(Doe C, 2017)

**Three modes of NB cell division.** The largest cells are NBs. Abbreviations: GMC, ganglion mother cell; INP, intermediate neural progenitor; n, neurons [all sibling neurons are either  $\text{Notch}_{\text{ON}}$  ( $n^*$ ) or  $\text{Notch}_{\text{OFF}}$  ( $n$ )].

# Neuroblast cell lineage

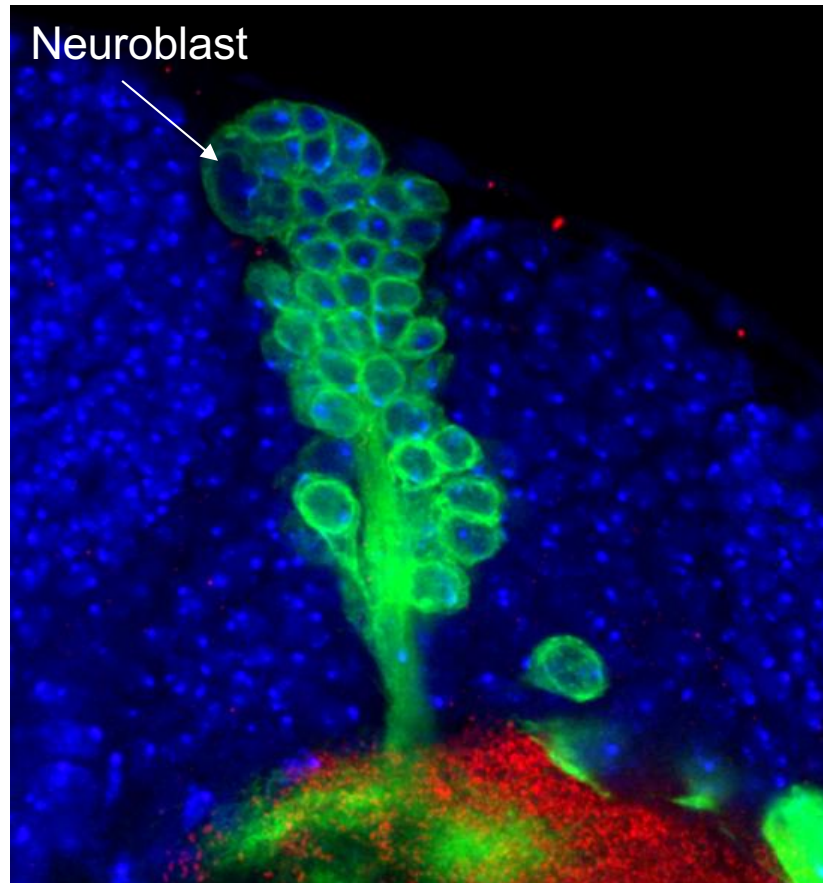


Homem et al (2015)

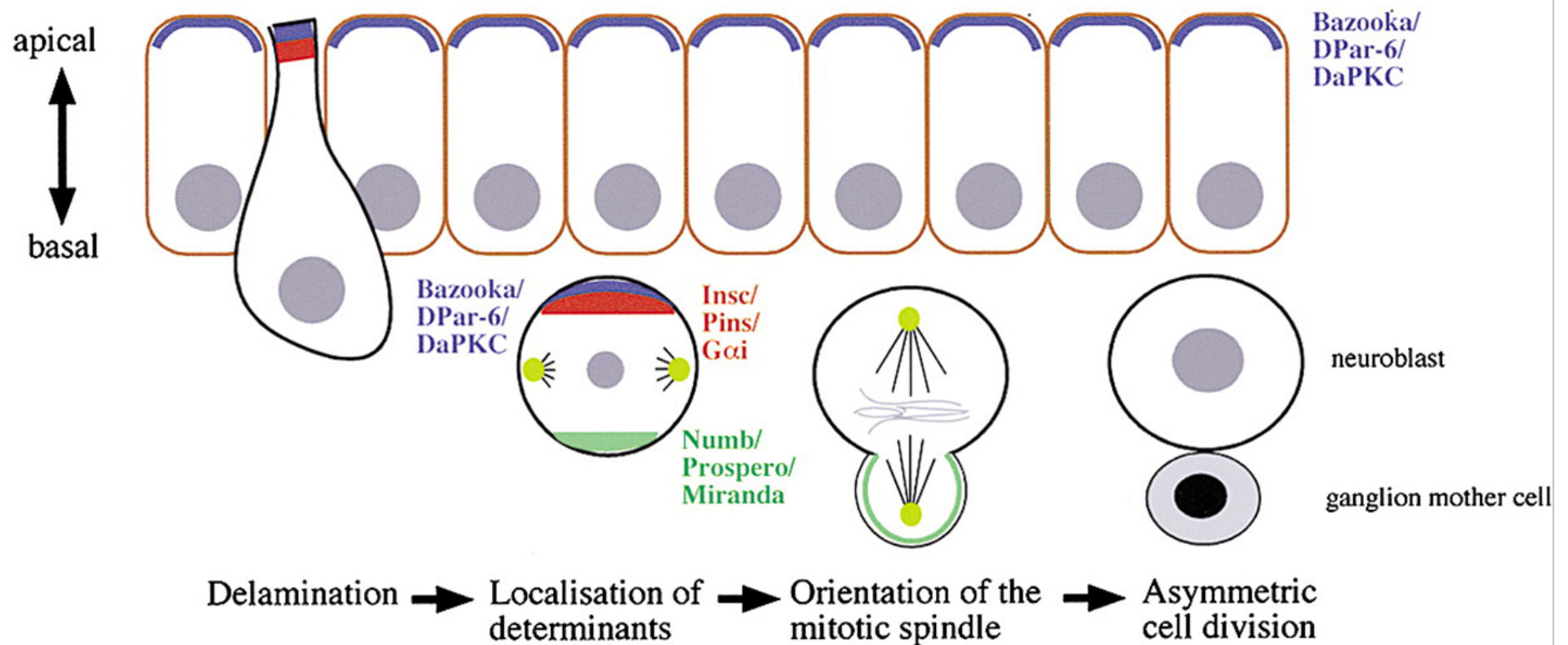
Development of the mouse neocortex.

NE neuroepithelial cells. RG radial glia cells. IPC Intermediate progenitor cell. oRG outer radial glia.

# Neuroblast cell lineage



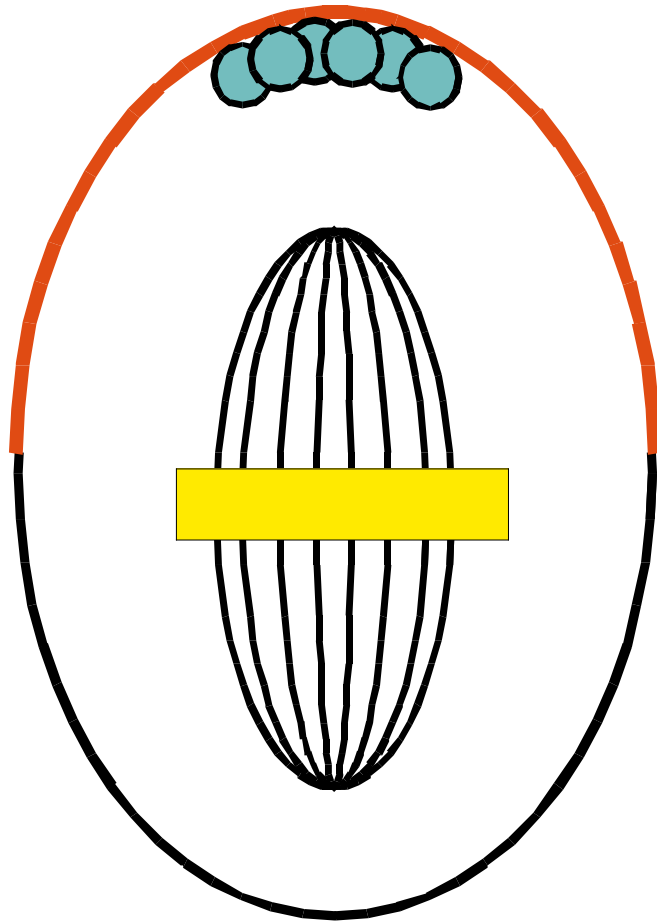
## Neuroblasts undergo a self-renewing asymmetric division



Neuroblasts divide asymmetrically

# Asymmetric cell division

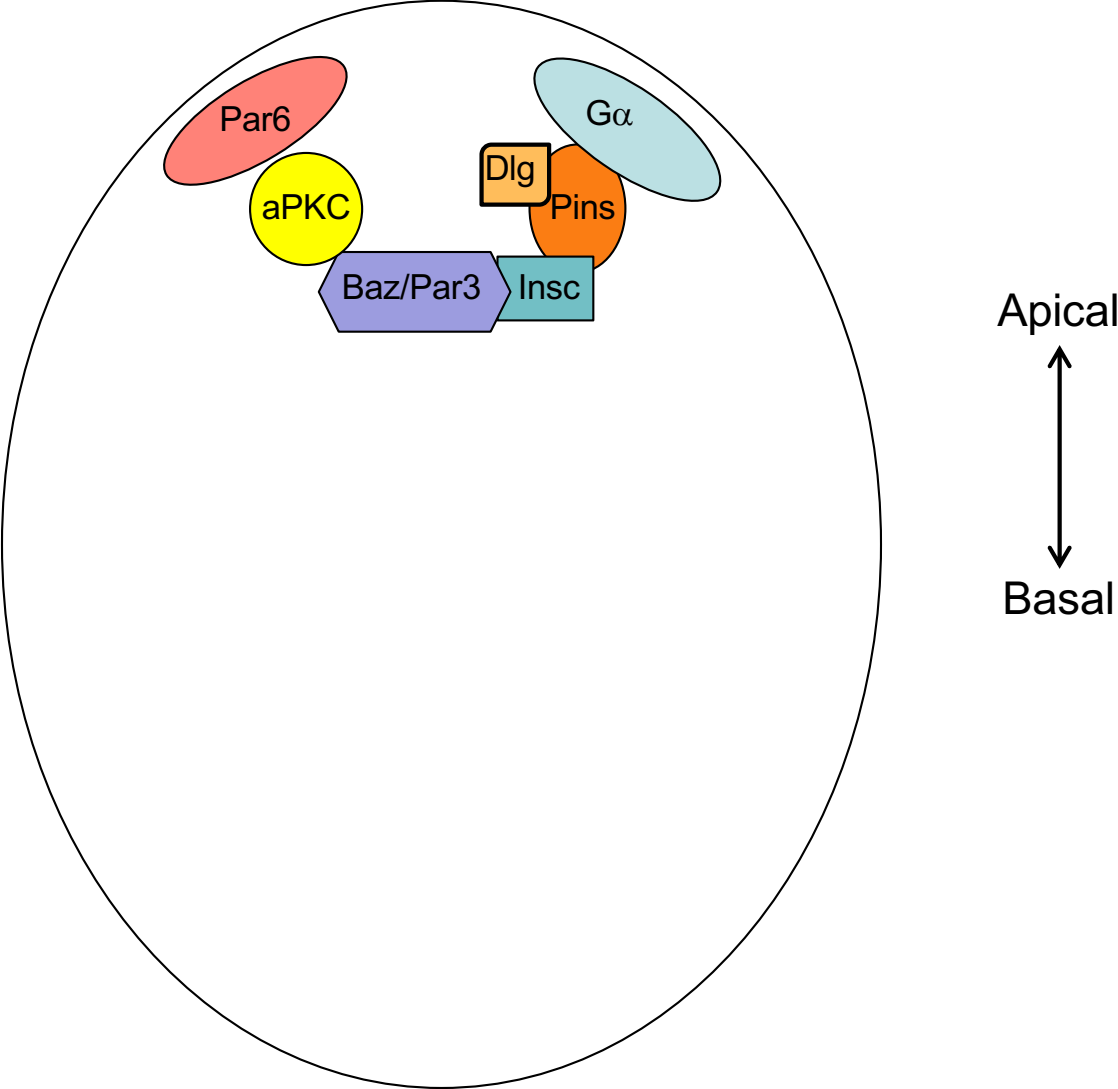
Requirements



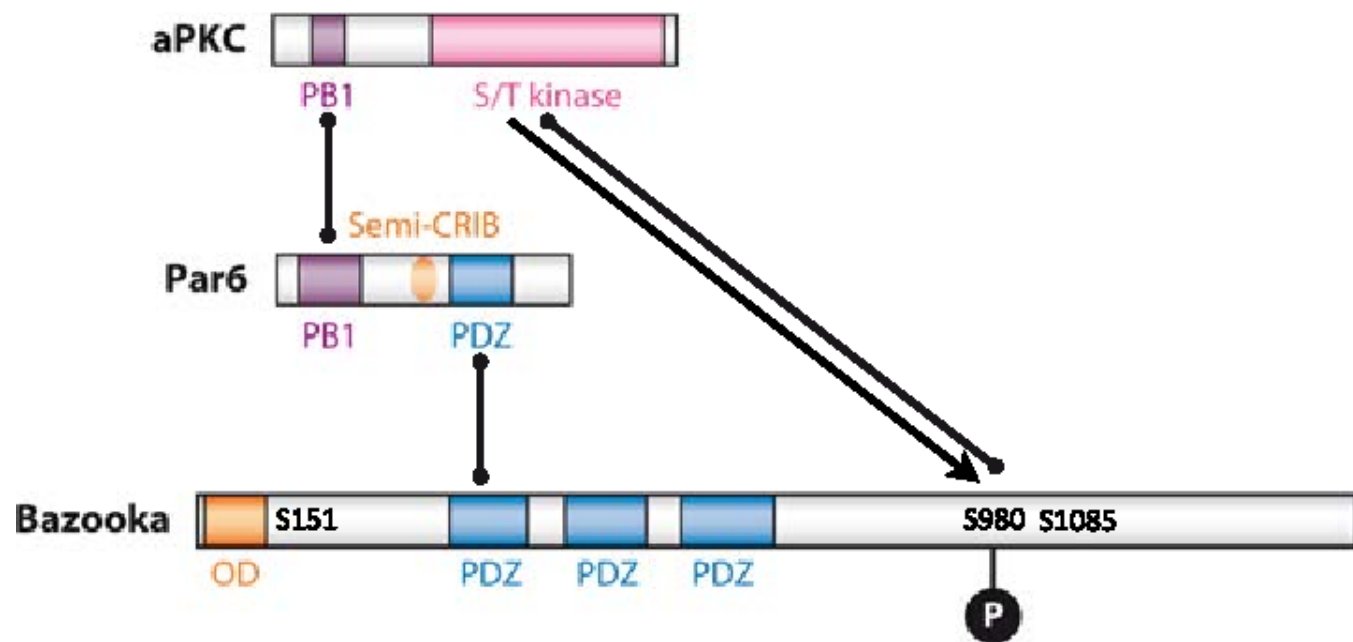
- (1) Cell polarity
- (2) Localisation of cell determinants
- (3) Mitotic spindle orientation

# Neuroblast asymmetric cell division

The apical complex

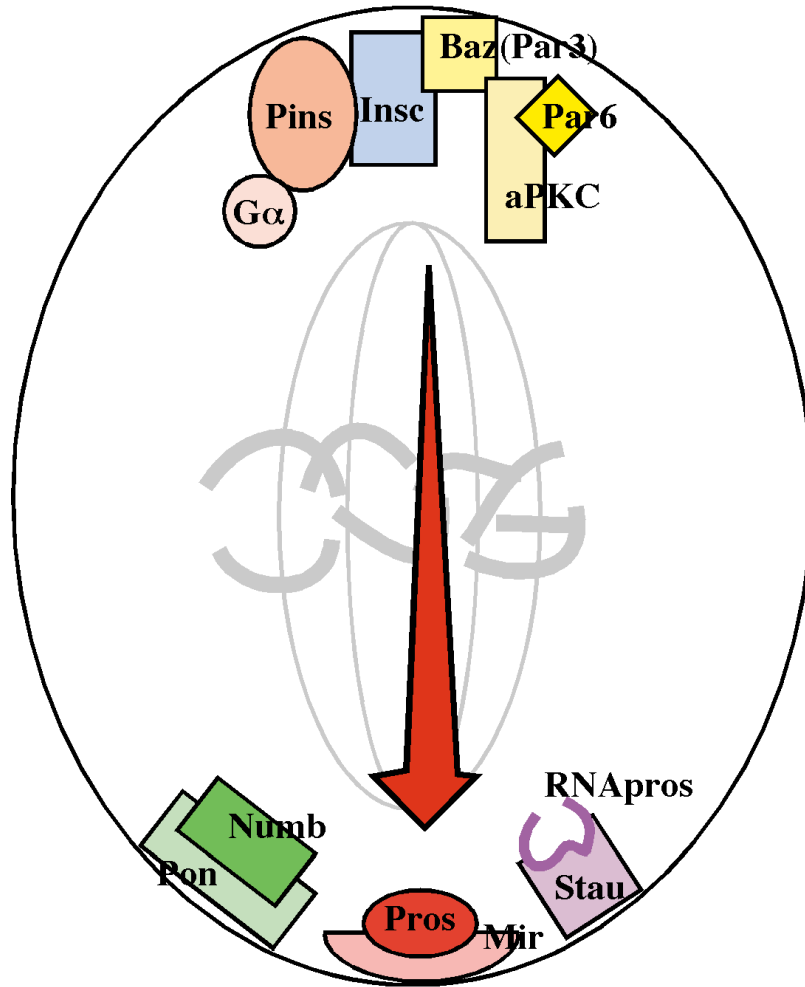






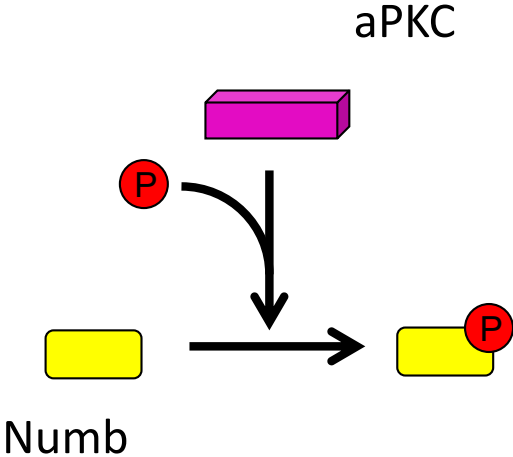
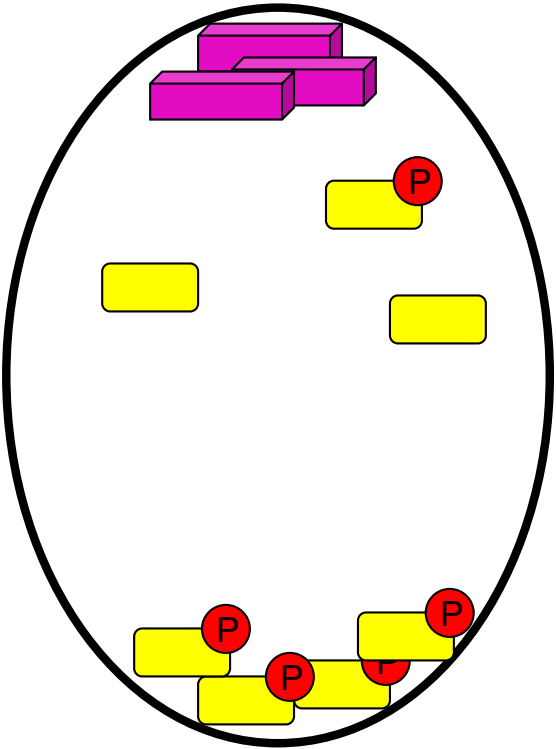
# Neuroblast asymmetric cell division

Apical complex controls basal localisation of factors during neuroblast division



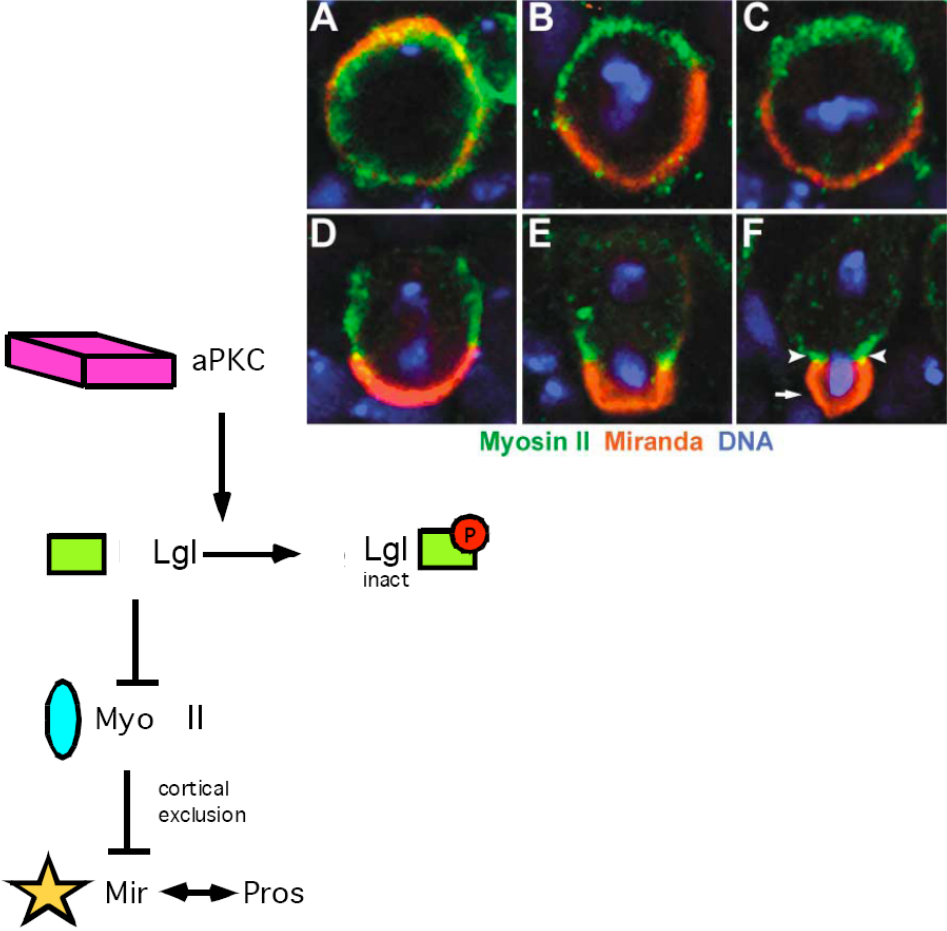
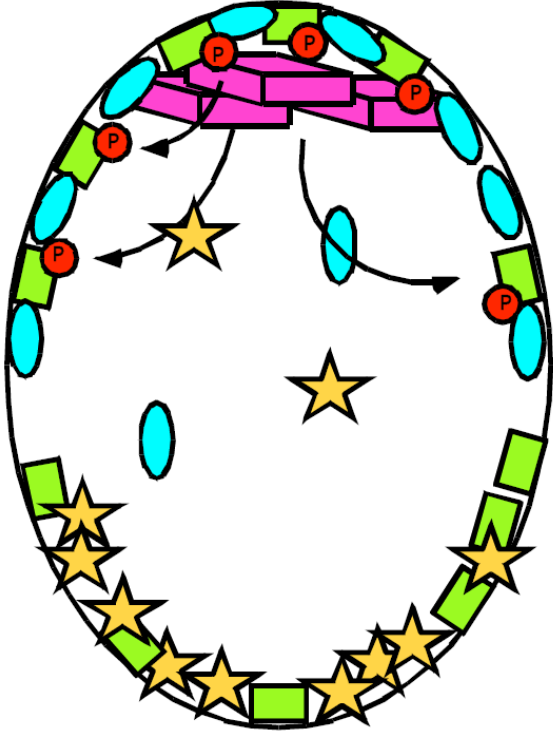
# Neuroblast asymmetric cell division

Numb



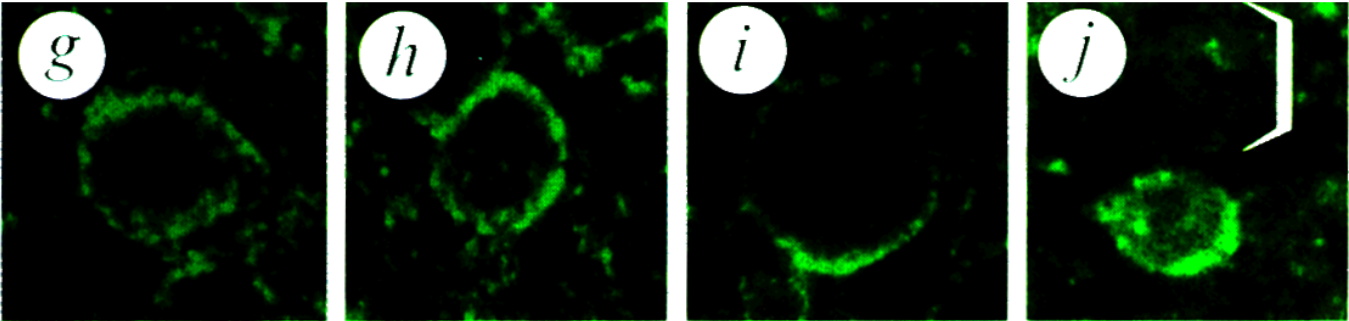
# Neuroblast asymmetric cell division

Prospero

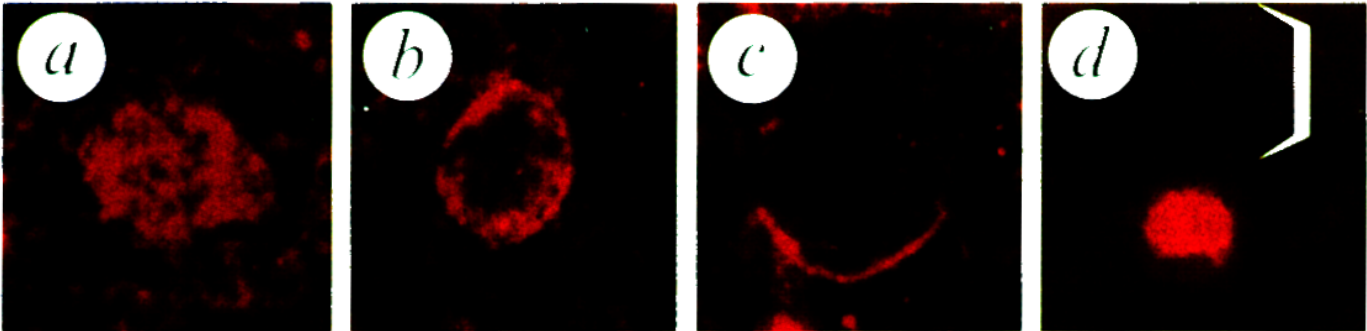


# Neuroblast asymmetric cell division

**Numb**

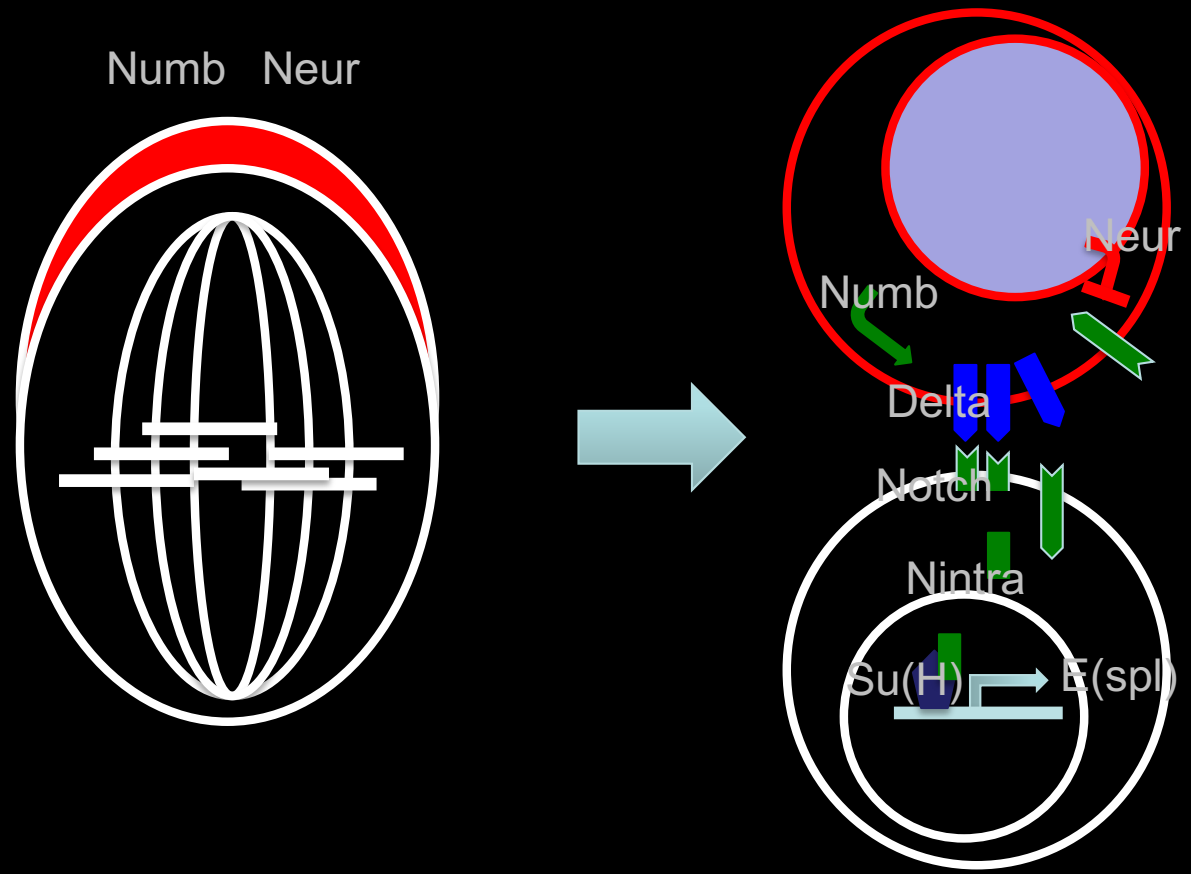


**Pros**

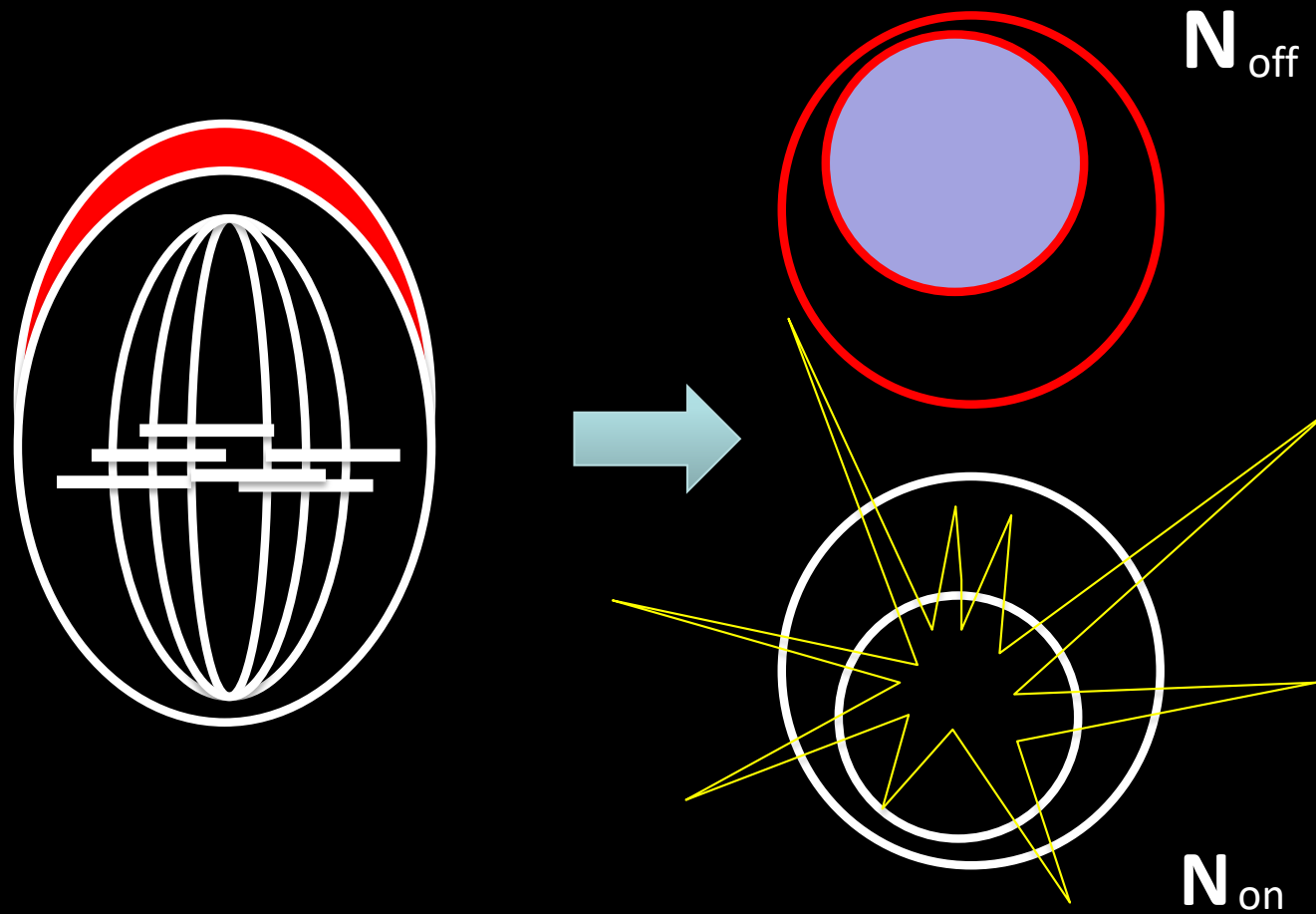


Knoblich (1995)

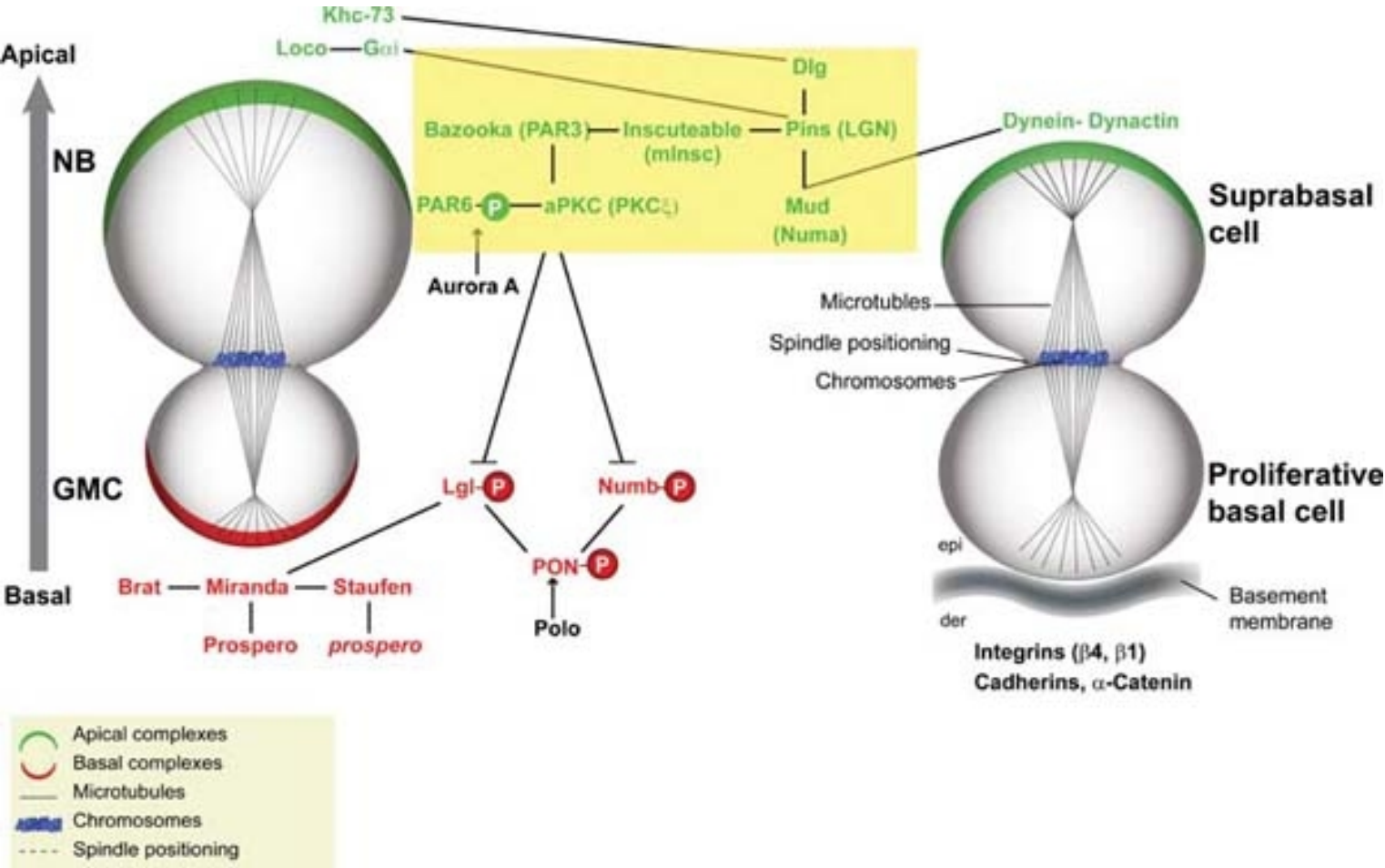
# Binary cell decisions and asymmetric cell divisions



# Binary cell decisions and asymmetric cell divisions



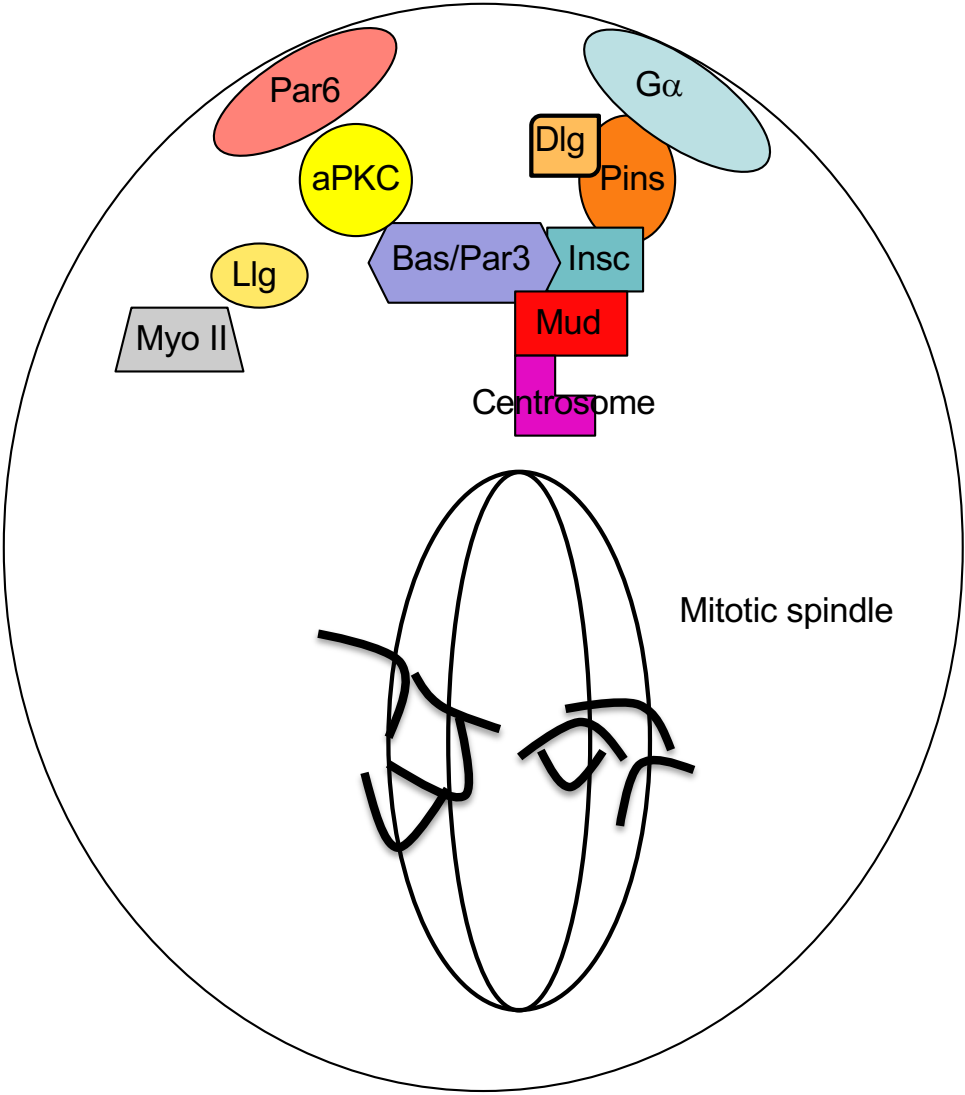
# Neuroblast asymmetric cell division



Baker NE, Brown NL. All in the family: proneural bHLH genes and neuronal diversity. (2018) Development;145.



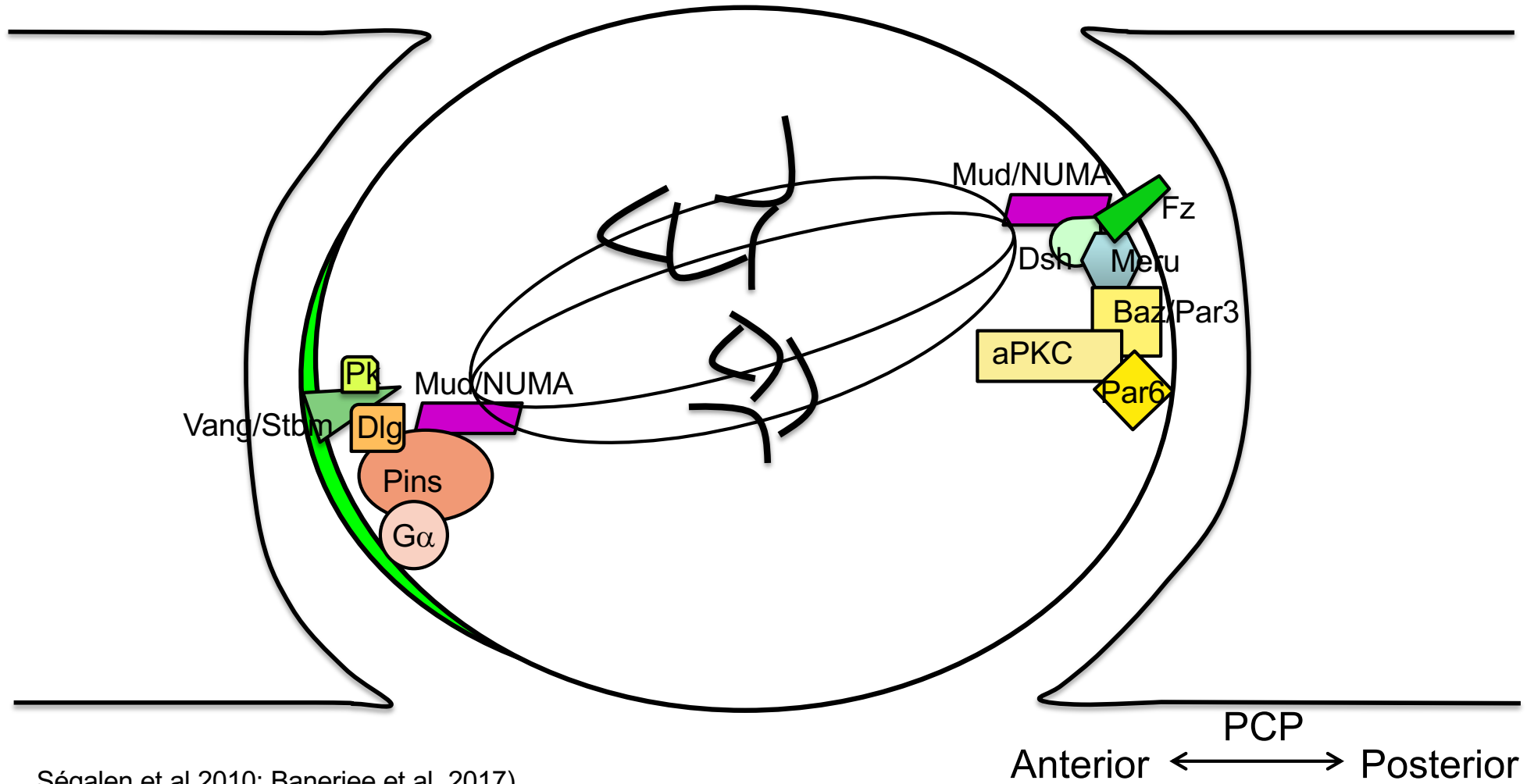
# Neuroblast asymmetric cell division



# SOP asymmetric cell division

Asymmetry is induced via the planar cell polarity (PCP) pathway

Posterior and anterior complexes control fate determinants segregation and spindle orientation



Ségalen et al 2010; Banerjee et al. 2017)

# The Peripheral Nervous System

# Sensory organs of the notum

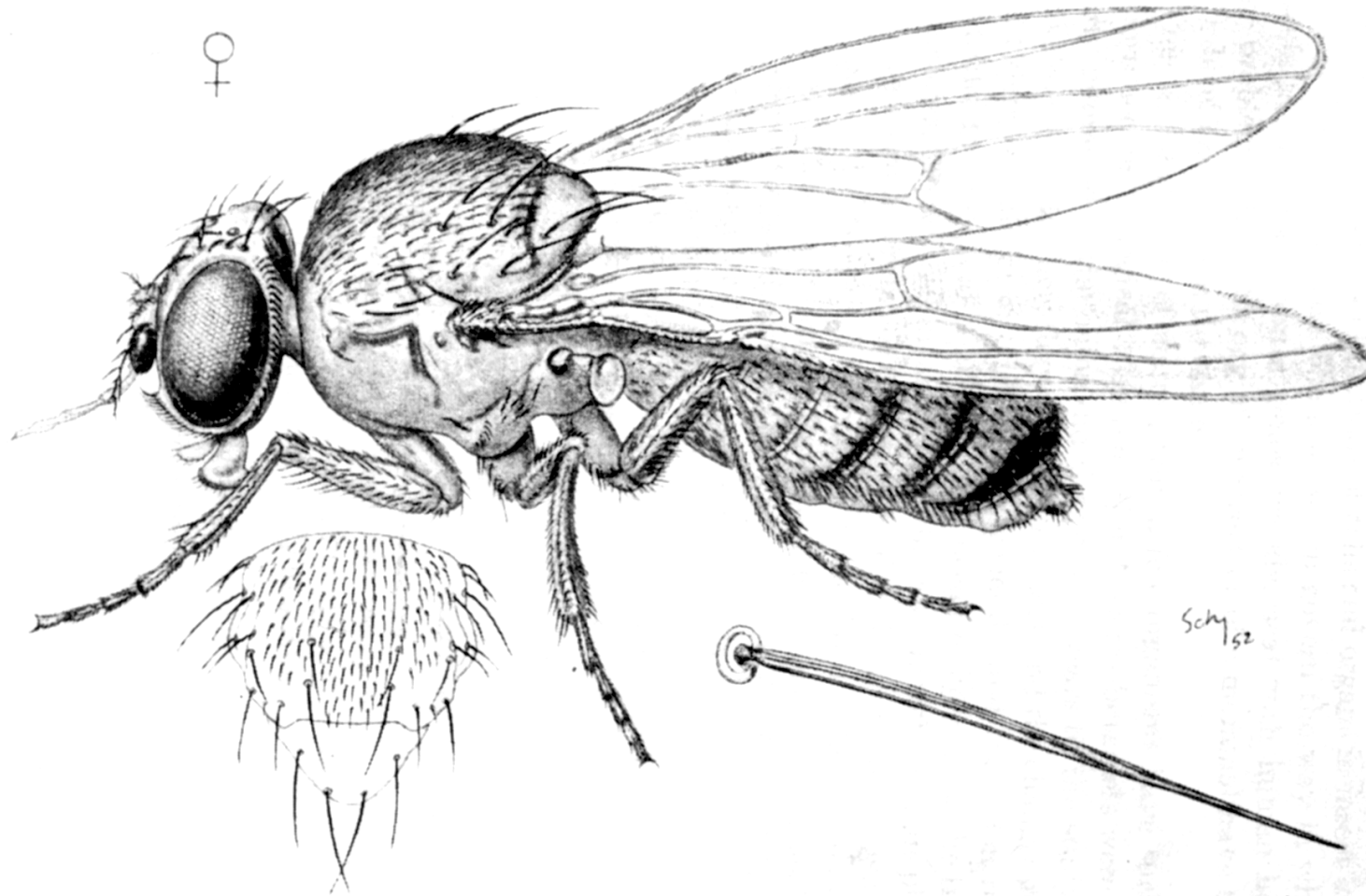
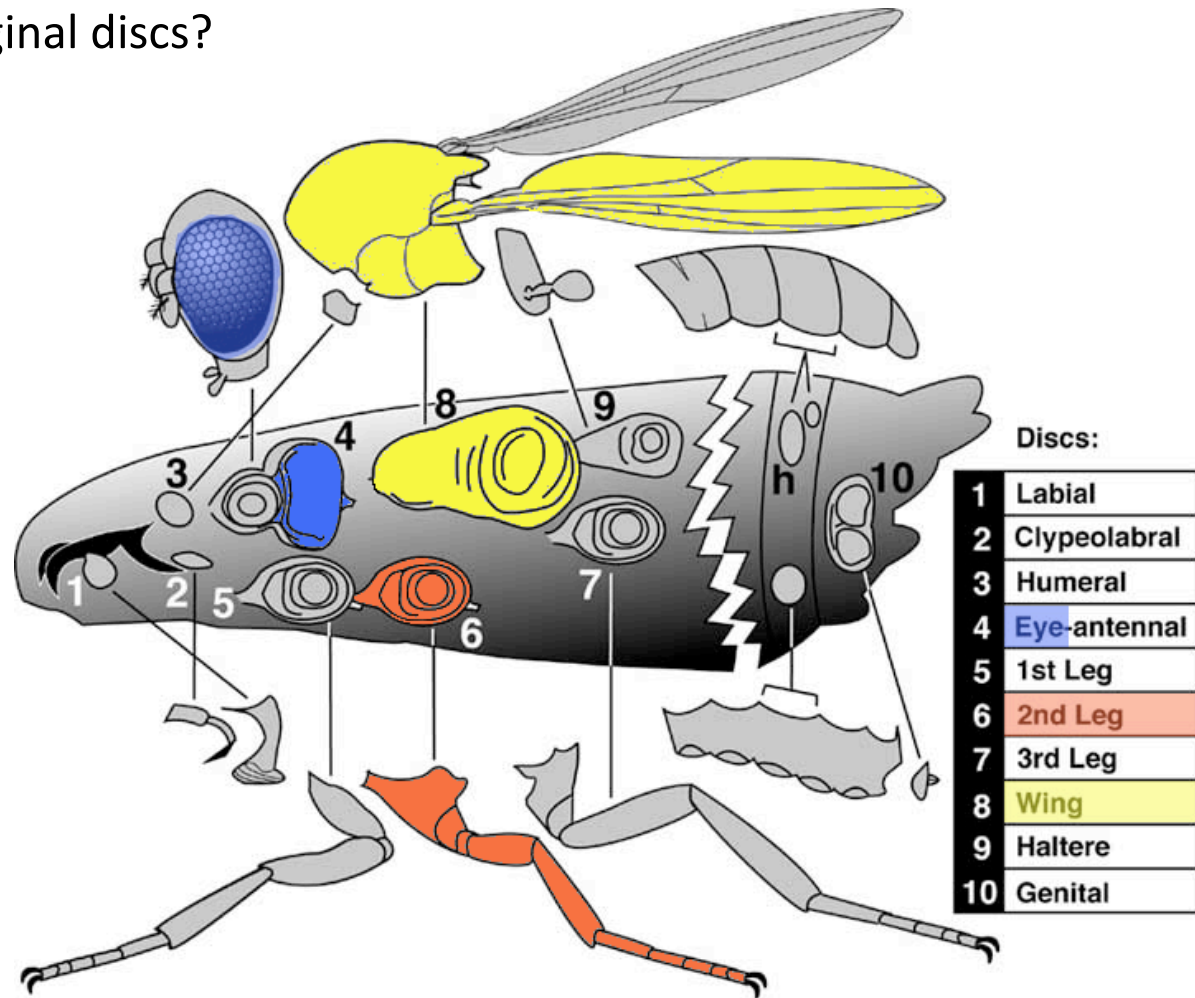


FIG. 1. *Drosophila melanogaster*. Above, side view, slightly oblique, of a female. Below, left, dorsal view of the thorax showing pattern of bristles and hairs. Below, right, a bristle in its socket.

# Sensory organs of the notum

Selection of the SOP/pl cell

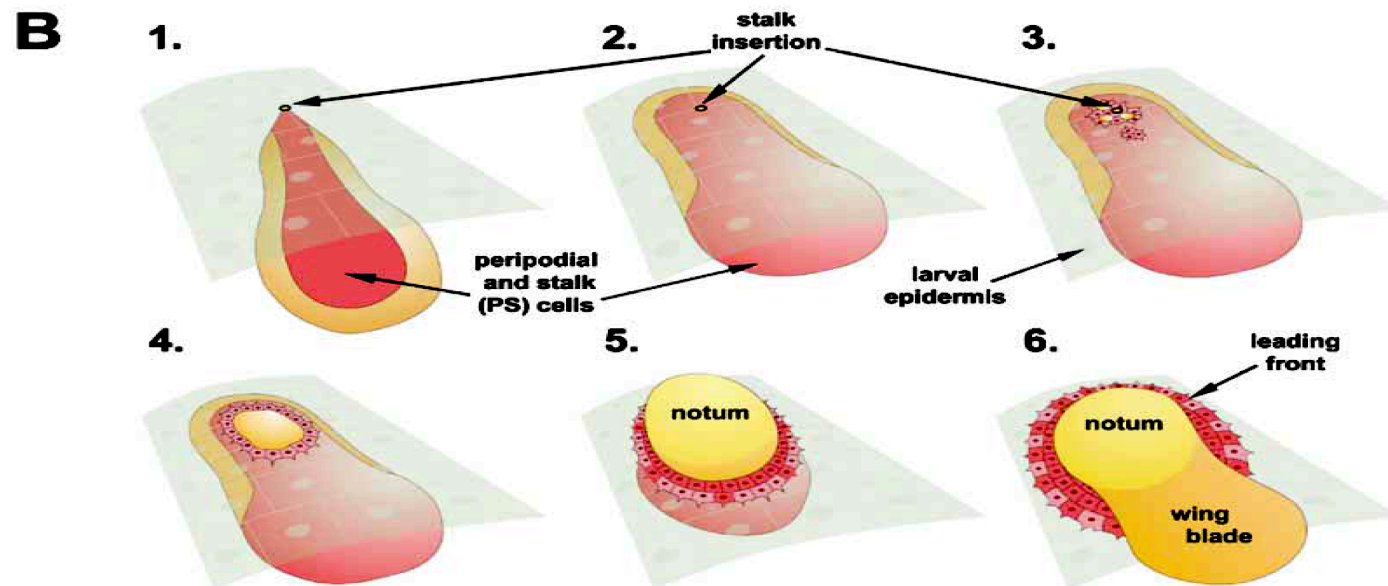
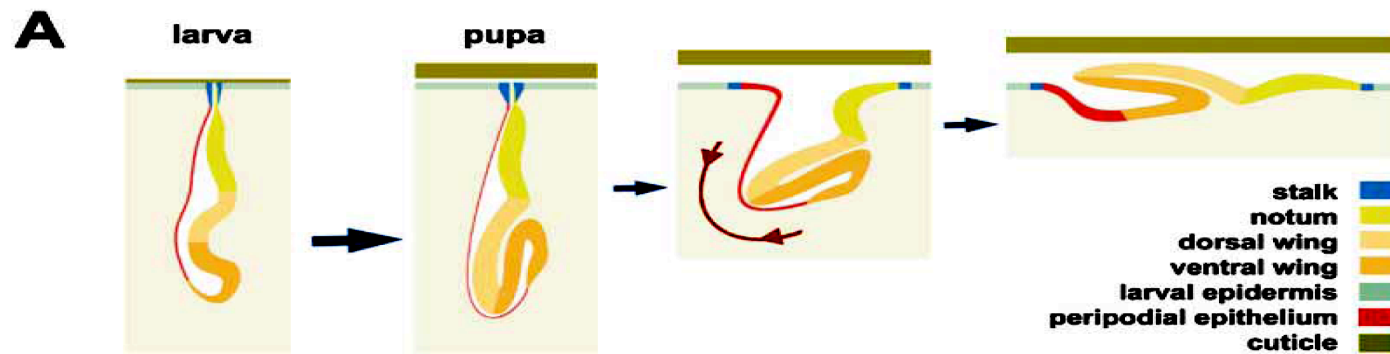
What are the imaginal discs?



# Sensory organs of the notum

Selection of the SOP/pl cell

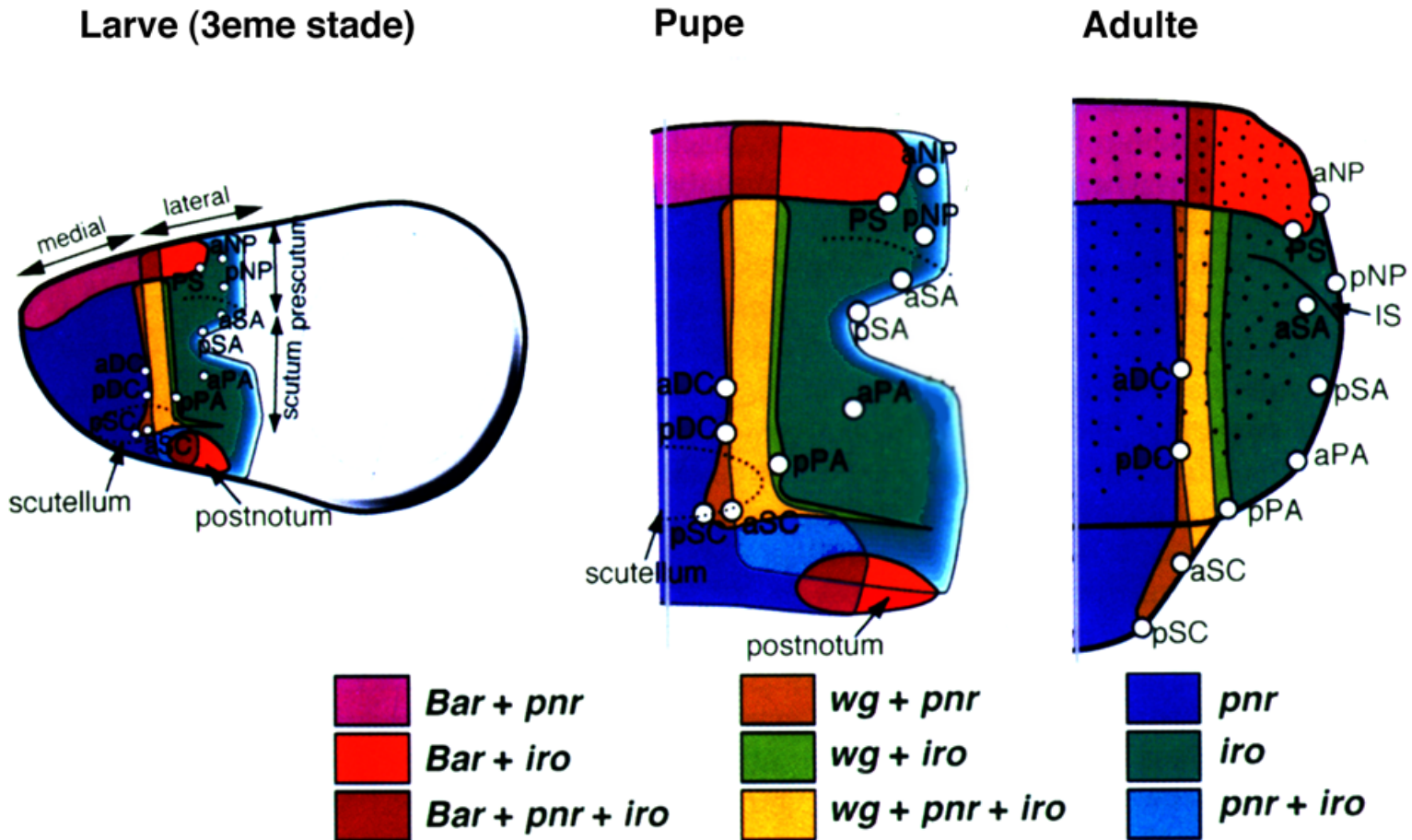
Wing discs eversion during metamorphosis



# Sensory organs of the notum

Selection of the SOP/pl cell

Prepattern genes



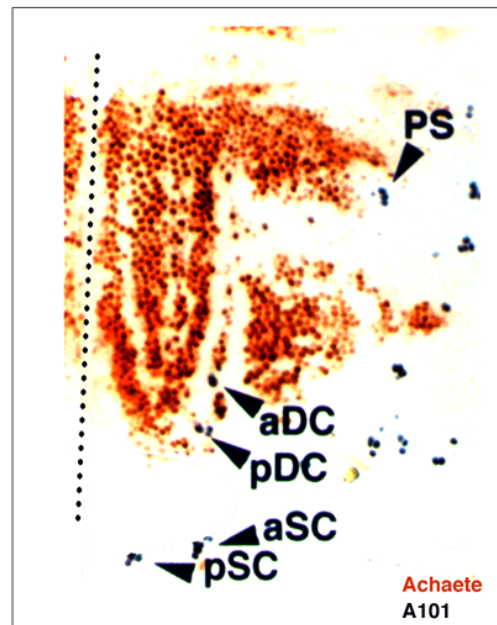
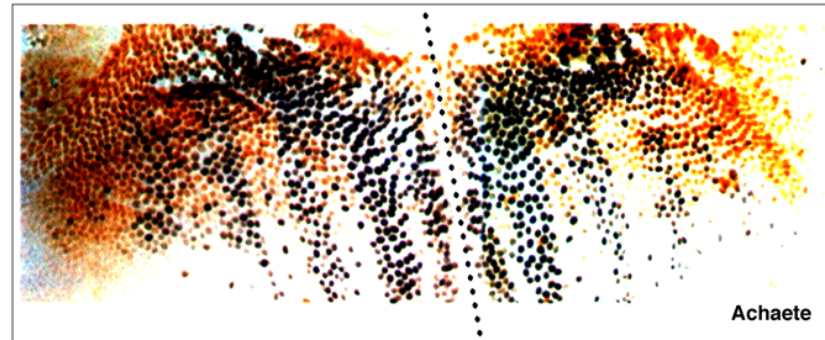


# Sensory organs of the notum

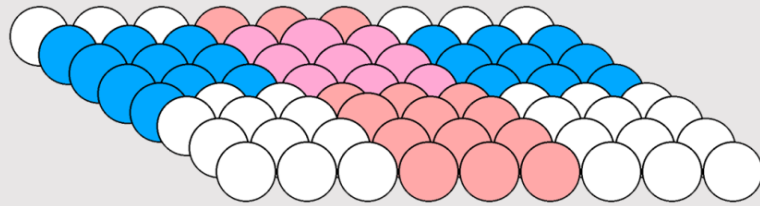
Selection of the SOP/pl cell

Proneural genes

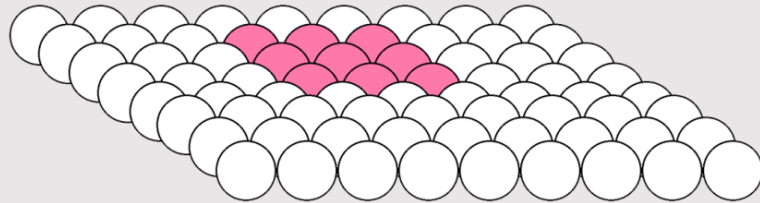
Expression d'achaete dans le notum (8 hrs APF)



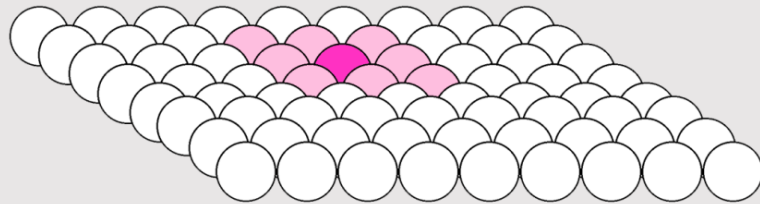
Sato et al., (1999) Development 126; 1457-1466



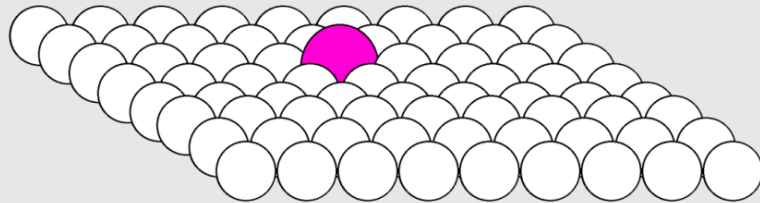
Pre-pattern



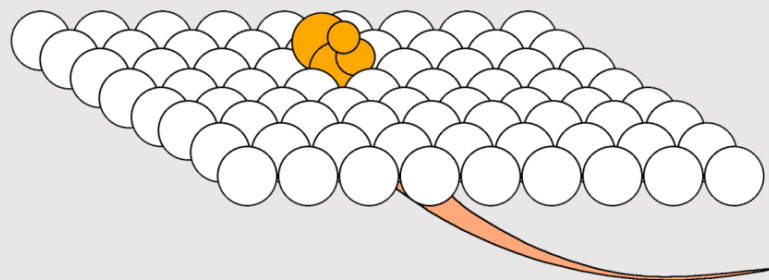
Formation d'un groupe proneural  
gènes proneuraux



Sélection d'une SOP  
gènes neurogéniques

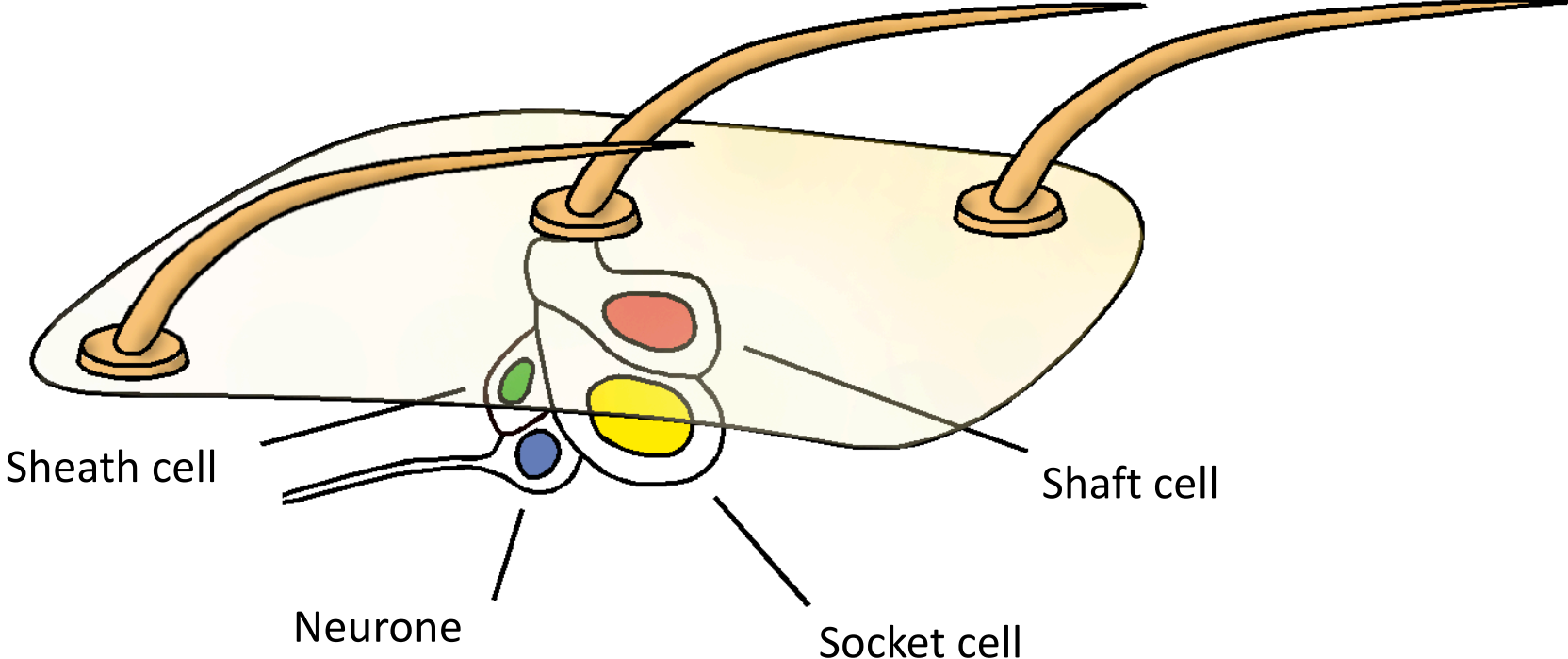


Spécification de la SOP  
gènes d'identité cellulaire



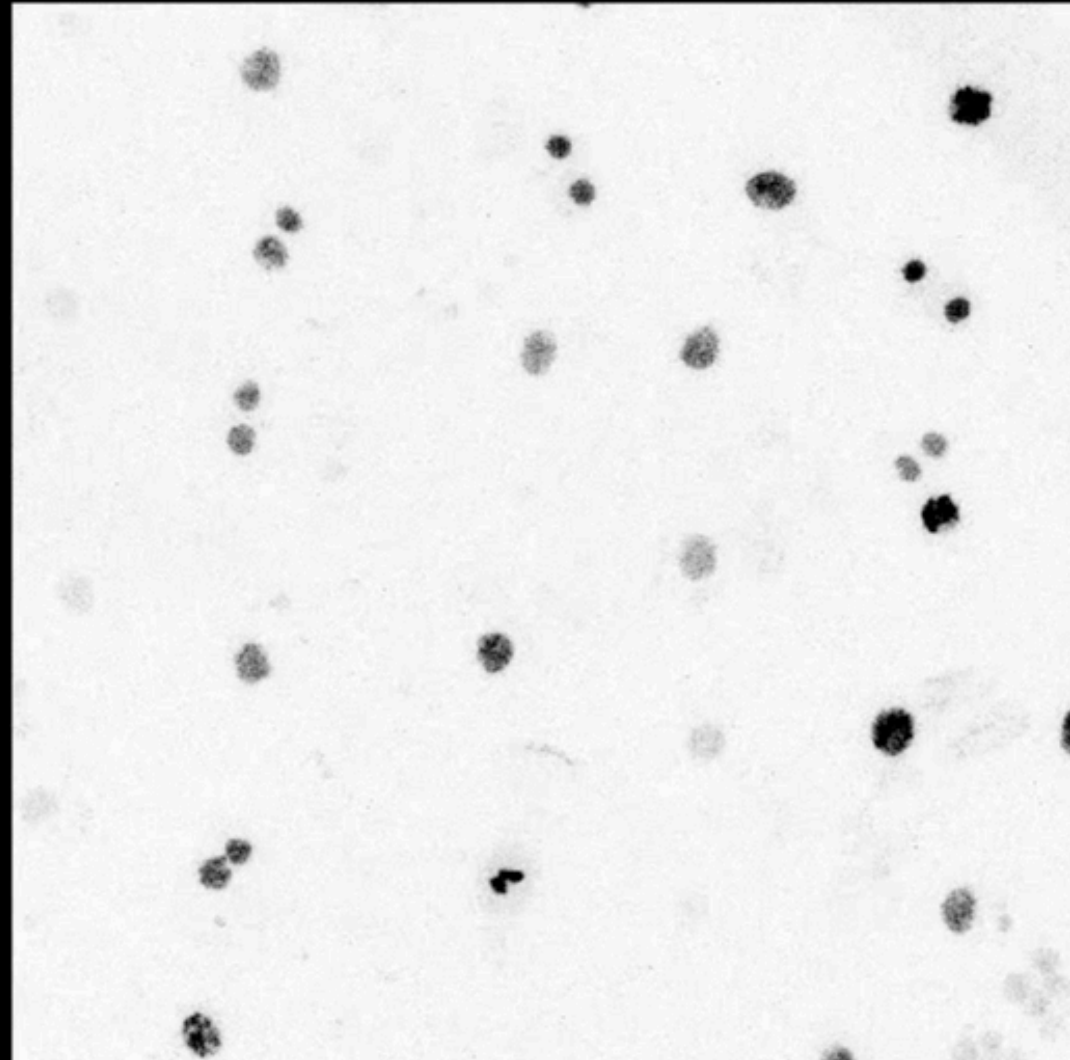
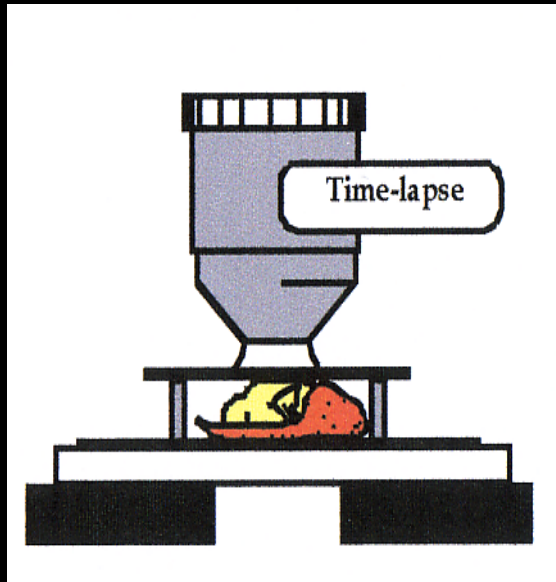
Division de la SOP et  
différenciation de l'organe sensoriel  
gènes d'identité cellulaire

# Sensory organs of the notum



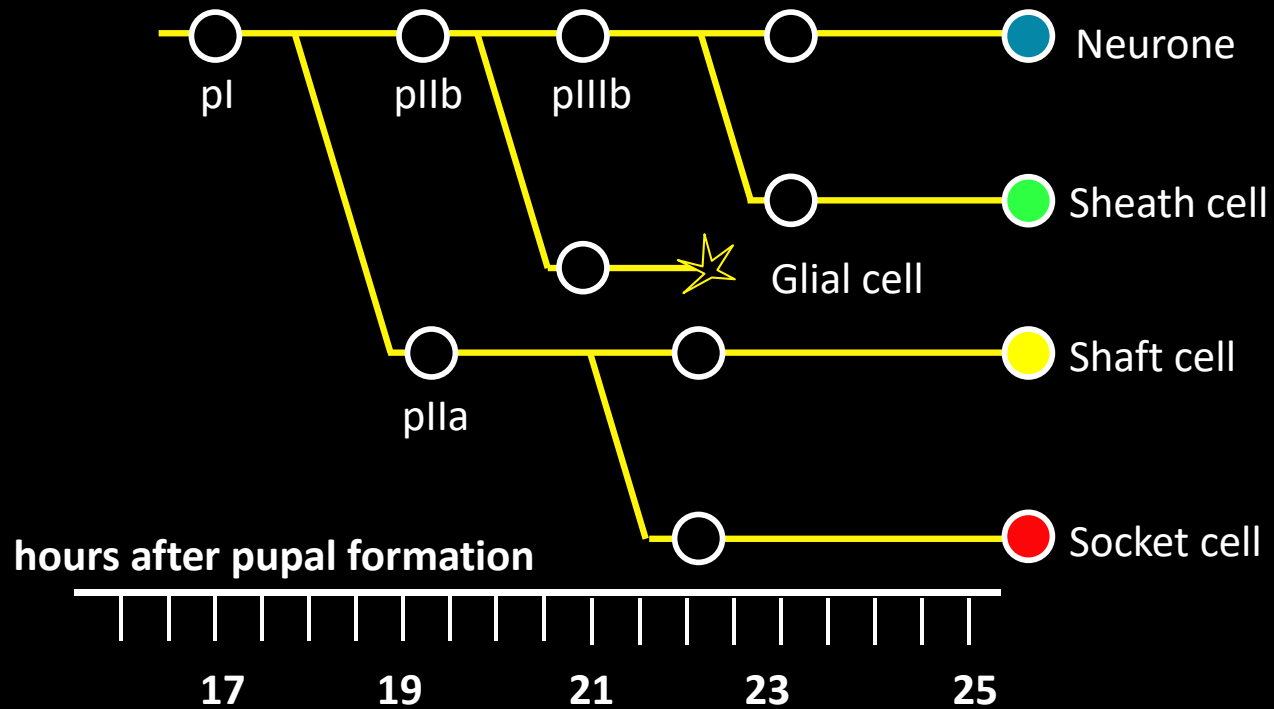
# Sensory organs of the notum

The bristle lineage



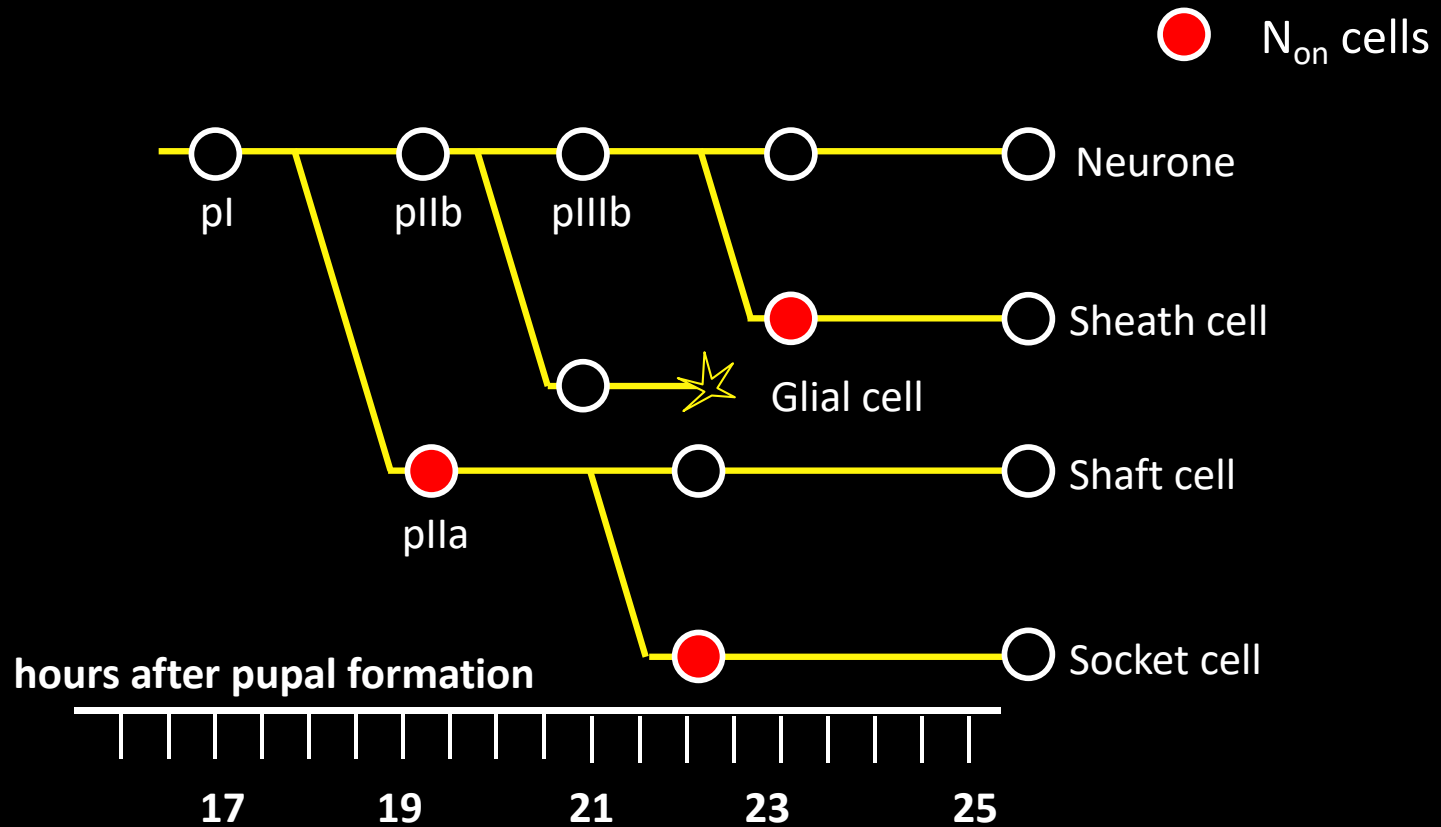
*neu-Gal4; UAS-H2B::YFP*

## *Drosophila* mechanosensory organs



The bristle lineage ( a model system to study cell diversification)

# *Drosophila* mechanosensory organs

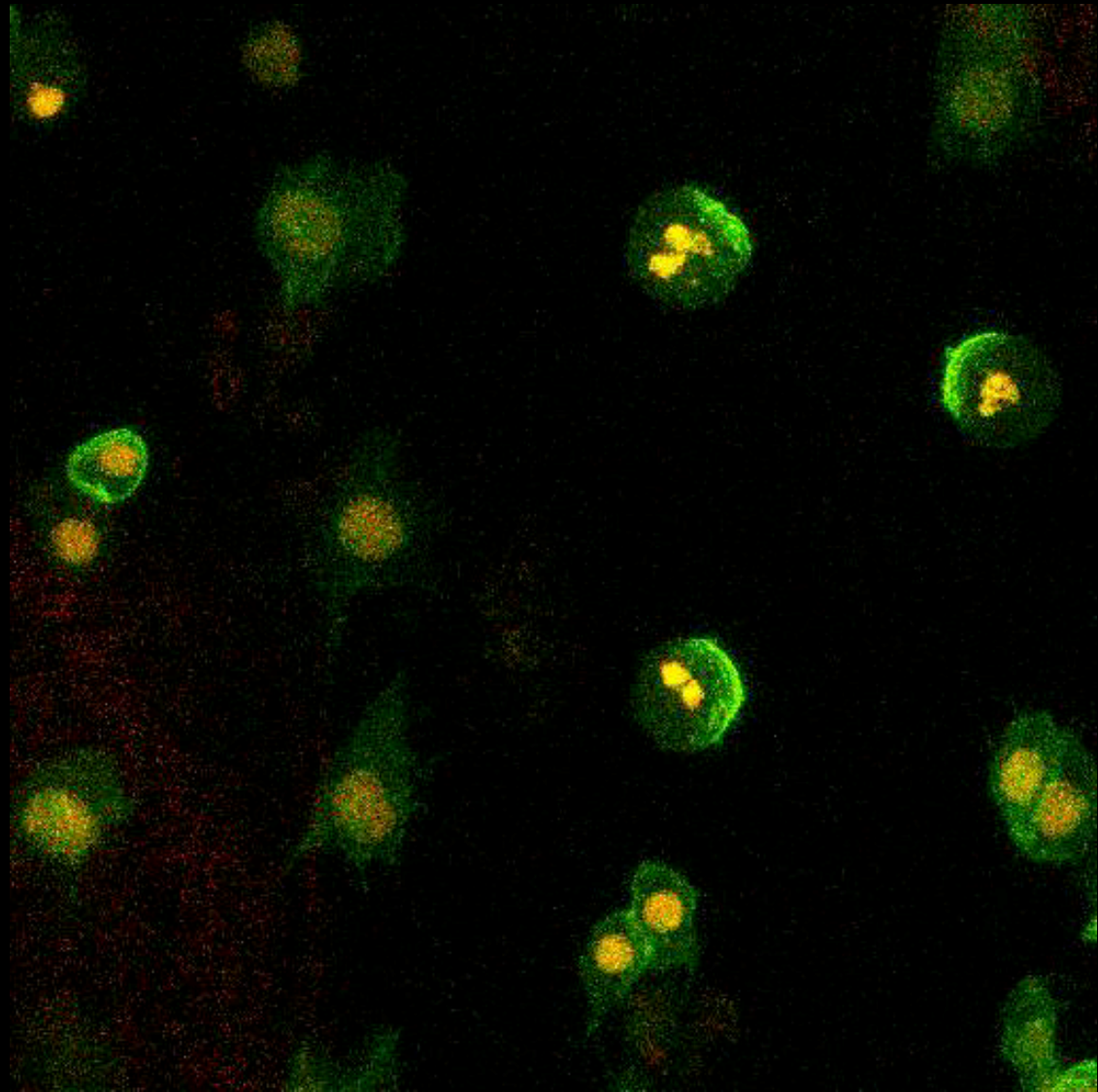


Binary cell fate decisions: The Notch pathway



# Sensory organs of the notum

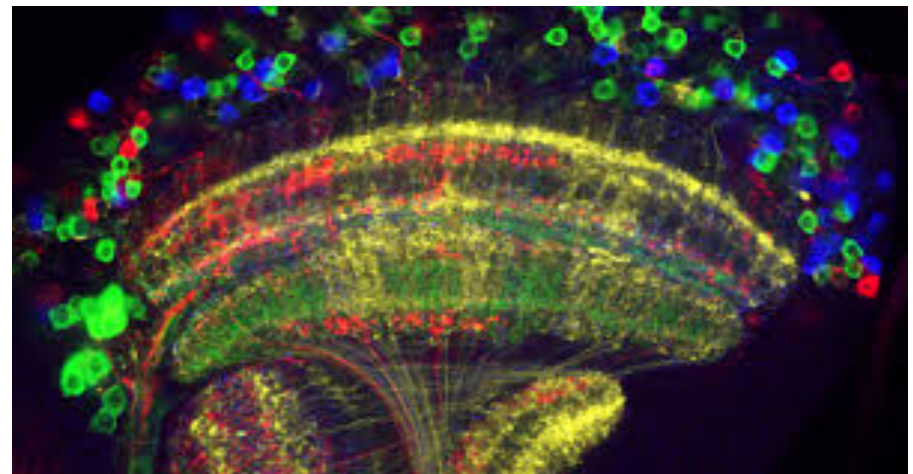
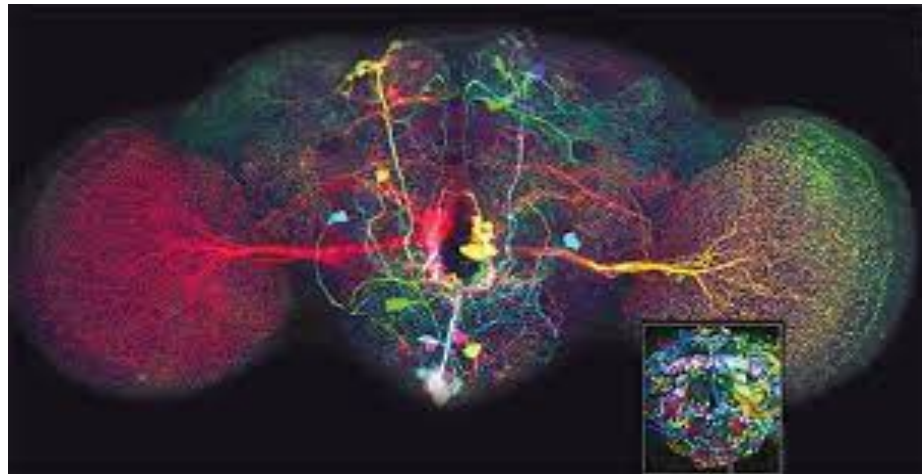
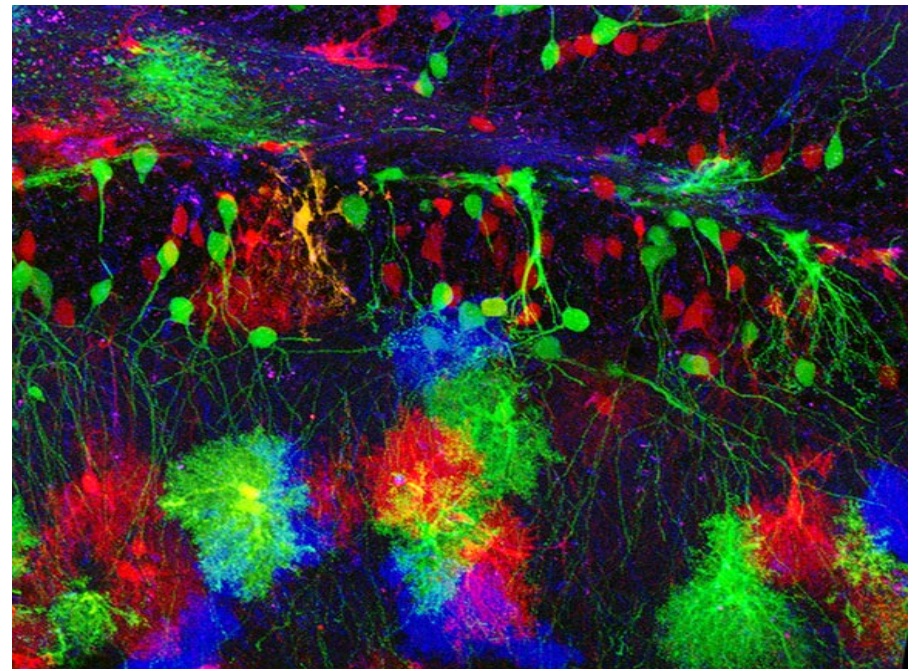
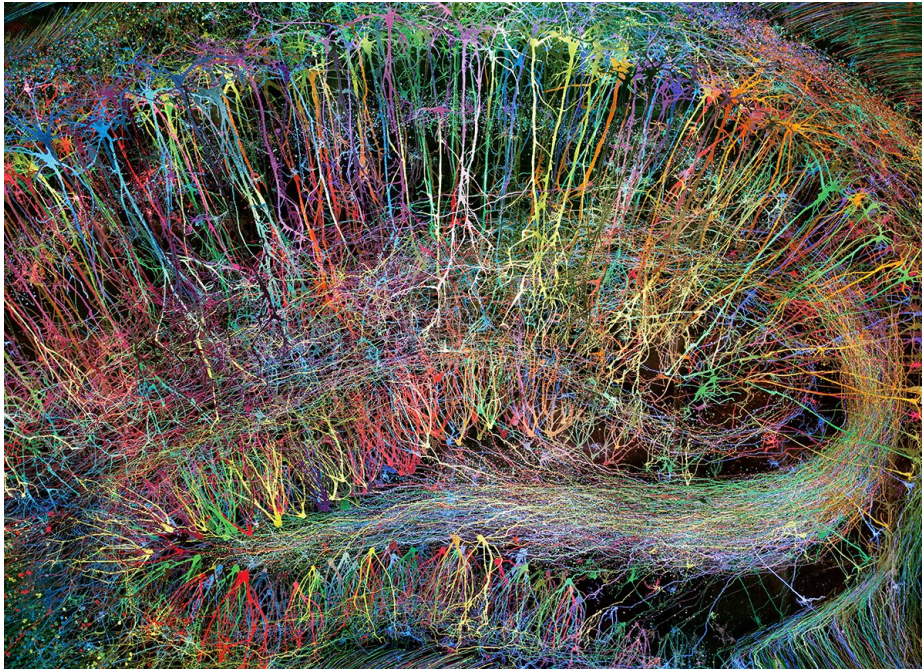
Asymmetric cell divisions



HisH2B-RFP Pon-GFP



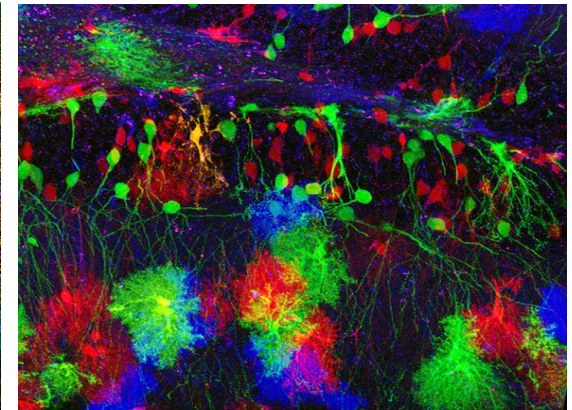
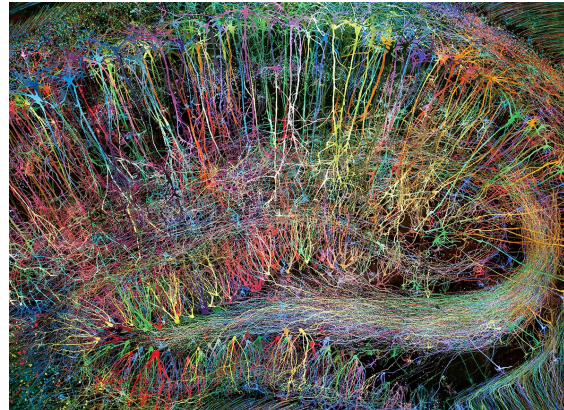
# Origin of neuron diversity



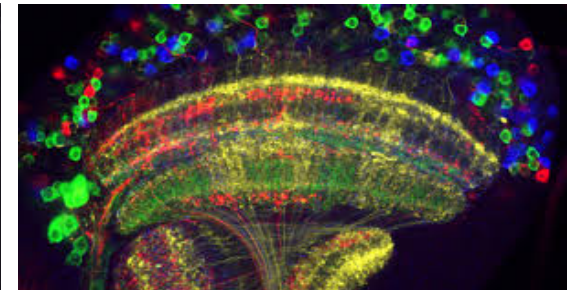
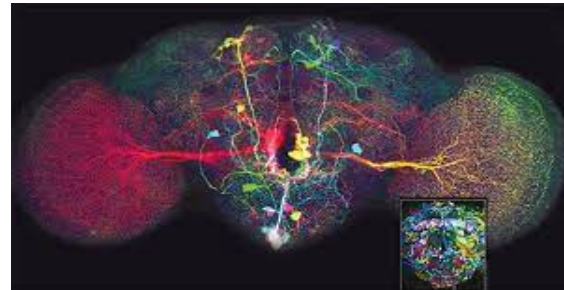


# Origin of neuron diversity

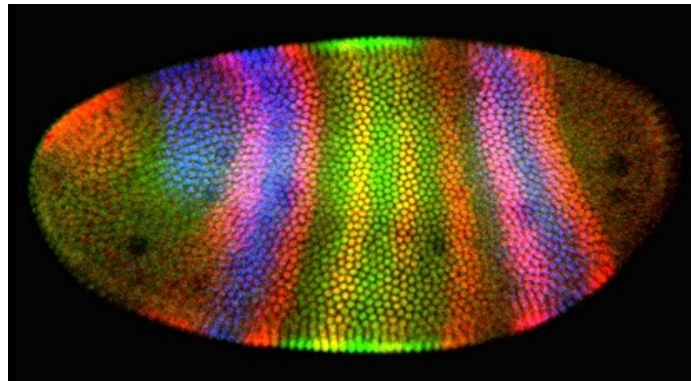
Spatial clues



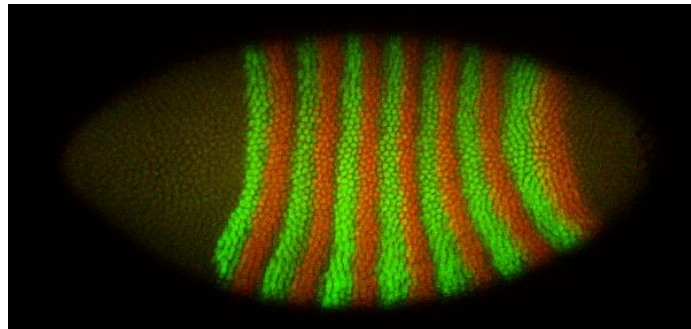
Temporal clues



# Spatial regulation of neuroblast identity

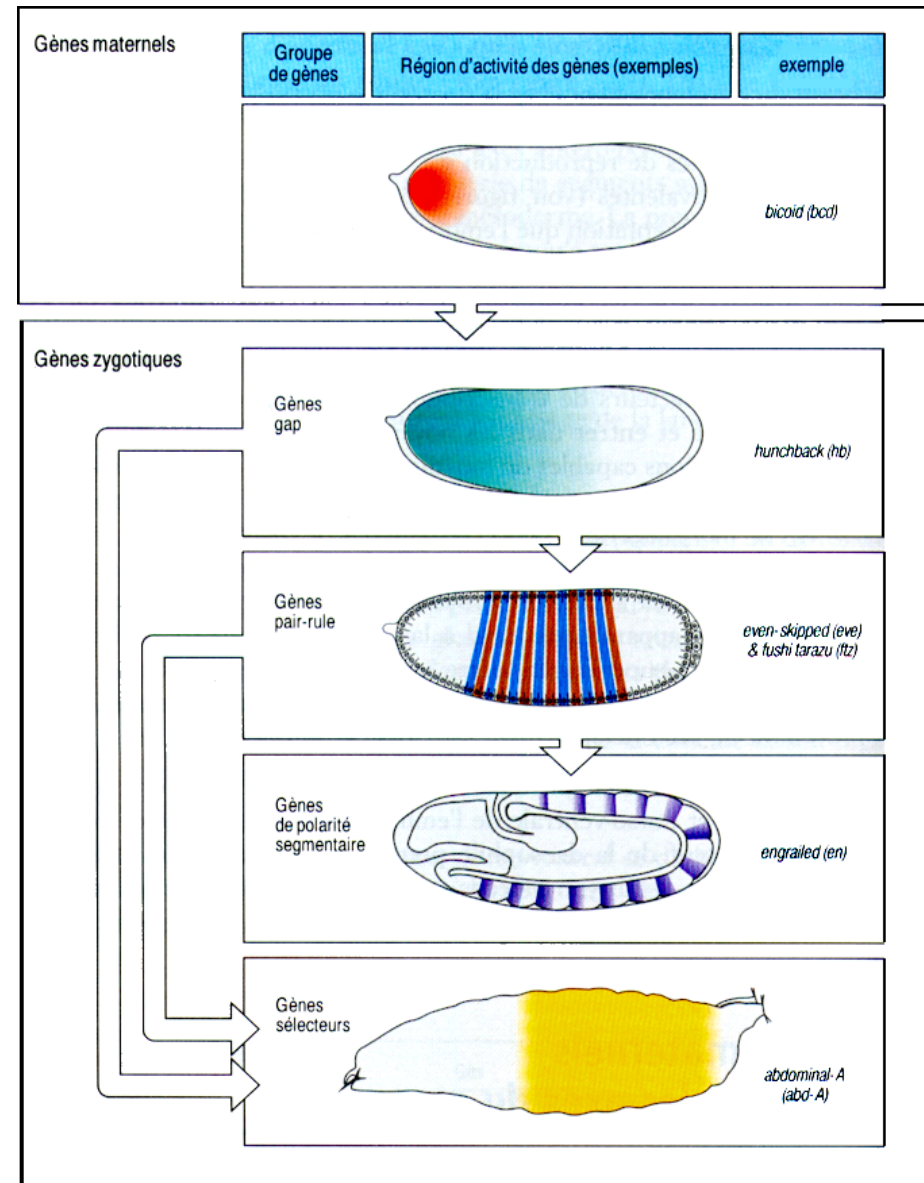


hairy krüppel giant



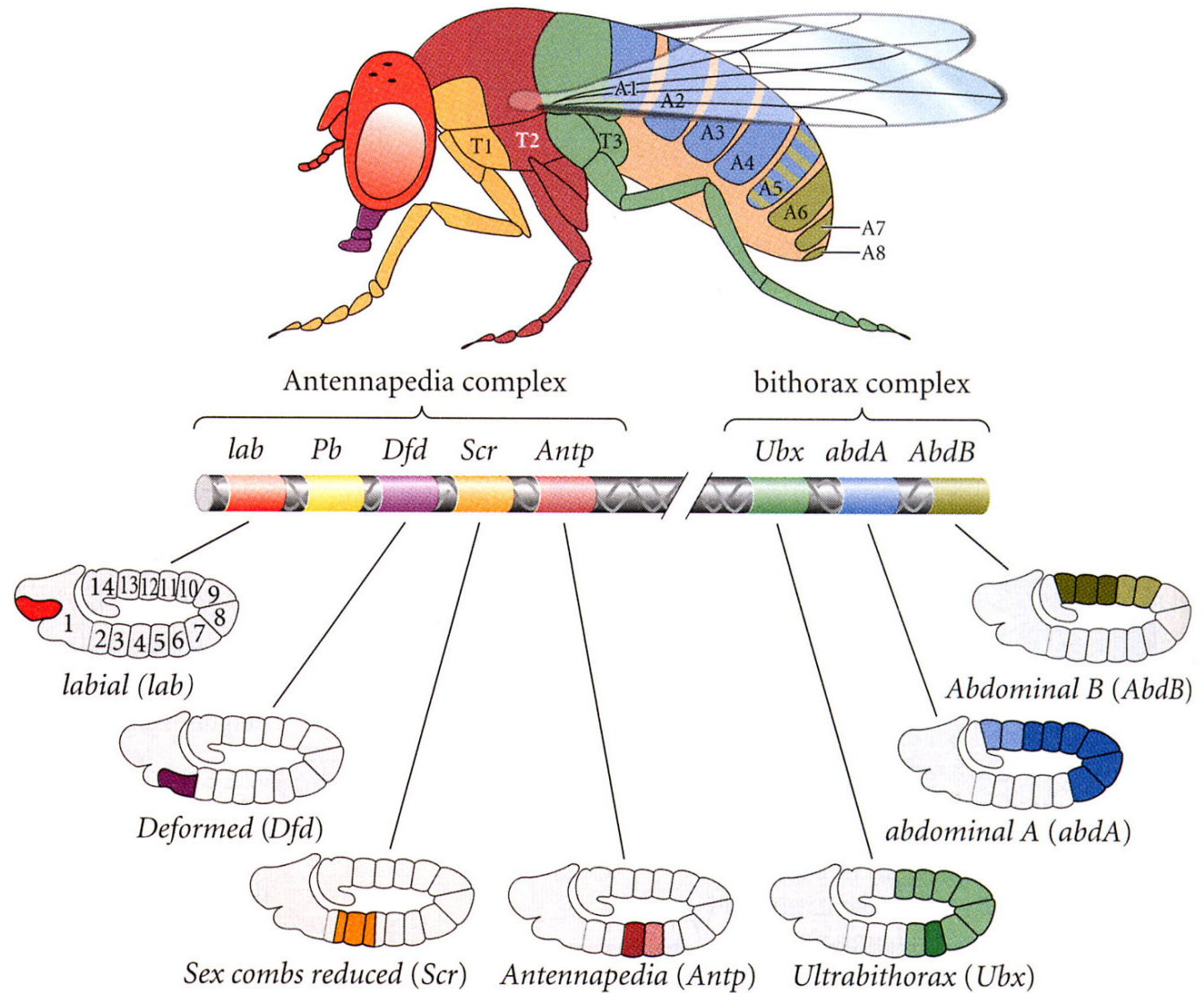
even-skipped fushi tarazu

Genes patterning following antero-posterior axis



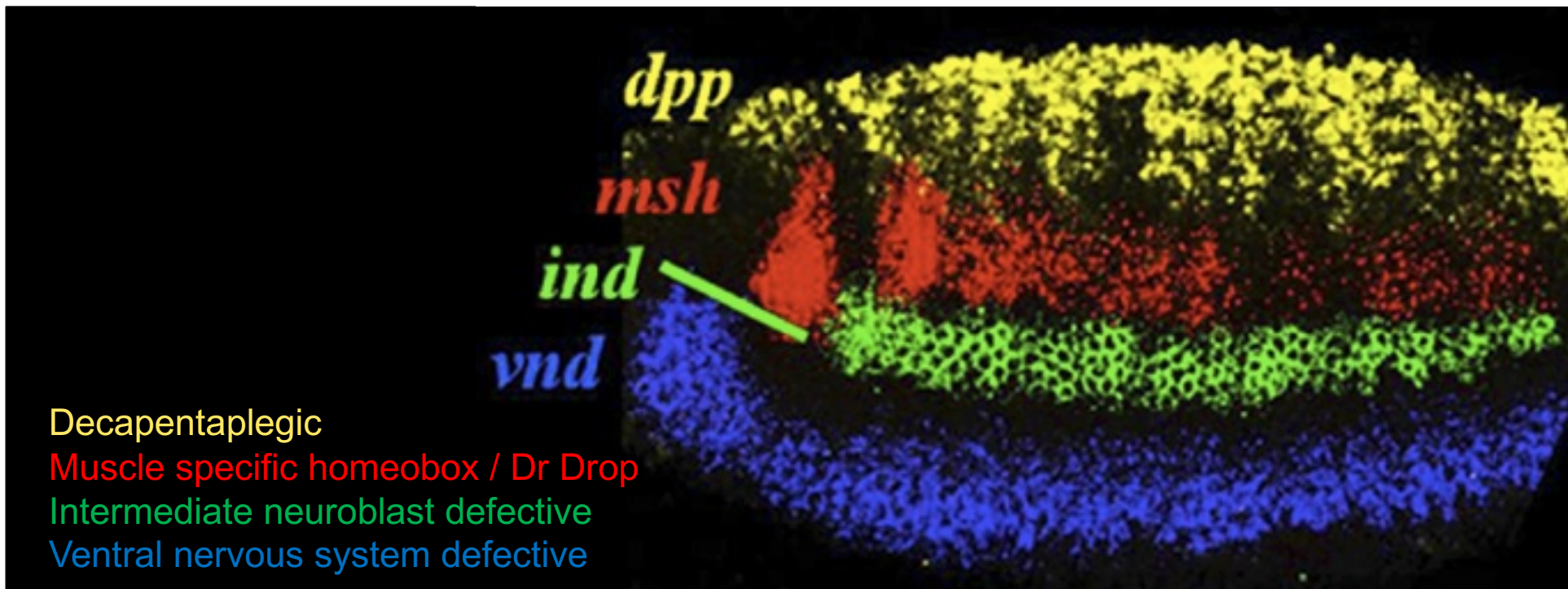
# Spatial regulation of neuroblast identity

## Hox genes





## Spatial regulation of neuroblast identity

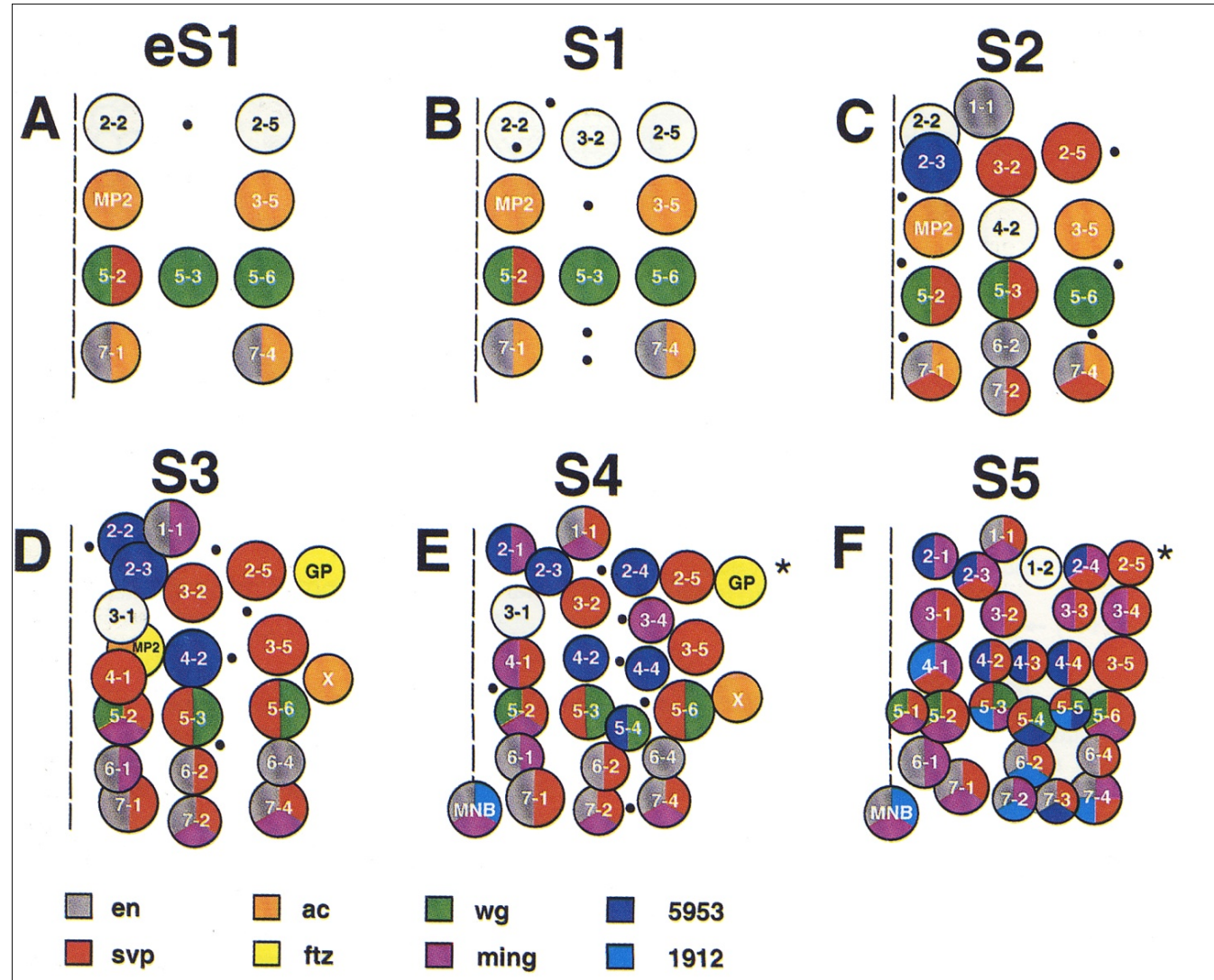


Mizutani et al. (2006)

Genes patterning following medio-lateral axis

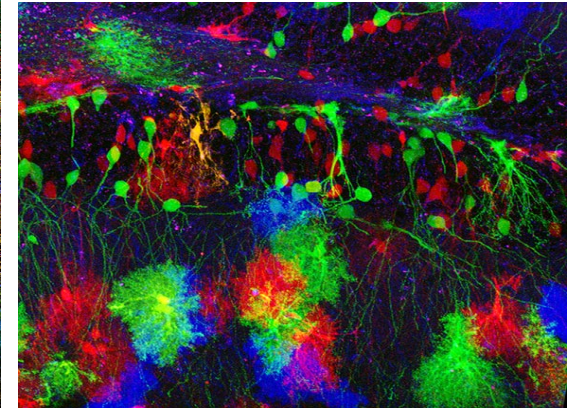
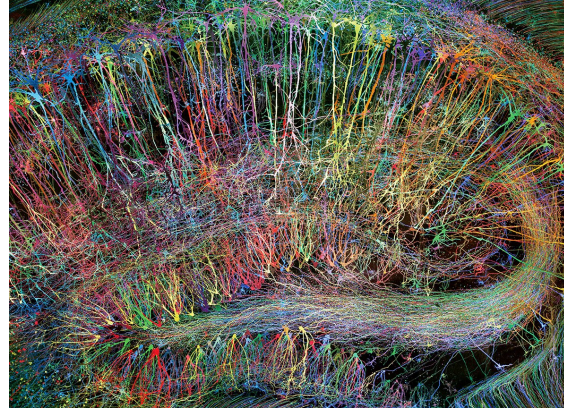
# Spatial regulation of neuroblast identity

## Neuroblasts in the embryonic ventral nerve cord

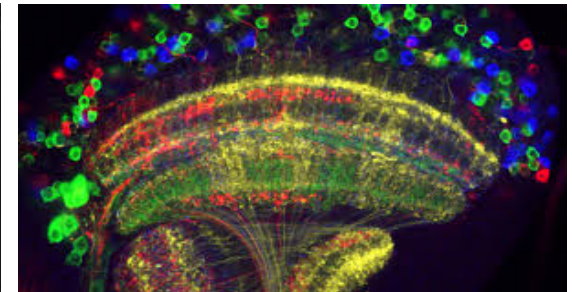
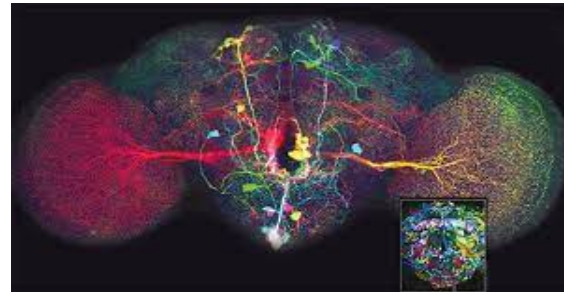


# Origin of neuron diversity

Spatial clues



Temporal clues





## **Temporal patterning in neurogenesis**

**A key challenge during neural development is to coordinate the birth and specification of diverse neuronal and glial cell types.**

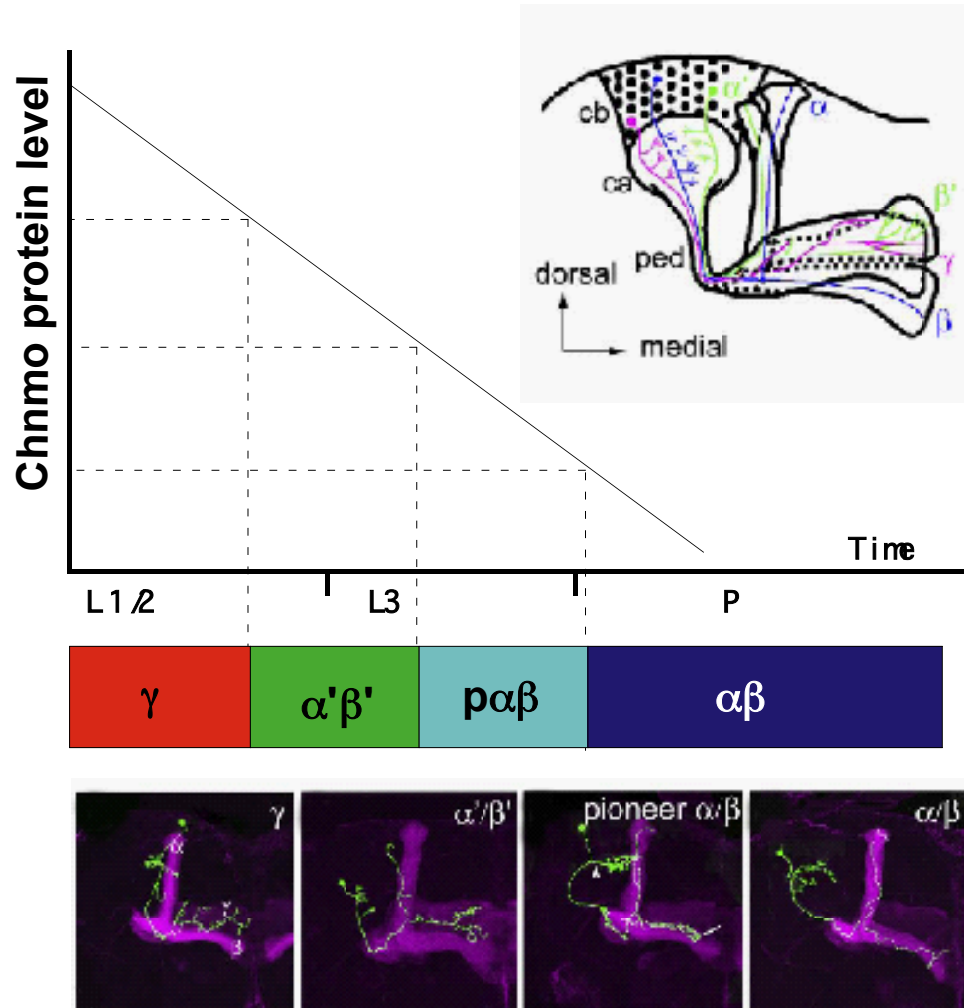
**Gradients of the *Drosophila*  
Chinmo BTB-Zinc Finger Protein  
Govern Neuronal Temporal Identity**

Sijun Zhu,<sup>1,2,5</sup> Suewei Lin,<sup>3</sup> Chih-Fei Kao,<sup>3</sup> Takeshi Awasaki,<sup>3</sup> Ann-Shyn Chiang,<sup>4</sup> and Tzumin Lee<sup>1,3,\*</sup>

Cell 127, 409–422, October 20, 2006

# Origin of neuroblast diversity

Temporal control of neuroblast identity



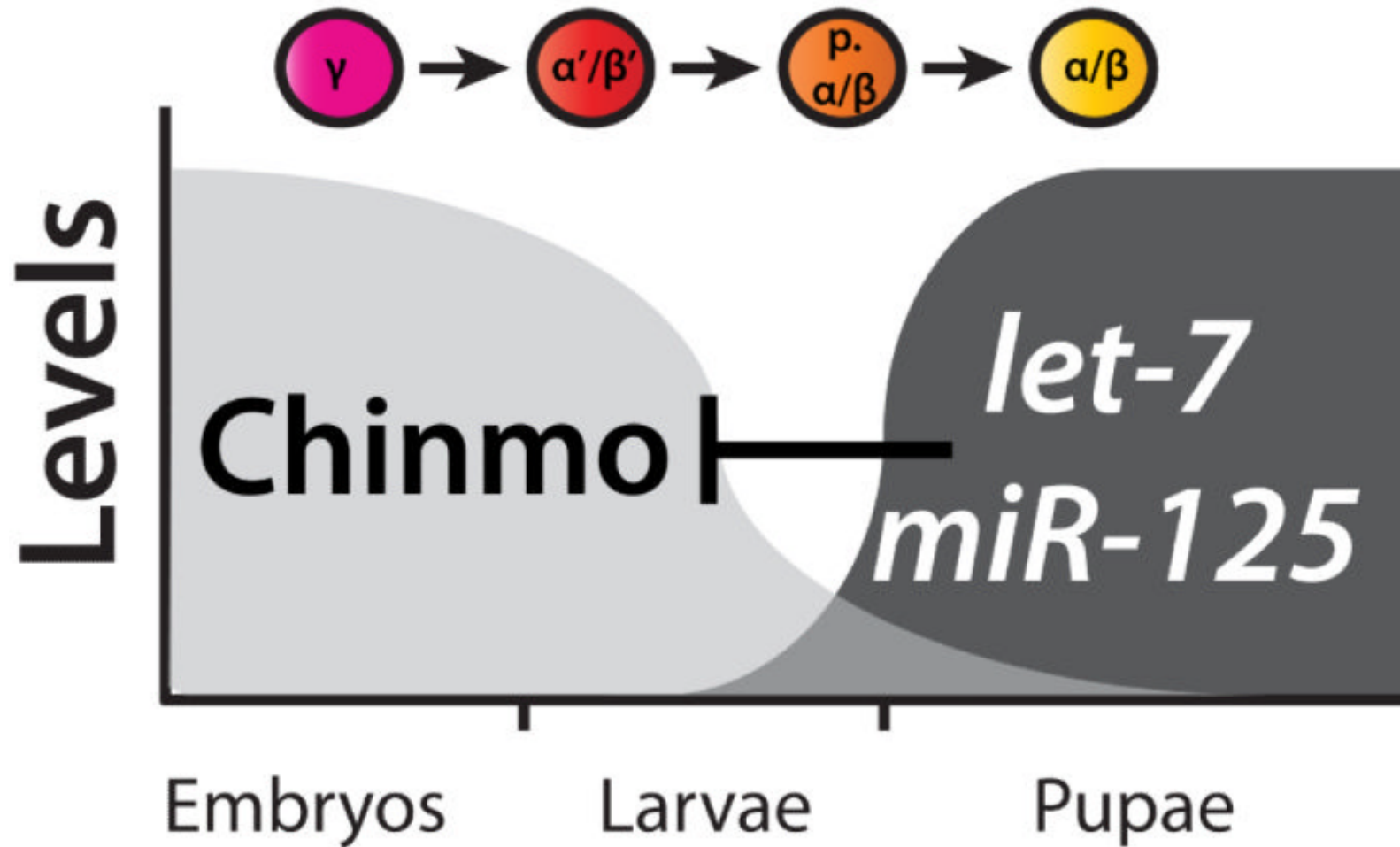
Zhu et al. (2006)

Temporal gradient of the Chinmo protein governs neuronal temporal identity

# Origin of neuroblast diversity

Temporal control of neuroblast identity

Wu..Sokol (2012)



Temporal gradient of the Chinmo protein governs neuronal temporal identity

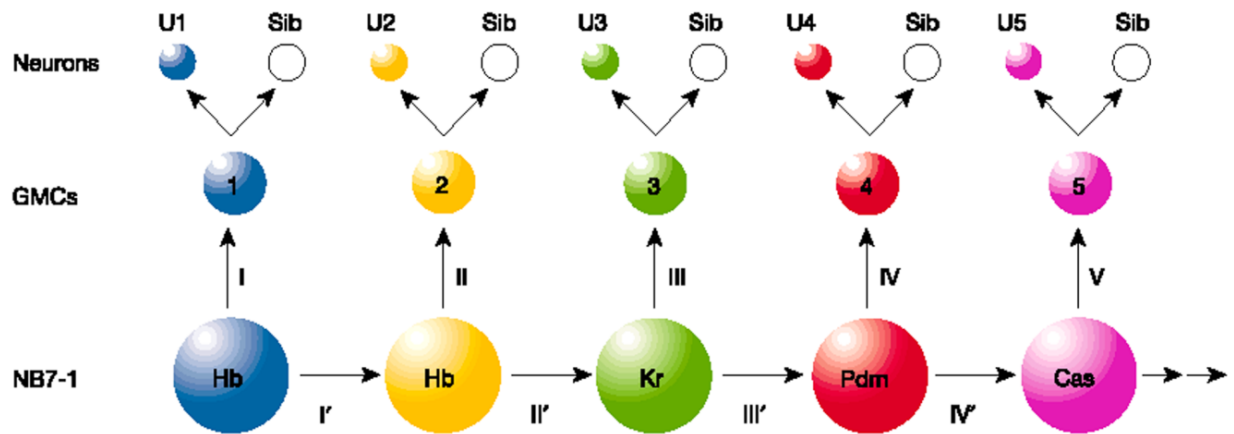
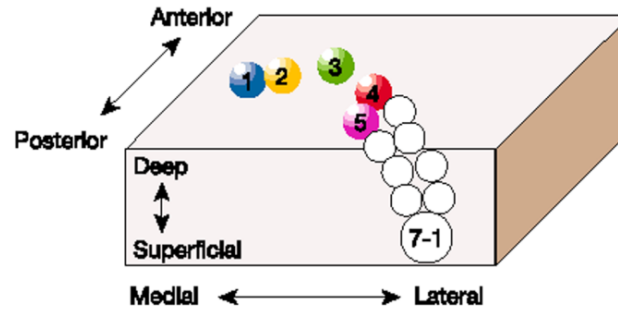
Cell, Vol. 106, 511–521, August 24, 2001, Copyright ©2001 by Cell Press

# ***Drosophila* Neuroblasts Sequentially Express Transcription Factors which Specify the Temporal Identity of Their Neuronal Progeny**

Takako Isshiki, Bret Pearson, Scott Holbrook,  
and Chris Q. Doe<sup>1</sup>

# Origin of neuroblast diversity

Temporal control of neuroblast identity

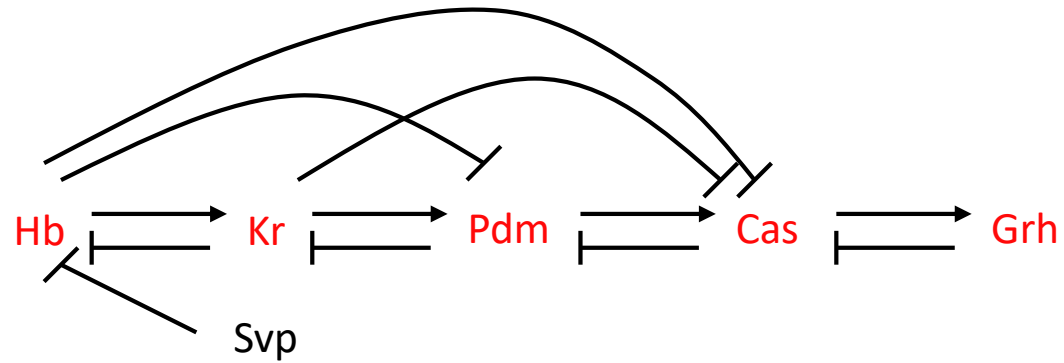


Doe (2003, 2005)

Multiple temporal identity transcription factors specify distinct neuronal fates within a single early competence window

# Origin of neuroblast diversity

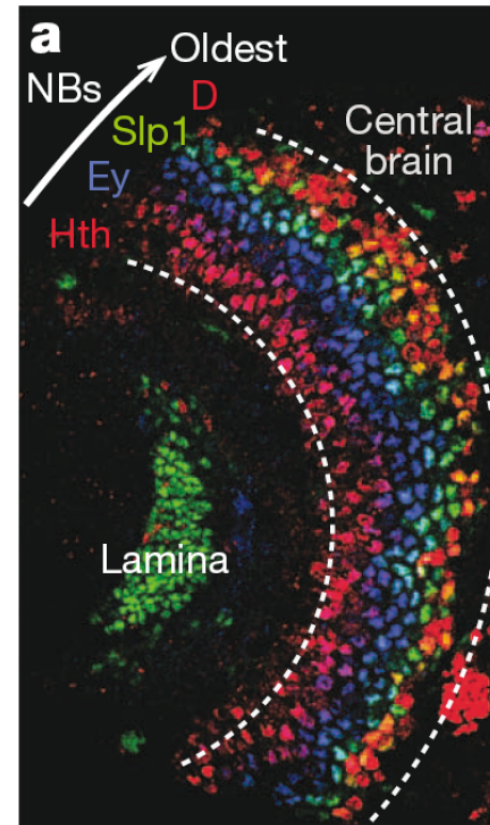
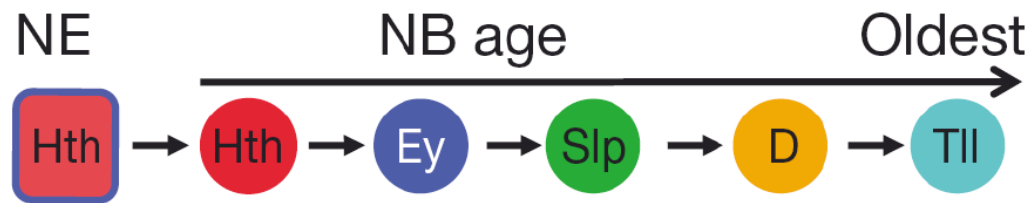
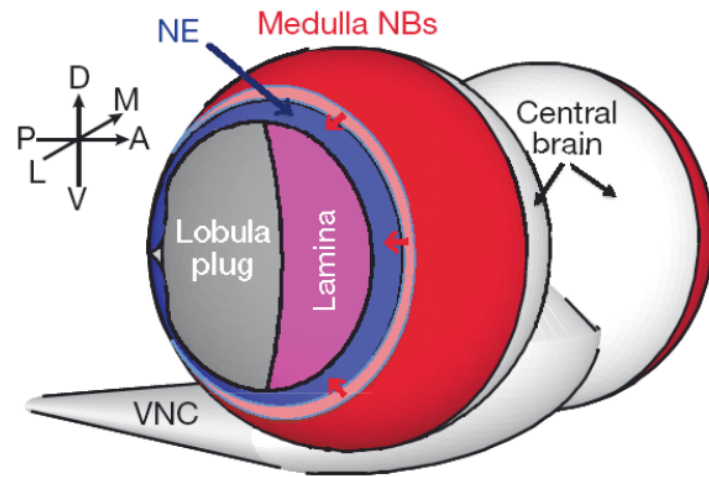
Temporal control of neuroblast identity



Transitions between TFS is controlled by feedforward and feedback repression cross-regulations

# Adult Neurogenesis

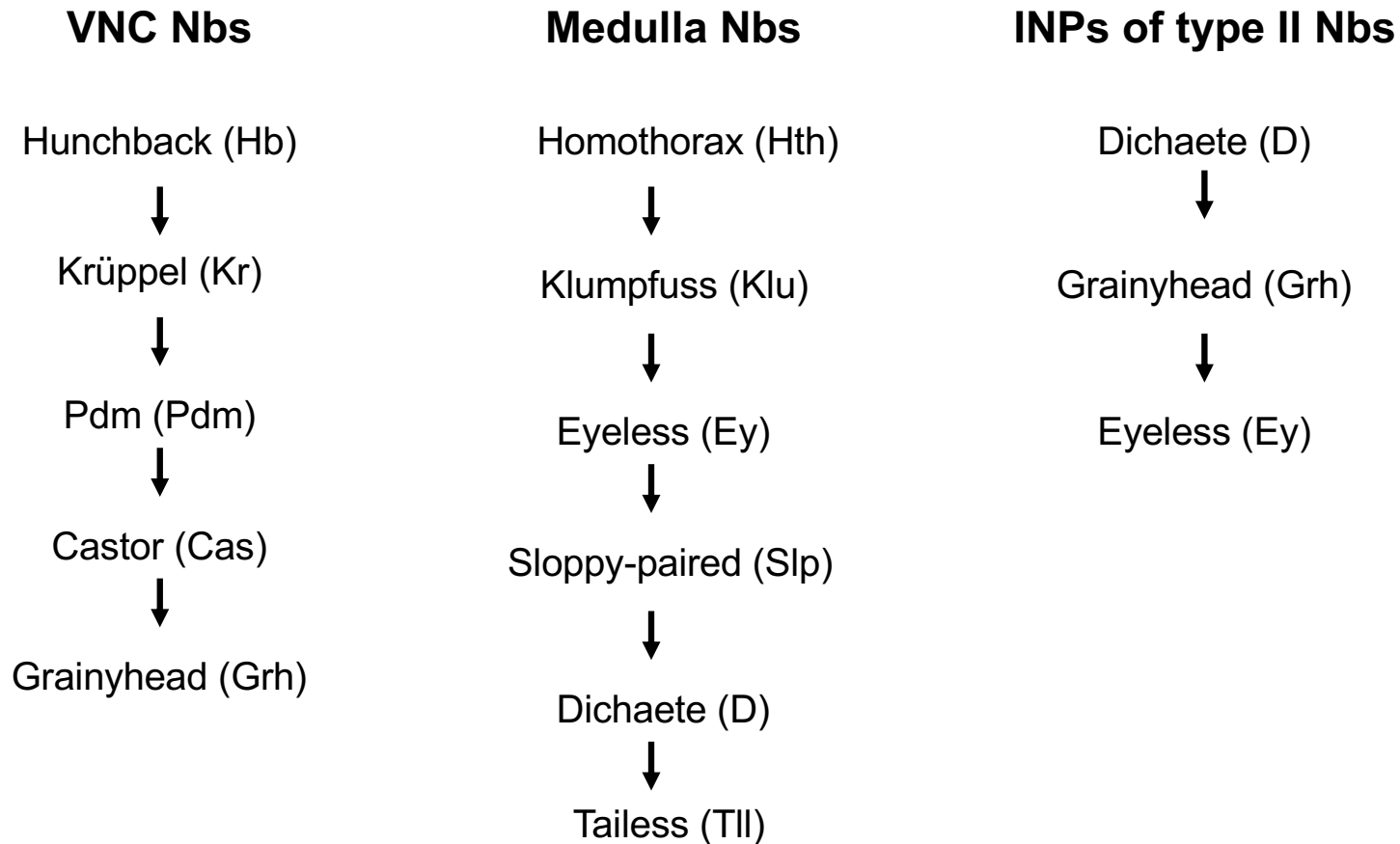
Temporal patterning of *Drosophila* medulla neuroblasts





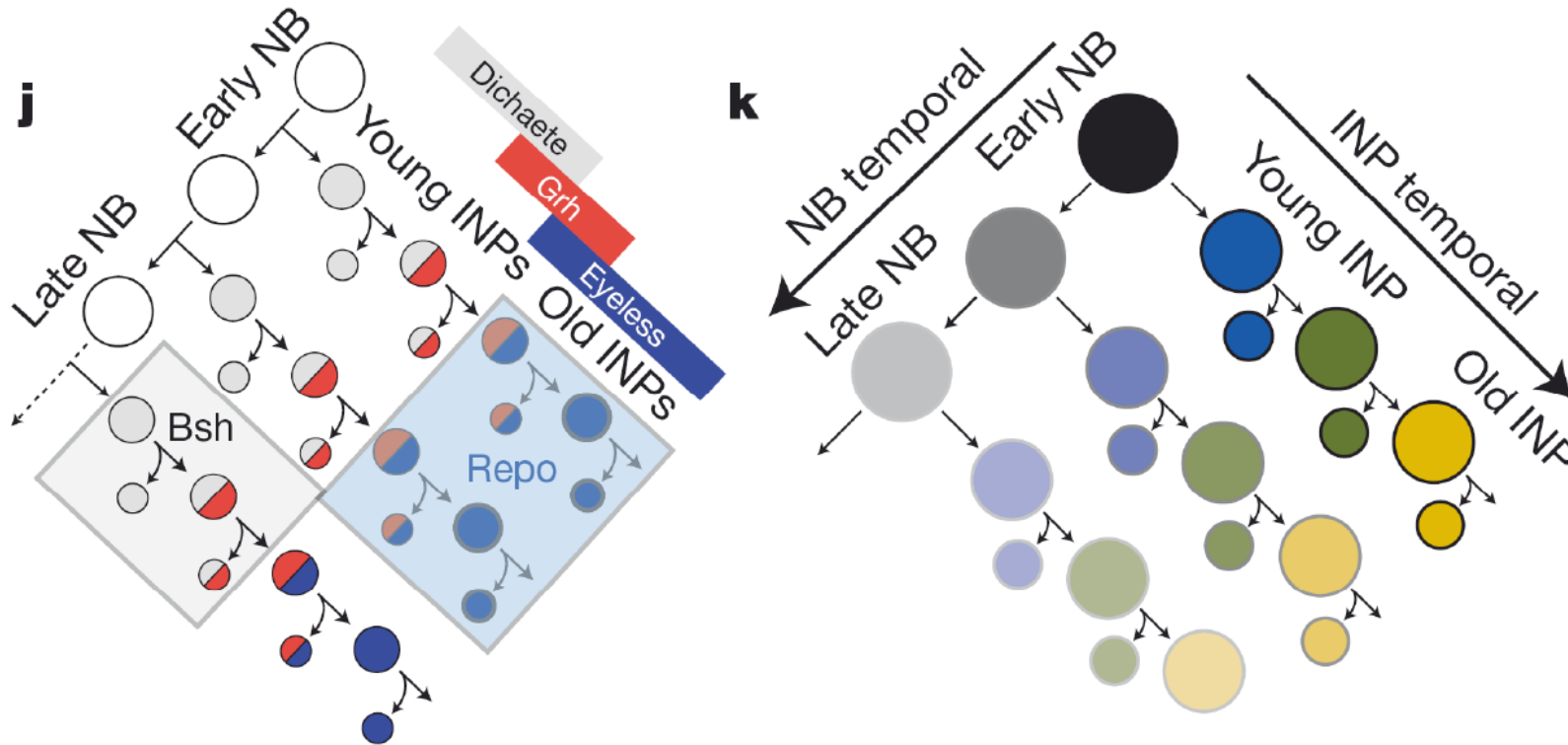
# Origin of neuroblast diversity

Temporal control of neuroblast identity



These TFs do not specify a certain neuron type, but control the birth-order-dependent neuronal identity

# Adult Neurogenesis



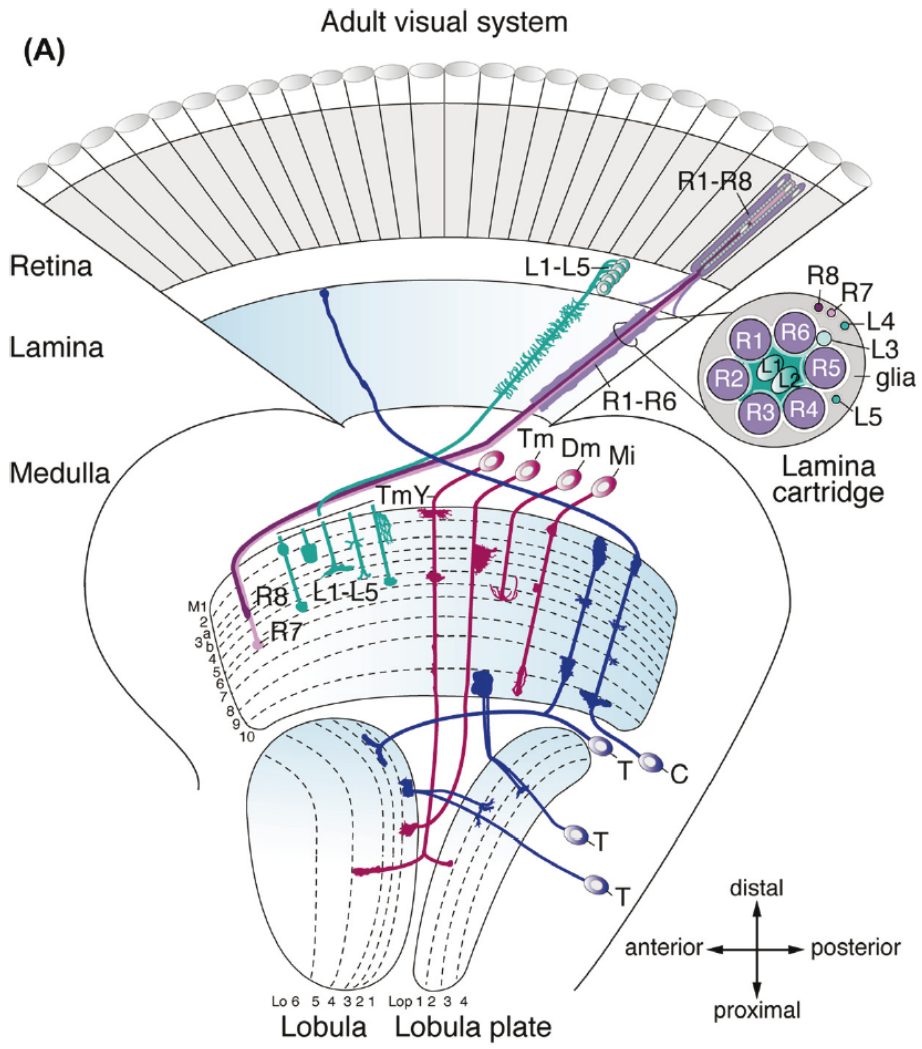
Distinct neural progeny are born from early versus late type II neuroblast lineages  
neuroblast and Intermedial Neural Progenitors temporal patterning act together to  
generate neural diversity

TF sequences on the two temporal axes act combinatorially to  
generate larger neural diversity

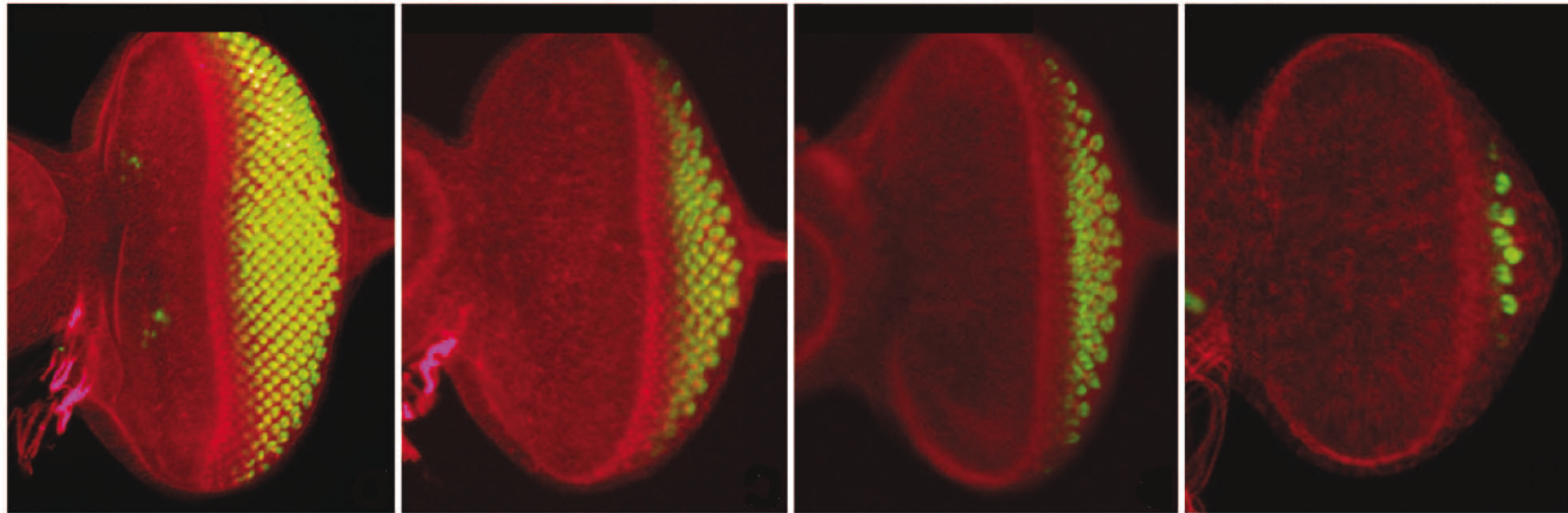
# **Glia relay differentiation cues to coordinate neuronal development in *Drosophila***

**Vilaiwan M. Fernandes,<sup>\*†</sup> Zhenqing Chen,<sup>†‡</sup> Anthony M. Rossi,  
Jaqueline Zipfel,<sup>§</sup> Claude Desplan**

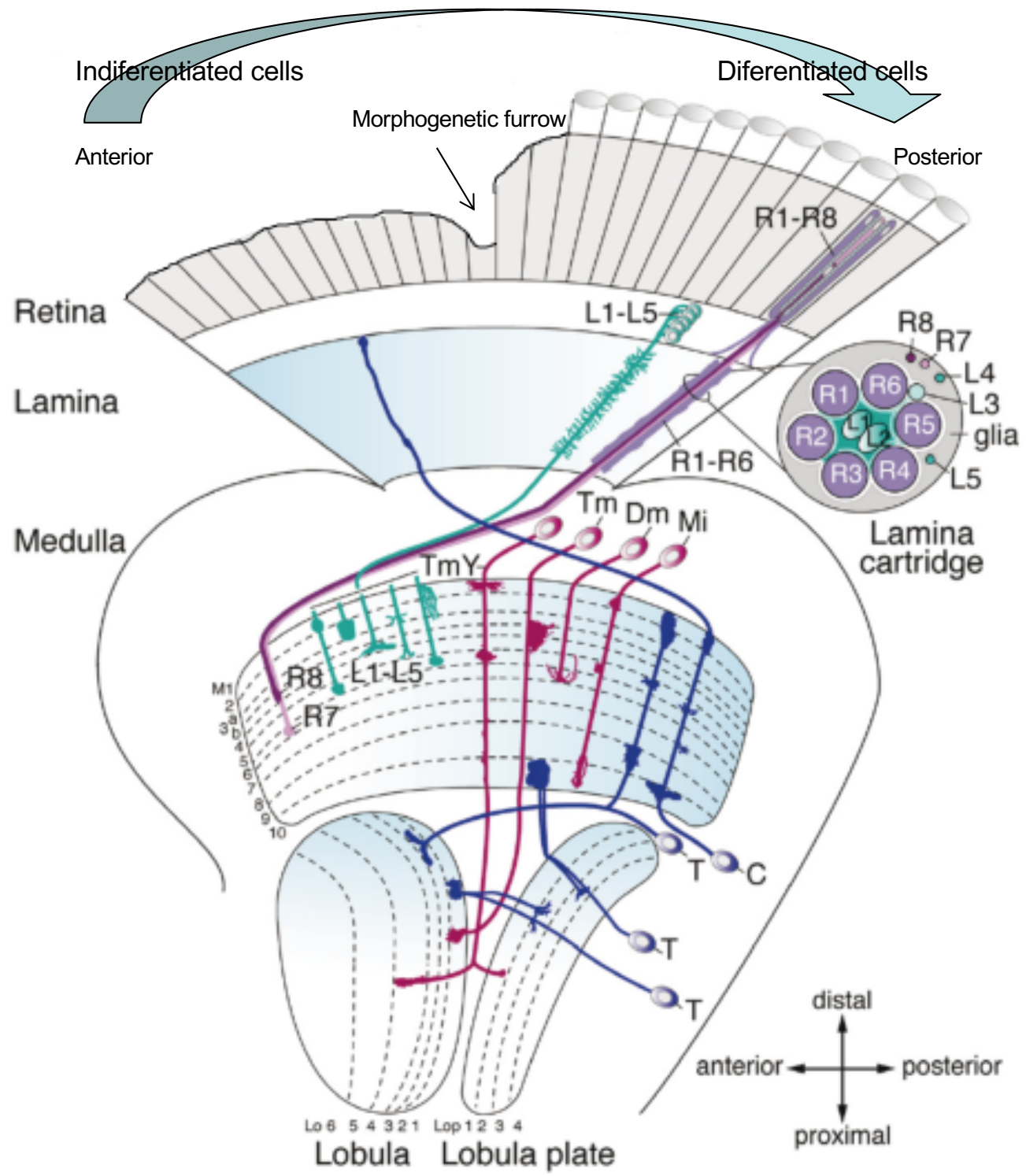
, *Science* **357**, 886–891 (2017) 1 September 2017



Time



Progression of the morphogenetic furrow across the developing eye field of *Drosophila*



Indifferentiated cells

Differentiated cells

Anterior

Morphogenetic furrow

Posterior

Retina

Lamina

Medulla

R1-R8

L1-L5

R8 R7

L4 L3 L5

glia

R1-R6

Lamina cartridge

Tm Dm Mi

M1

R8 L1-L5 R7

T C

T

T

distal

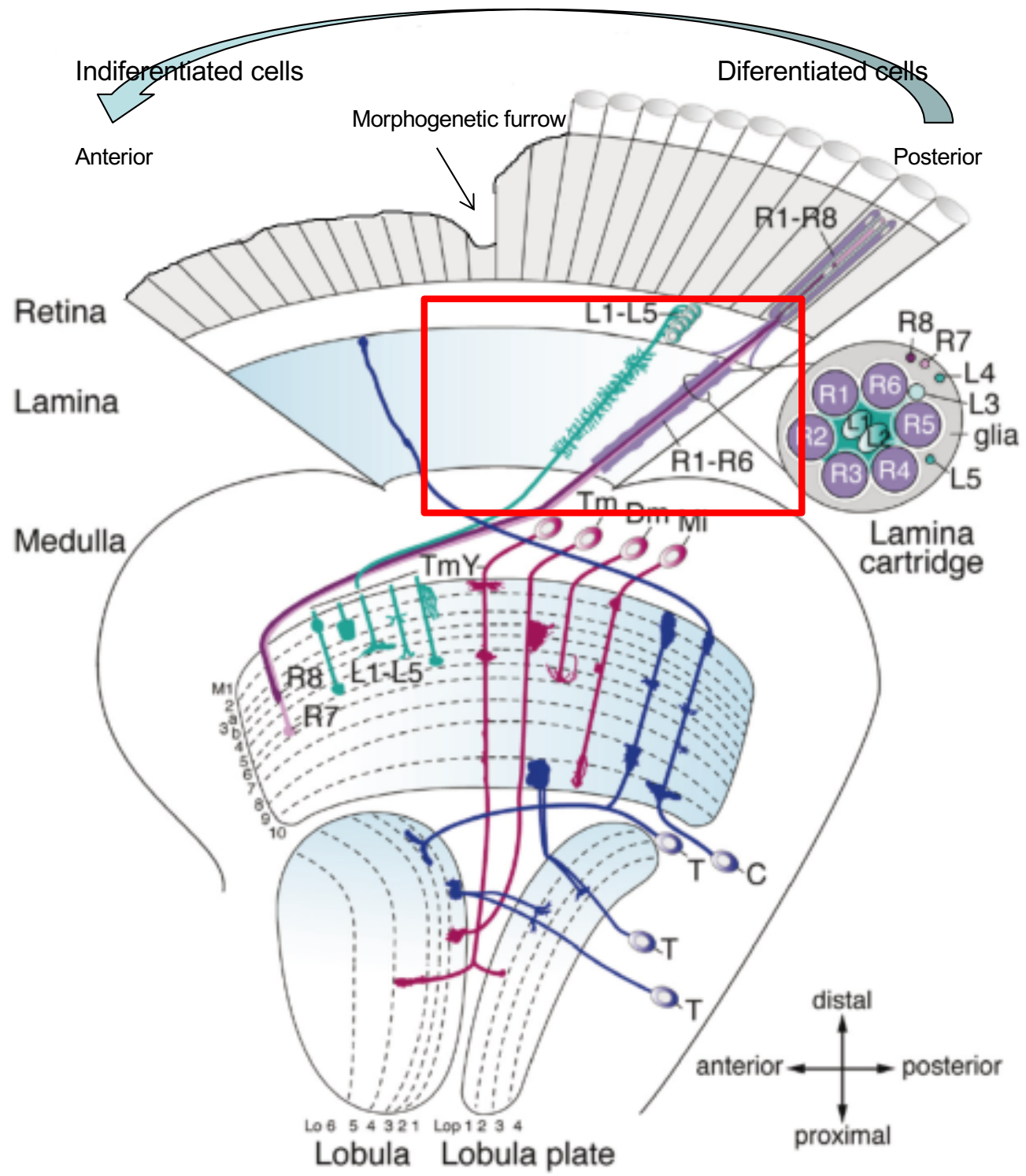
anterior

posterior

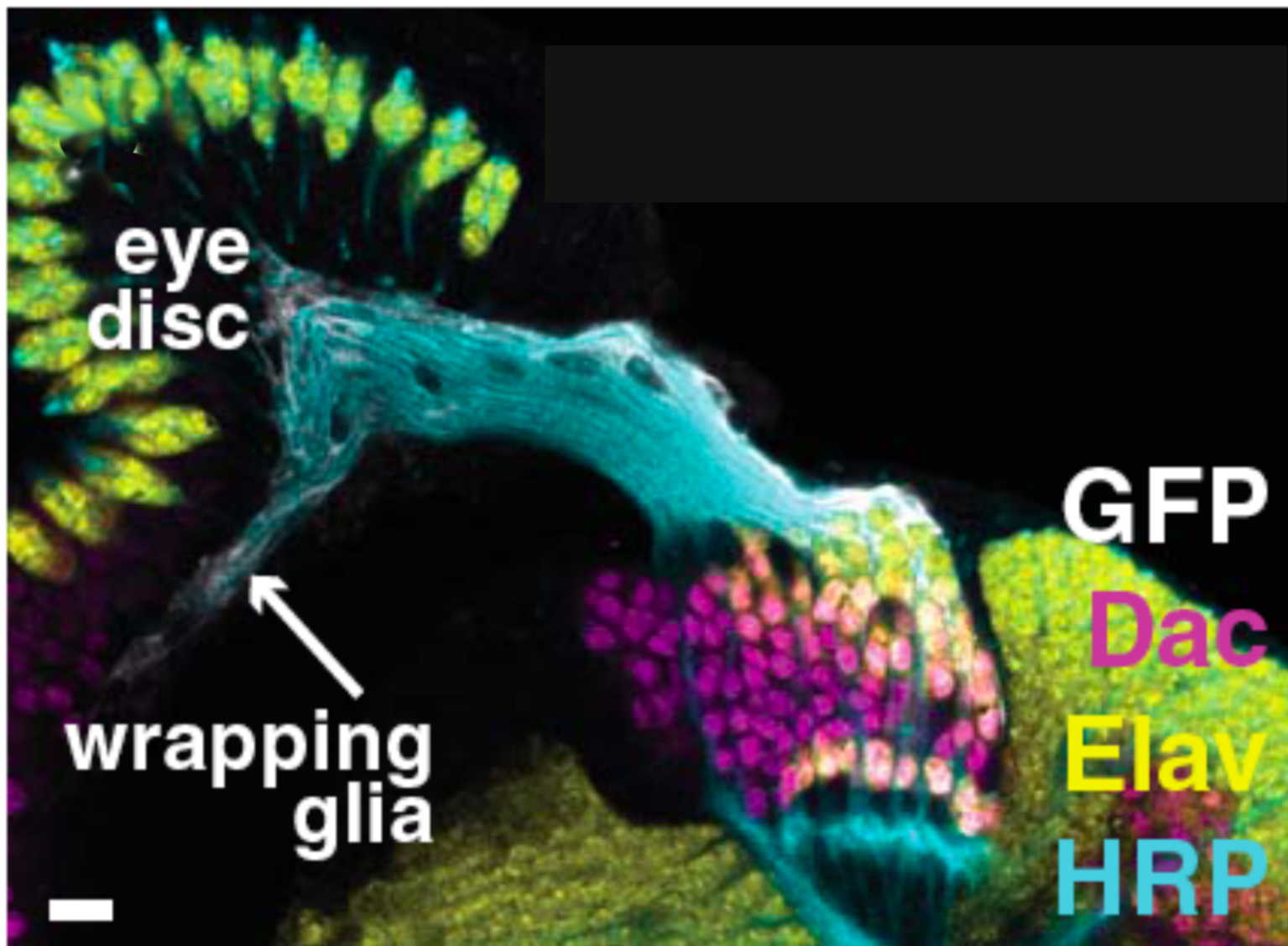
proximal

Lo 6 5 4 3 2 1 Lop 12 3 4

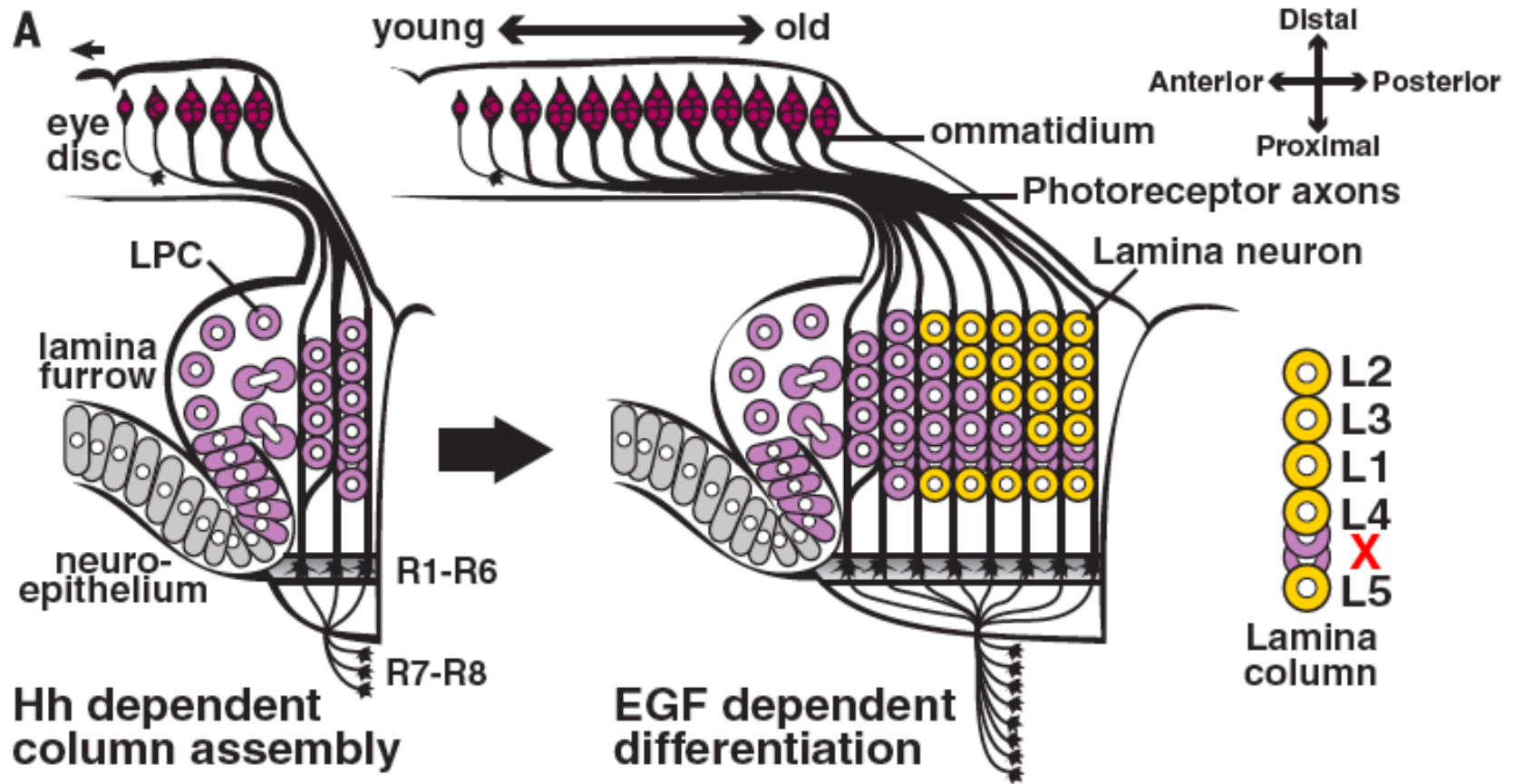
Lobula Lobula plate





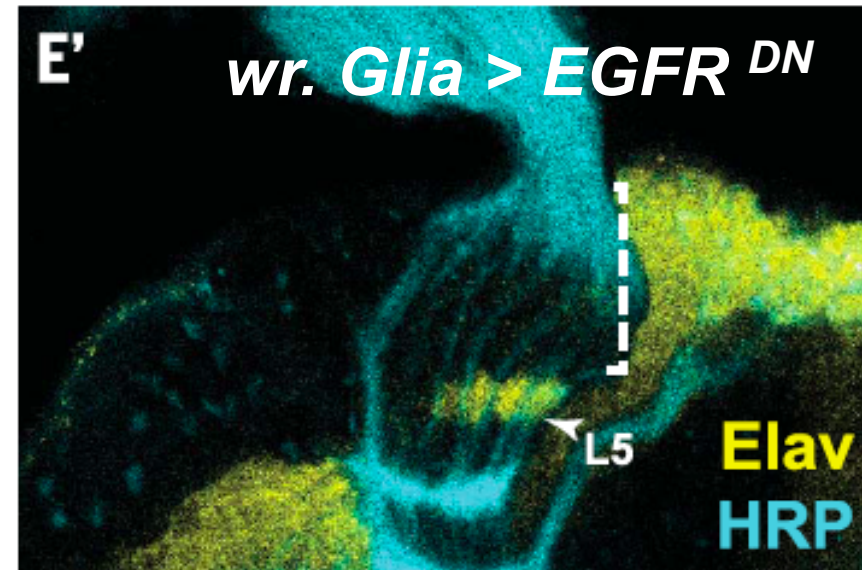
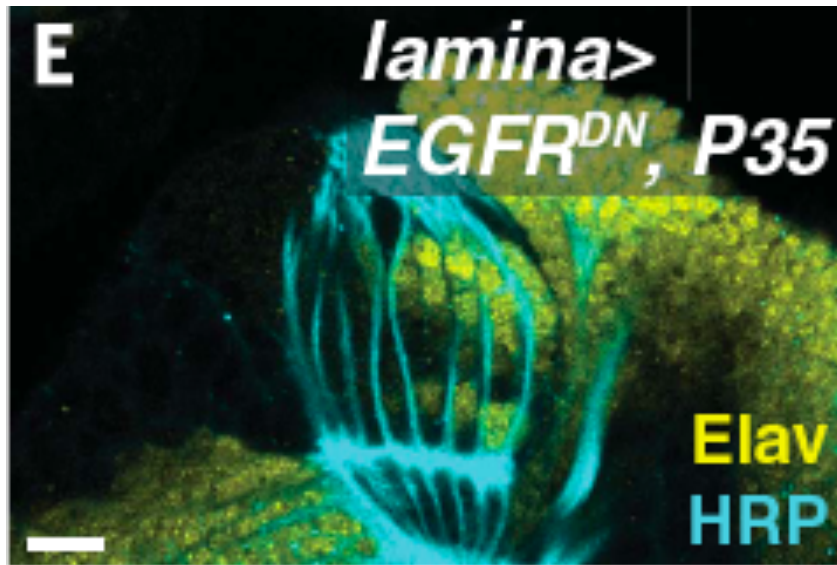




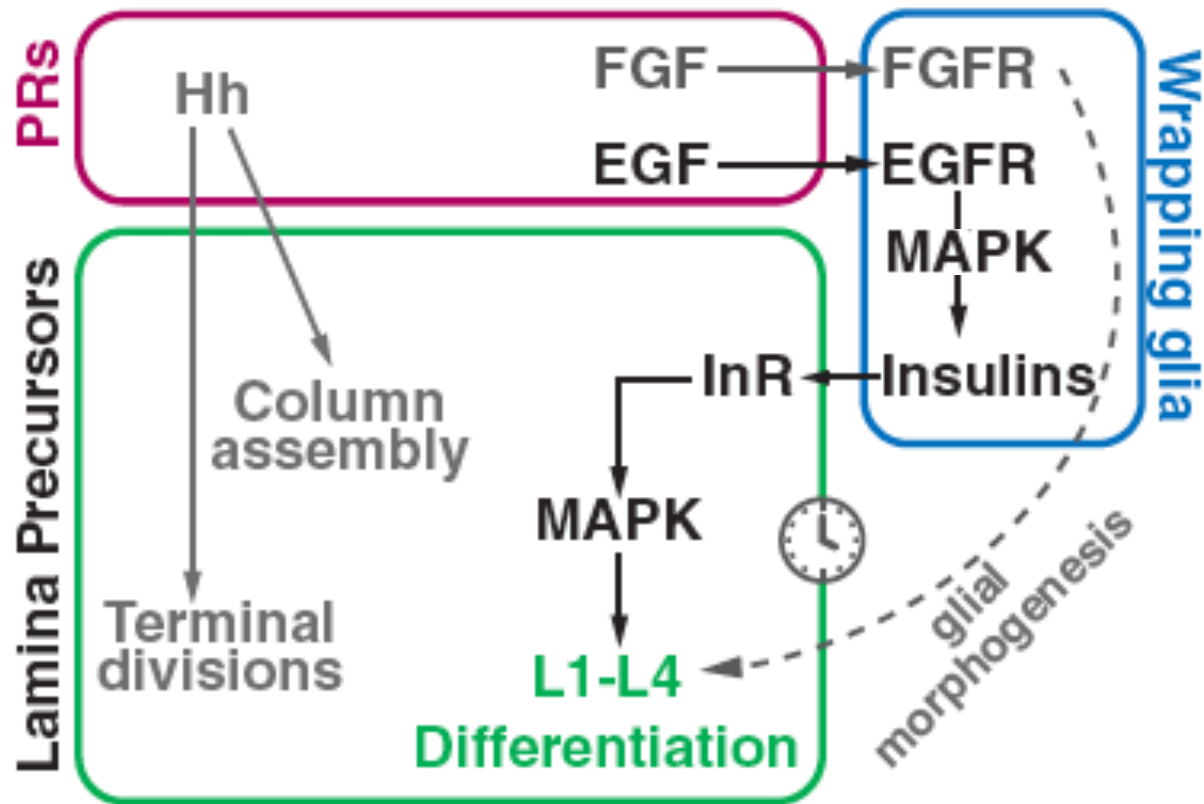


Signals from photoreceptors axons control lamina precursor cells cell divisions, column assembly and differentiation

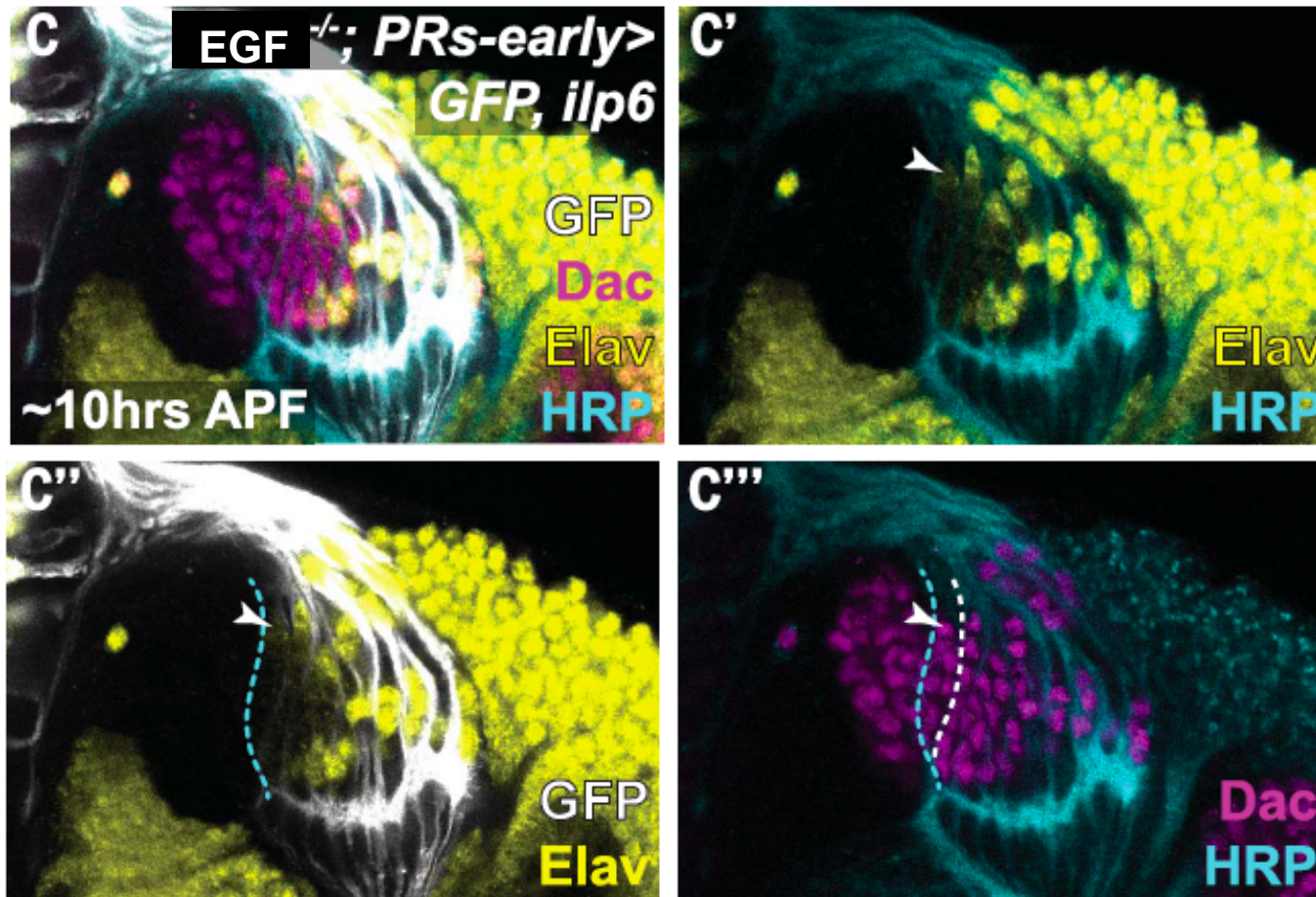
However, precursors differentiate only after column assembly is completed.



EGFR in wrapping glia, but not in lamina precursor cells, is required for lamina cells differentiation into neurons.



The connection between from photoreceptors to wrapping glia to lamina precursors induced a delay in the MAPK signalling that results in the assembly of lamina cells in columns followed by they differentiation



Activation of MPAK signalling in lamina precursor cells directly by photoreceptor axons disorganises column assembly.

The connection between from photoreceptors to wrapping glia to lamina precursors induced a delay in the MAPK signalling that results in the assembly of lamina cells in columns followed by they differentiation

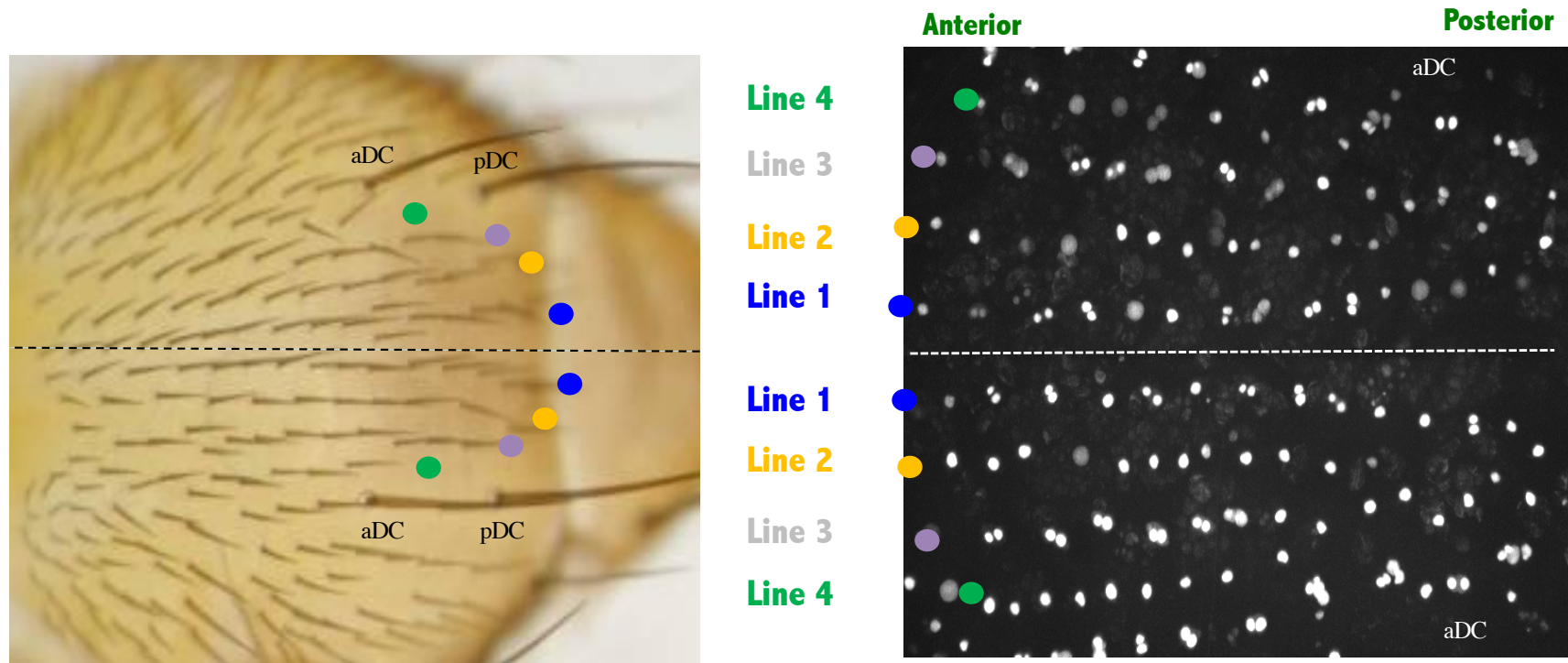
> [Elife](#). 2022 Mar 7;11:e75746. doi: 10.7554/eLife.75746.

## **A neural progenitor mitotic wave is required for asynchronous axon outgrowth and morphology**

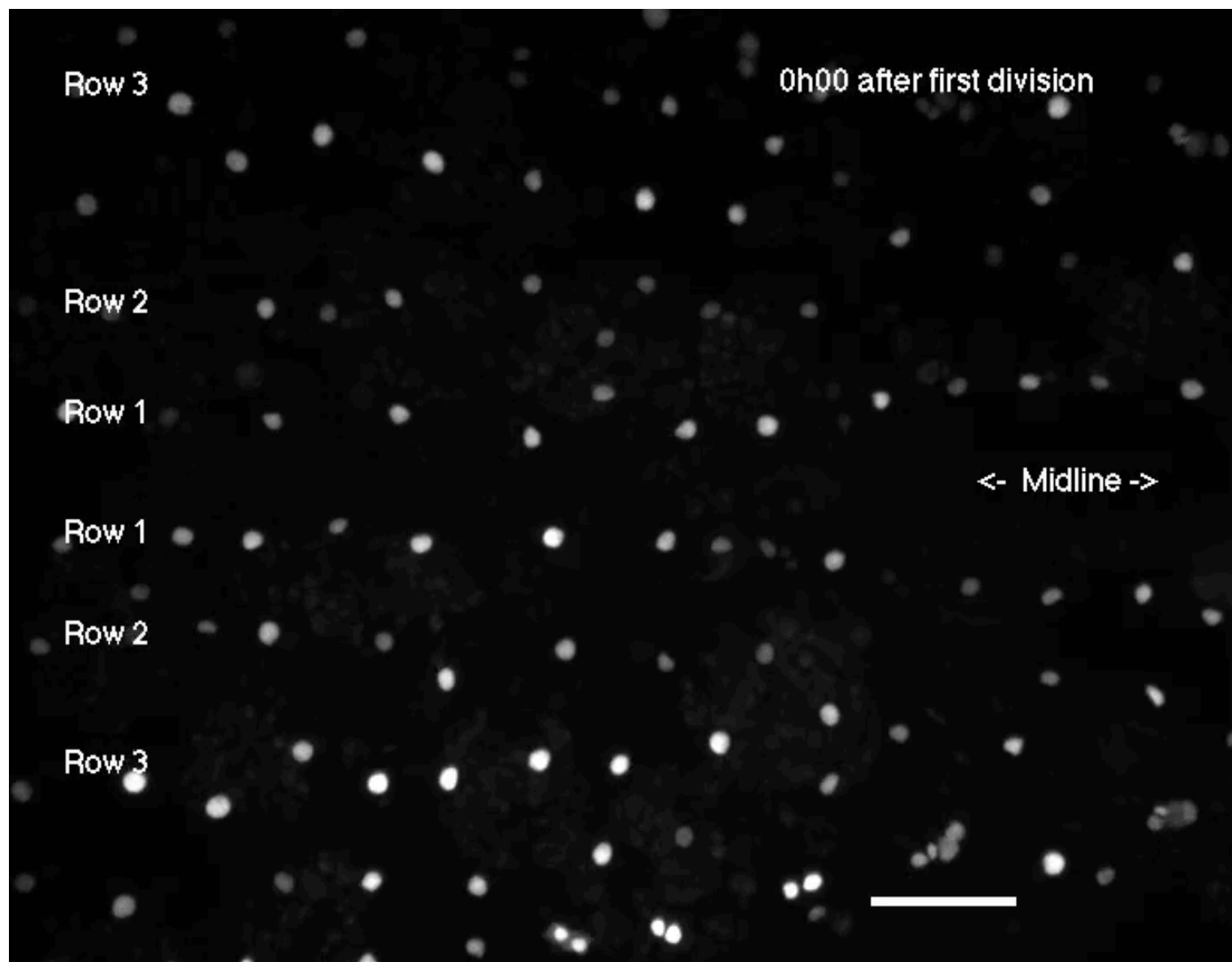
Jérôme Lacoste <sup>1</sup>, Hédi Soula <sup>2</sup>, Angélique Burg <sup>1</sup>, Agnès Audibert <sup>1</sup>, Pénélope Darnat <sup>1</sup>, Michel Gho <sup># 1</sup>, Sophie Louvet-Vallée <sup># 1</sup>

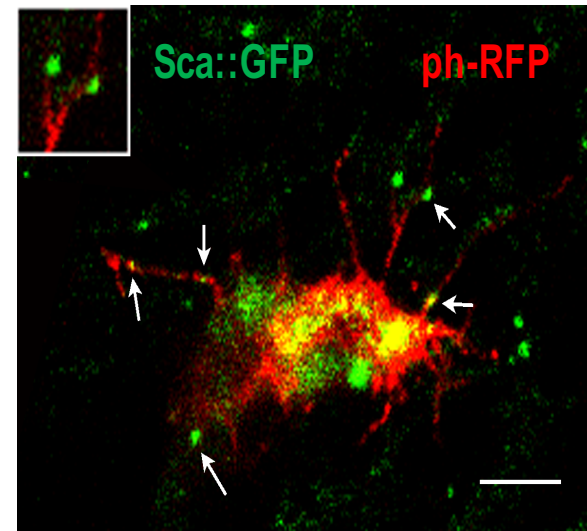
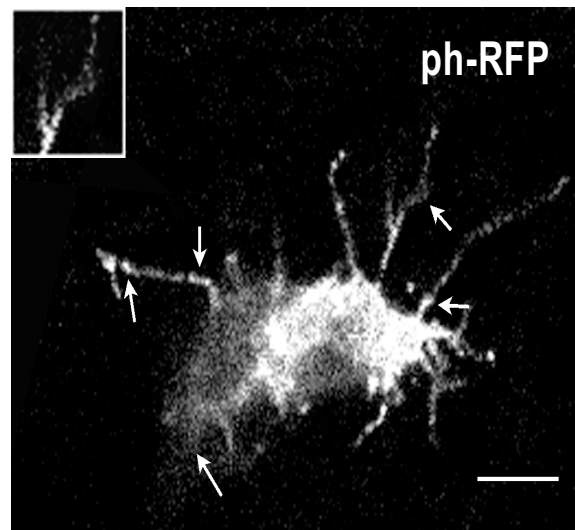
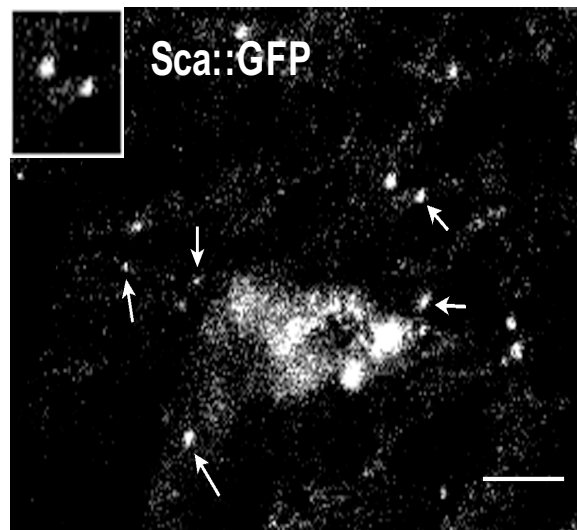
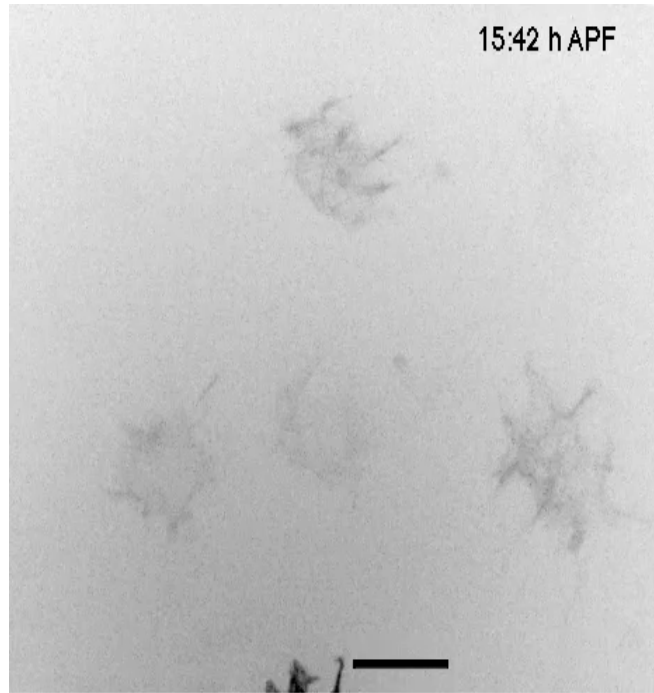


The Sensory Organ Precursor cells (SOP cells) producing the mechanosensory organs in *Drosophila* are among the few cases of normal G2-arrested somatic cells.

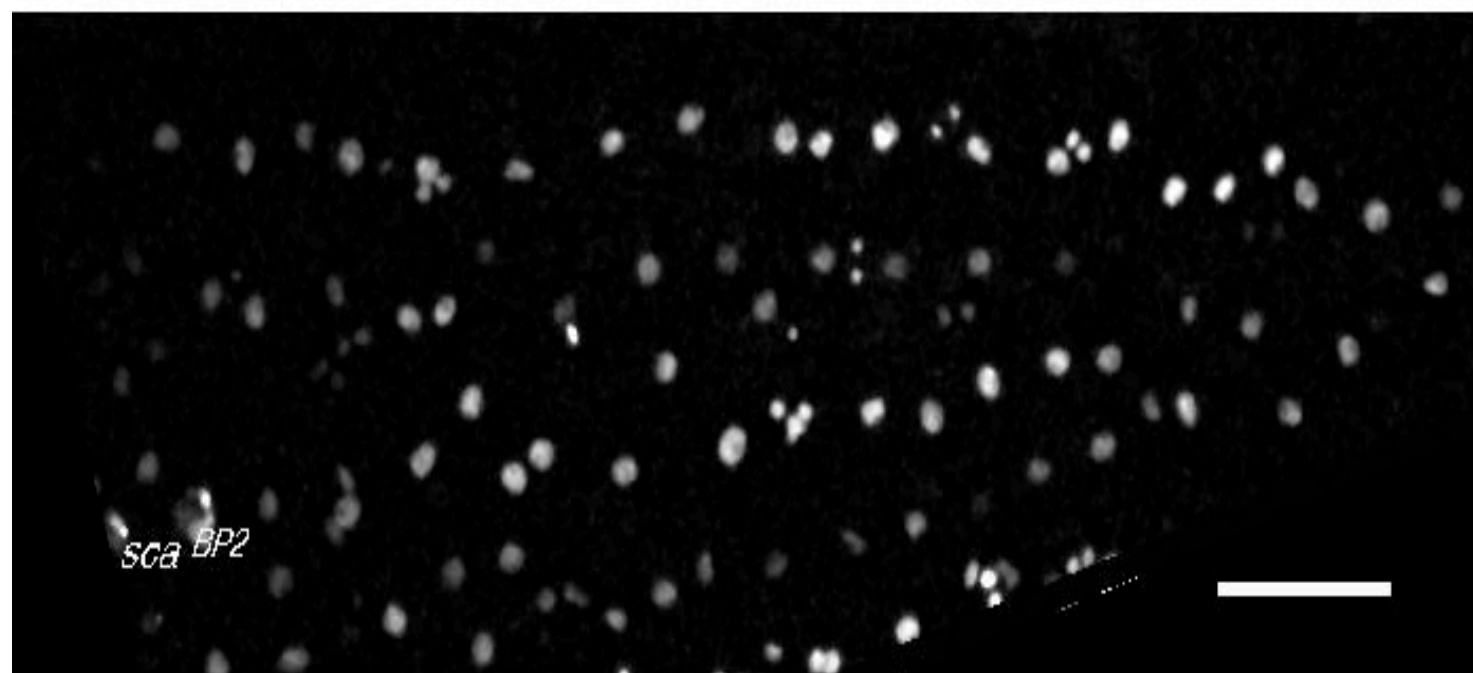
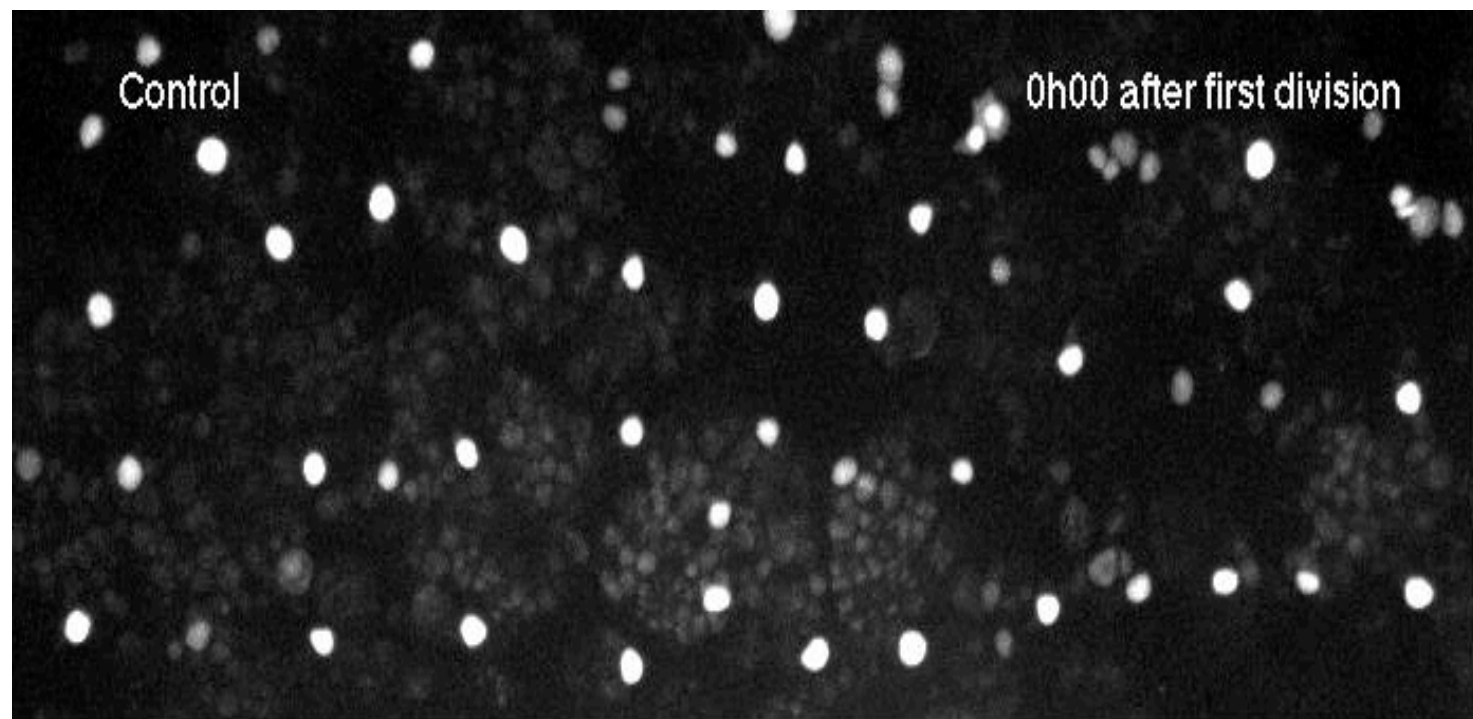


What is the dynamics of the resumption of the G2-arrest in these cells?



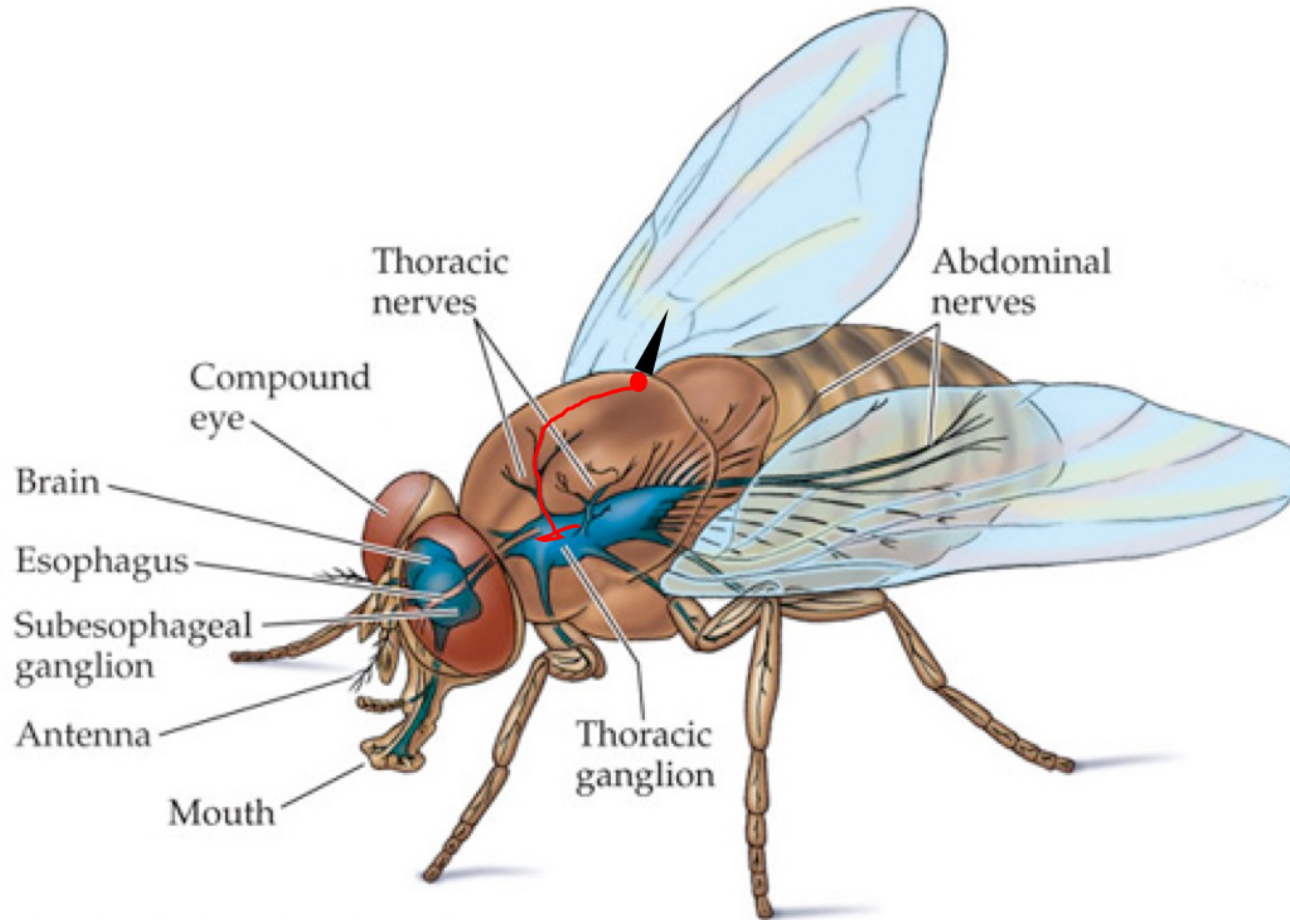






## The mitotic wave: physiological consequences

### Morphometry of axonal projections in the thoracic ganglion



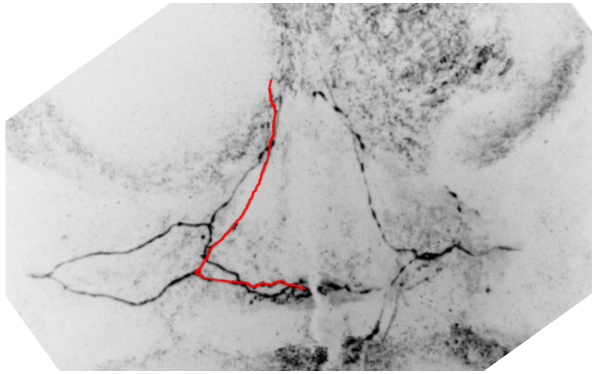
## The mitotic wave: physiological consequences

### Morphometry of axonal projections in the thoracic ganglion

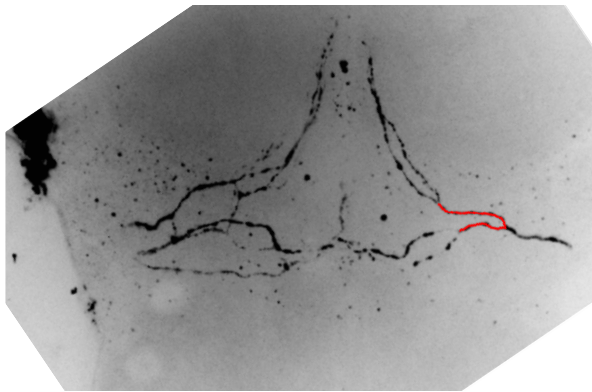
➤ **Pnr Gal4 > UAS-PH::GFP**



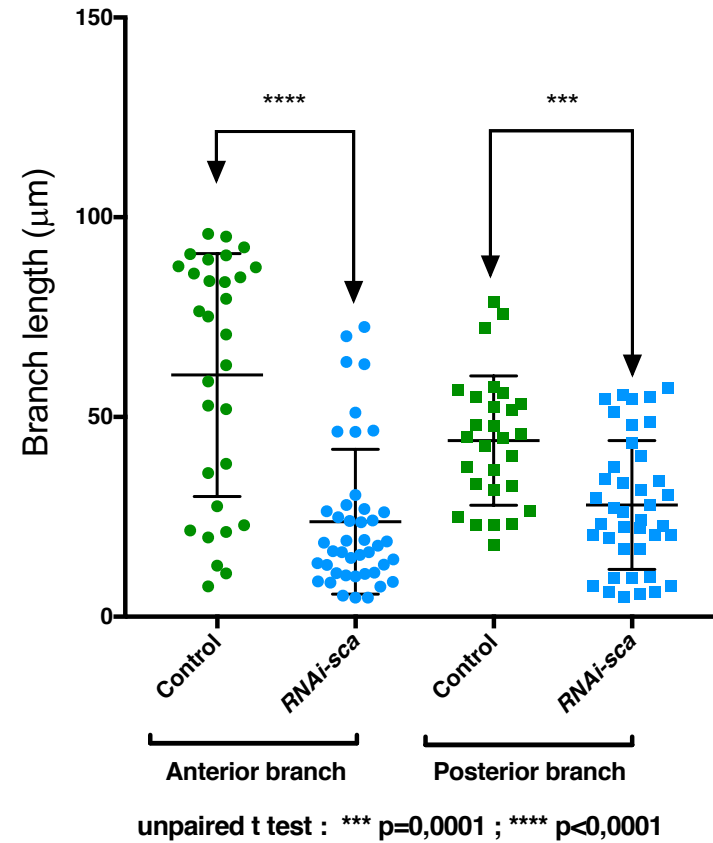
## The mitotic wave: physiological consequences



Control



*RNAi-sca*



When mitotic wave was specifically abolished, sensory organ axon terminals branched more than in control flies.



The cleaning reflex

# The mitotic wave: physiological consequences

## The cleaning reflex

**Control**



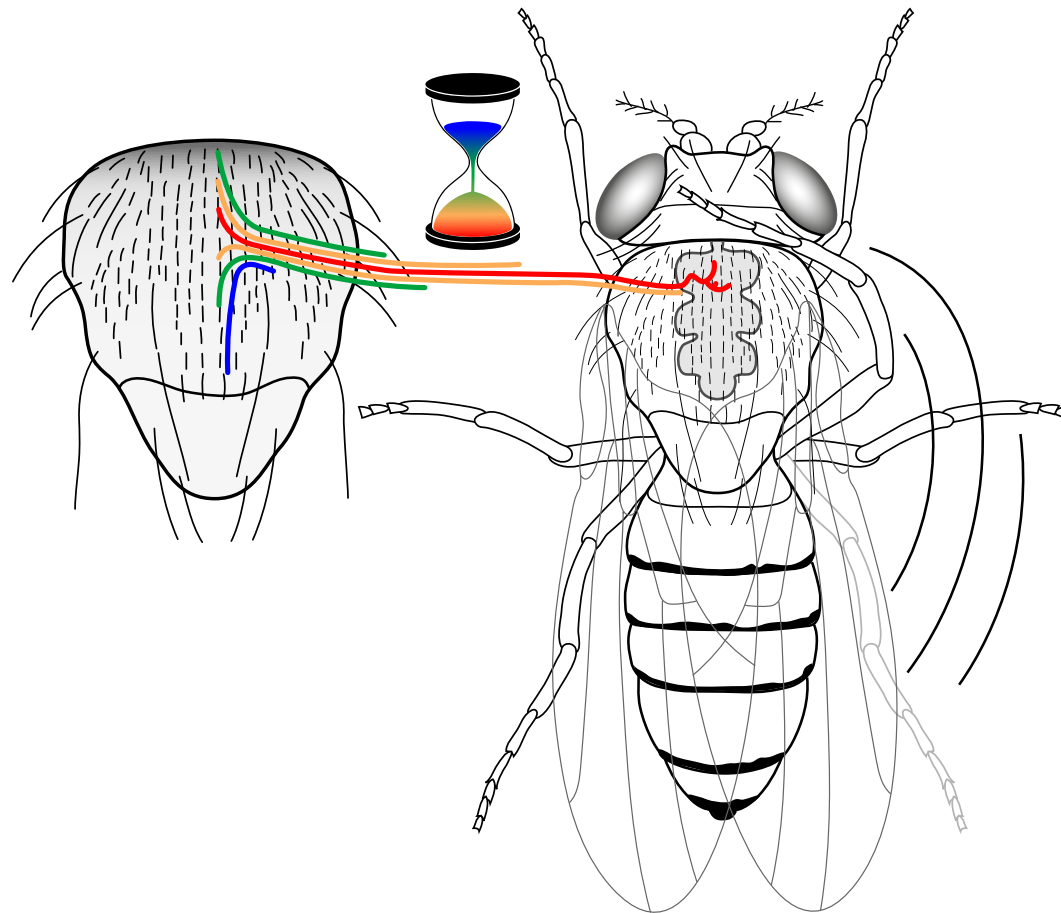
# The mitotic wave: physiological consequences

The cleaning reflex

When mitotic wave  
was abolished







The mitotic wave observed in precursor cells is related to progressive arrival of axon terminals into the central ganglion, and that this is required for a correct axon branching and appropriate behavior

In development, time is not only a dimension but also a parameter



